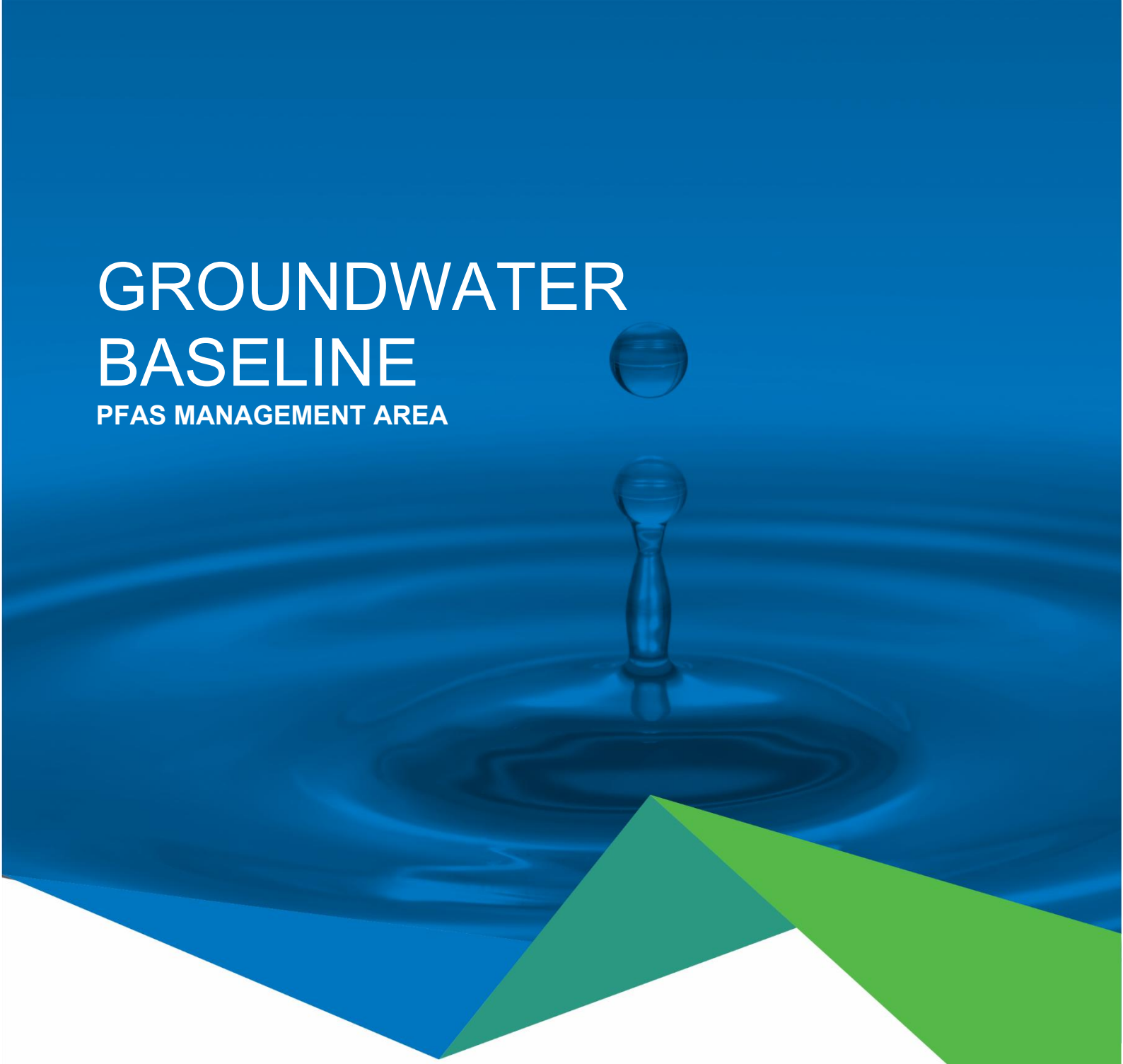
GROUNDWATER BASELINE

PFAS MANAGEMENT AREA



Western Sydney University
Richmond NSW

Hamilton Locke – February 2026

Geo-Logix
environment • geotech

DOCUMENT CONTROL

GROUNDWATER BASELINE

PFAS Management Area
Western Sydney University, Hawkesbury Campus
2 College Street
Richmond NSW

PREPARED FOR

Hamilton Locke

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EXECUTIVE SUMMARY

Geo-Logix Pty Ltd (Geo-Logix) was engaged by Hamilton Locke Pty Ltd, on behalf of Western Sydney University (WSU), to undertake a groundwater baseline assessment for the PFAS Management Area (PMA) at the Hawkesbury Campus, Richmond NSW. The assessment establishes baseline PFAS groundwater conditions as of October 2025 to inform ongoing PFAS management, risk assessment, and future decision-making within the PMA.

Groundwater samples were collected from twenty-five (25) monitoring wells comprising nineteen (19) shallow wells targeting a discontinuous perched groundwater system and six (6) deep wells screened within the regional groundwater system. Sampling and analysis were undertaken for PFAS by a NATA-accredited laboratory, with quality assurance and quality control procedures implemented in accordance with relevant Australian Standards and industry guidance.

The hydrogeological assessment confirms the presence of two distinct groundwater systems beneath the PMA: a shallow, localised perched system within sandier lenses of the Londonderry Clay, and a deeper, regionally continuous groundwater system characterised by low hydraulic gradients and very slow groundwater velocities. Shallow groundwater was found to be hydraulically discontinuous, exhibiting significant spatial variability, while the deeper groundwater system displayed consistent elevations and flow directions broadly aligned with historical investigations.

Analytical results indicate that PFAS impacts are largely confined to shallow groundwater and are spatially localised. Perfluorooctanesulfonic acid (PFOS) was detected above the applicable freshwater guideline value in a limited number of shallow wells, with the highest concentrations recorded adjacent to the student accommodation area. Perfluorooctanoic acid (PFOA) was not detected above guideline values in any groundwater samples. Deep groundwater impacts were limited, with PFOS exceeding guideline values in only one well, located adjacent to an area of known shallow groundwater impact. These findings are consistent with previous investigations and support the conceptual model of a localised PFAS source associated with historical fill placement.

Overall, the baseline assessment provides a robust snapshot of groundwater conditions within the PMA as of October 2025. To support any future regulatory, risk-based, or management decisions, it is recommended that bi-annual groundwater monitoring be undertaken to etablish temporal variability and PFAS concentration trends.

TABLE OF CONTENTS

1. INTRODUCTION 1
1.1 Objective 1
2. PFAS MANAGEMENT AREA 1
3. HYDROGEOLOGY 1
4. SITE ASSESSMENT CRITERIA 3
5. BASELINE SCOPE 3
5.1 Groundwater Monitoring Well Selection 3
5.2 Scope of works 3
5.3 Quality Assurance 4
6. INVESTIGATION RESULTS 5
6.1 Well Gauging Data 5
6.2 Field Parameter Data 6
6.3 Groundwater Analytical Results 7
7. CONCLUSIONS 10
8. RECOMMENDATIONS 10
9. REFERENCES 12

FIGURES

Figure 1: Site Location
Figure 2: Site Map
Figure 3: Shallow Groundwater Monitoring Locations
Figure 3a: Groundwater Elevation Map – Shallow Groundwater
Figure 4: Deep Groundwater Monitoring Locations
Figure 5: Groundwater Elevation Map – Deep Groundwater
Figure 6: PFOS SAC Exceedances – Shallow Groundwater
Figure 7: Total PFAS Concentration – Shallow Groundwater
Figure 8: PFOS SAC Exceedances – Deep Groundwater
Figure 9: Total PFAS Concentration – Deep Groundwater

TABLES

Table 1: Summary of Shallow Groundwater PFAS Analytical Data

Table 2: Summary of Deep Groundwater PFAS Analytical Data

Table 3: Summary of QA/QC

ATTACHMENTS

Attachment A: Groundwater Sampling Logs

Attachment B: Equipment Calibration Certificates

Attachment C: Monitoring Well Survey Data – Additional Works

Attachment D: Laboratory Reports

1. INTRODUCTION

Geo-Logix Pty Ltd (Geo-Logix) was engaged by Hamilton Locke Pty Ltd (Client Agent) on behalf of Western Sydney University (WSU) to undertake the 2025 groundwater baseline assessment for the PFAS Management Area (PMA), WSU Hawkesbury Campus (Figures 1 & 2). The baseline assessment included the collection of groundwater samples from twenty-five (25) monitoring wells, with laboratory analysis for per- and polyfluoroalkyl substances (PFAS).

1.1 Objective

Establish the concentrations of PFAS in both the shallow and deep groundwater systems underlying the PMA as at October 2025. These results will define the baseline groundwater conditions for the PMA against which future monitoring and management performance can be assessed.

2. PFAS MANAGEMENT AREA

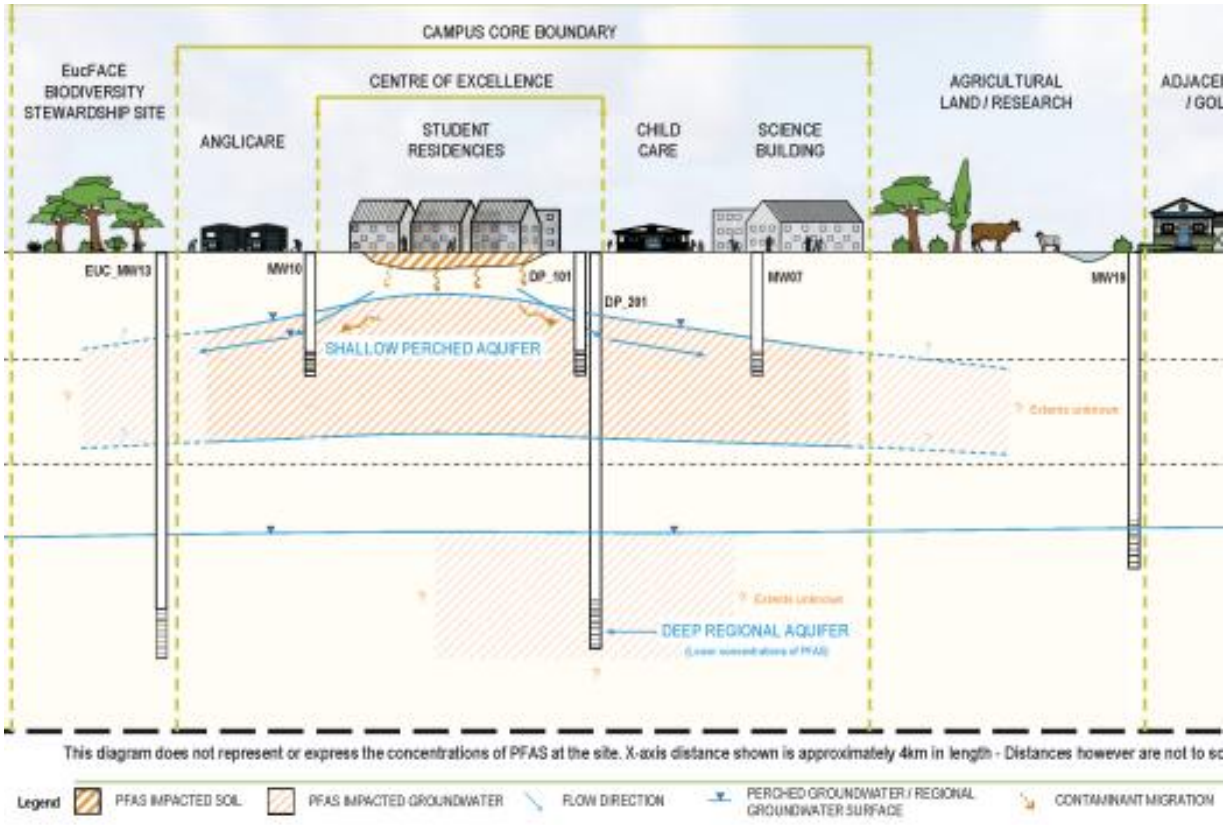
PFAS was first identified at the Hawkesbury Campus in 2021 during groundwater sampling for the former Centre of Excellence (CoE) development. Subsequent investigations confirmed that PFAS impacts are highly localised within the central campus core, specifically beneath and adjacent to the former CoE development footprint and the student residences. Multiple lines of evidence indicate the PFAS source is fill of unknown origin placed during 2011–2012.

The PMA, as shown in Figure 1, is approximately 15.5 hectares and encompasses the portion of the campus where PFAS impacts have been historically detected. It includes the student accommodation precinct, the former centre of excellence (CoE) development area, and the interconnected east and west drainage channels, which represent the principal surface water migration pathways. The boundary captures the grassed open areas between these features, where perched groundwater has been encountered and interacts with stormwater infrastructure. The management area forms an irregular polygon extending from the residences in the north to the confluence of drainage channels in the south. This area represents the zone of interest for ongoing PFAS management. An interim Long Term Management Plan for the area is currently being prepared.

3. HYDROGEOLOGY

Two hydrogeological units exist beneath the PFAS Management Area. The first is a shallow, discontinuous perched groundwater system that occurs within localised sandier lenses which pinch and swell within the Londonderry Clay, and is likely recharged by irrigation across the student grounds. The second is a deeper regional groundwater system forming a continuous saturated zone within the Londonderry Clay. The conceptual model and hydrogeological unit characteristics are summarised below.

CONCEPTUAL MODEL



Source – Aurecon 2022.

Table 1: Shallow Groundwater

PERCHED GROUNDWATER SYSTEM	
Discontinuous zone of saturation 0.9 – 4mbg within sandier clay lenses of the Londonderry Clay.	
Water Quality Characteristics	
Electrical Conductivity	Fresh to brackish
Hydrogeological Characteristics	
Groundwater Gradient	0.0175 – 0.023 (Aurecon 2023)
Flow Direction	Radial - Groundwater elevation map Figure 3a.

Table 2: Deep Groundwater

REGIONAL GROUNDWATER SYSTEM	
The primary aquifer for solute transport to surface water receptors. A zone of continuous saturation within the Londonderry Clay encountered between 7 – 15 mbg.	
Water Quality Characteristics	
Electrical Conductivity	Brackish
Hydrogeological Characteristics	
Groundwater Flow Direction	Southeast beneath the PMA – Figure 5.
Groundwater Gradient	0.0008
Hydraulic Conductivity	Hydrogeological tests performed by Aurecon (2023) indicate an average K value of 0.0295 m/day
Seepage Velocity	Assuming an effective porosity of 33% and gradient of 0.0006, Aurecon estimates the groundwater flow rate to be 5.36 x 10^{-5} m/day (equivalent to 2 cm movement per year).

4. SITE ASSESSMENT CRITERIA (SAC)

- Perfluorooctanesulfonic acid (PFOS) – 0.13 ug/L
- Perfluorooctanoic acid (PFOA) – 220 ug/L

Australian and New Zealand Guidelines for Fresh and Marine Water Quality – interim freshwater default guideline values (SAC) for PFOS and PFOA (ANZG 2023), 95% species protection.

5. BASELINE SCOPE

5.1 Groundwater Monitoring Well Selection

The majority of PFAS impact exists within shallow groundwater. Accordingly, the selection of monitoring wells was designed to characterise both hydrogeological units while providing adequate spatial coverage across the PMA. A total of nineteen (19) shallow wells (Figure 3) and six (6) deep groundwater monitoring wells (Figure 4) were selected for the baseline. Shallow wells were selected along six defined east – west transects, allowing assessment of PFAS distribution and groundwater flow conditions within the perched/shallow zone. The deep wells were selected around the footprint of the baseline assessment area.

5.2 Scope of works

The scope of works for assessment included:

- Review of historical groundwater assessment reports.
- Site walkover to assess sample locations and logistics.
- Groundwater level gauging: All selected wells were opened and equilibrated prior to measurement. Groundwater level, total well depths and casing stick-ups were recorded. Gauging was completed on the same day.

- Groundwater sampling from deep groundwater wells via Hydrasleeves and measurement of physiochemical parameters.
- Groundwater sampling from shallow groundwater wells via Low Flow sampling technique and measurement of physiochemical parameters.
- Collection of a sample from each well in a 500mL PET/HDPE bottle, labelled, placed on ice in an esky and transported under chain of custody to a NATA Accredited Laboratory (Eurofins) for PFAS analysis.
- Sample analysis for PFAS by a NATA-accredited laboratory.
- Provision of this Groundwater Baseline Assessment Report.

Groundwater sample logs are presented in Attachment A. Equipment calibration certificates are presented in Attachment B.

5.3 Quality Assurance

Quality control (QC) sampling was undertaken in general accordance with specifications outlined in AS4482.1, Guide to Sampling and Investigation of Potentially Contaminated Soil. Field QC samples included:

- Inter and Intra Laboratory Duplicates
- Field Blanks
- Equipment Rinsates

Sample Identification	Date	Sample Type	Sample Matrix	Rate of Collection
DW1	21/10/2025	Field duplicate of BH108	Water	1 in 5 samples
TW1	21/10/2025	Field triplicate of BH108	Water	1 in 5 samples
DW2	23/10/2025	Field duplicate of GW128	Water	1 in 5 samples
TW2	23/10/2025	Field triplicate of GW128	Water	1 in 5 samples
DW3	23/10/2025	Field duplicate of BH107	Water	1 in 5 samples
TW3	23/10/2025	Field triplicate of BH107	Water	1 in 5 samples
DW4	28/10/2025	Field duplicate of BH111	Water	1 in 5 samples
TW4	28/10/2025	Field triplicate of BH111	Water	1 in 5 samples
R1	21/10/2025	Equipment Rinsate	Water	1 per day of sampling
R2	23/10/2025	Equipment Rinsate	Water	1 per day of sampling
R2	28/10/2025	Equipment Rinsate	Water	1 per day of sampling
BLANK1	21/10/2025	Field Reagent Blank	Water	1 per batch
BLANK2	23/10/2025	Field Reagent Blank	Water	1 per batch
BLANK3	28/10/2025	Field Reagent Blank	Water	1 per batch

Field quality assurance (QA) procedures included:

- Changing disposable gloves between samples.

- Between each sampling location, the groundwater interface probe was decontaminated by rinsing it in a Liquinox® (PFAS-free) detergent solution, followed by two sequential rinses using laboratory-prepared PFAS-free deionised water.
- Use of PFAS “safe” products including HDPE HydraSleeves, HDPE sample tubing, PVC bailers and polypropylene bailer cord.
- Bottles were labelled after sample collection.

6. INVESTIGATION RESULTS

6.1 Well Gauging Data

Aquifer	Well ID	Well Cover	Stick Up (m)	Top of Casing (mAHD)	Total Depth of Well (mBTOC)	Standing Water Level 27/10/2025 (mBTOC)	Groundwater Elevation 27/10/2025 (mAHD)
Shallow	MW03_s	Gatic	0	23.440	5.912	1.708	21.732
	MW10	Gatic	0	24.091	7.014	6.761	17.33
	MW02	Gatic	0	23.879	1.299	2.300	21.579
	BH101	Monument	1.075	24.662	5.917	2.379	22.283
	MW11[1]	No cover	1.07	24.382	6.952	2.680	21.702
	GW2	Well could not be located.					
	BH103	Monument	1.05	24.053	5.506	Dry	-
	BH108	Monument	0.655	23.880	4.275	2.976	20.904
	BH107	Monument	0.99	24.237	4.941	3.164	21.073
	BH101h	Monument	0.94	24.531	5.377	1.971	22.56
	BH101i[2]	Gatic	0	23.056	1.430	0.964	22.092
	BH109	Monument	0.96	24.248	4.635	1.872	22.376
	BH105	Monument	0.98	23.598	4.715	2.015	21.583
	BH101g[3]	Monument	0.96	23.886	5.505	*	-
	BH106	Monument	0.97	23.805	5.120	3.062	20.743
	BH111	Gatic	0	23.191	4.986	0.979	22.212
	BH102_s	Monument	0.90	24.313	5.219	2.069	22.244
	BH101e	Gatic	Well cover was broken, preventing safe opening and access for measurement and sampling. Repair was attempted; however, it was not possible because the well casing could not be located.				
	BH104	Well could not be located.					
Deep	GW128	Gatic	0	22.702	14.845	12.321	10.381
	GW129	Gatic	0	22.632	14.328	12.305	10.327

Aquifer	Well ID	Well Cover	Stick Up (m)	Top of Casing (mAHD)	Total Depth of Well (mBTOC)	Standing Water Level 27/10/2025 (mBTOC)	Groundwater Elevation 27/10/2025 (mAHD)
	GW153	Monument	0.73	23.911	17.028	13.485	10.426
	GW155	Monument	0.75	23.502	15.602	13.348	10.154
	MW03_d	Gatic	0	23.4465	14.879	12.586	10.860
	BH201	Monument	0.62	24.175	20.630	13.507	10.668

[1] Well not consistent with historical records. The well identified as MW11 does not correspond to the previously mapped MW11 location.
[2] A rigid base was encountered during measurement with the interface probe, although the well contained water and showed no evidence of cementing.
[3] Well could not be measured on 27 October 2025, as plastic tubing from a previous sampling event was lodged too deeply inside the casing, preventing insertion of the interface probe. The tubing was removed the following day using additional tools, and the well was subsequently measured and sampled.

The well survey data collected during the additional works is provided in Attachment C.

Shallow Groundwater Observations

Gauging confirmed the presence of a shallow, discontinuous perched groundwater system across the PMA. Well BH103 was dry. Groundwater was encountered in multiple shallow wells; however, elevations show significant variability over relatively short distances, ranging from approximately 20.904 to 22.58 mAHD.

MW10 contained only a minimal volume of water, insufficient for sampling, and its screened interval intersects deeper materials rather than the shallow perched zone observed in the remaining wells. Accordingly, MW10 was not used in defining the interpolated shallow groundwater elevation.

Deep Groundwater Observations

Gauging indicates a continuous regional groundwater system, with most wells recording standing water levels between 10 and 11 mAHD. The regional groundwater flow direction is towards the south – southwest consistent with historical observations (Figure 5).

6.2 Field Parameter Data

Well	EC (us/cm)	pH	Redox (mV)	DO (mg/L)	Temp (°C)
Shallow Wells					
MW03_s	437.1	5.83	-117.3	7.96	17.5
MW10	903	5.54	-83.6	9.24	18.2
MW02	854	6.67	-151.4	0.61	17.6
BH101	1501	6.05	-139.9	1.20	18.0
MW11	583.1	5.77	-199.8	3.17	16.2
BH108	365	6.45	-156.5	0.33	18.2
BH107	278	5.95	278.0	0.53	17.0
BH101h	1071	6.35	-42.8	0.57	17.5

Well	EC (us/cm)	pH	Redox (mV)	DO (mg/L)	Temp (°C)
BH101i	2162	6.32	-195.6	1.36	19.6
BH109	783	6.58	-204.3	0.19	20.5
BH105	1280	6.87	-15.6	3.75	17.0
BH101g	899	6.75	-145.0	4.28	17.2
BH106	519	6.32	-66.9	0.38	21.0
BH111	1410	6.82	-176.2	1.69	18.8
BH102_s	518	6.54	-145.7	1.70	19.6
Deep Wells					
GW128	585	5.46	-47.1	5.04	20.7
GW129	4722	5.78	-254.7	4.65	20.1
GW153	9740	5.93	-76.8	1.48	21.5
GW155	7497	6.19	-94.5	5.14	20.6
MW03_d	598	5.65	-37.1	2.60	21.4
BH201	1094	6.18	-139.1	1.32	20.2

Note: Dissolved oxygen readings were not relied upon based on anomalous responses. DO meters are notoriously unreliable.

Key observations:

- Shallow groundwater is fresh, slightly acidic to near neutral. Redox values across the shallow wells predominantly range from –80 to –205 mV, indicating chemically reducing groundwater consistent with limited oxygen availability and poor hydraulic connectivity. One well (BH107) exhibited a strongly oxidising Eh (+278 mV), likely reflecting a localised recharge pathway or a thin oxidised sand lens. This variability supports the interpretation that shallow groundwater occurs as isolated perched zones with limited lateral continuity.
- Deep groundwater is primarily brackish and slightly acidic. Field redox potentials in the deeper groundwater system ranged from approximately –40 to –255 mV, indicating consistently non-oxidising to reducing conditions.

6.3 Groundwater Analytical Results

Groundwater analytical results are presented in Tables 1 to 3 and on Figures 6 to 9. Laboratory reports are presented in Attachment D.

Shallow Groundwater

- PFOS was detected above SAC in shallow groundwater samples BH101, BH101i, BH102_s and MW03_s.
- PFOA was not detected above SAC in any samples.

Total PFAS (sum of the 30 PFAS analytes) was detected at its highest concentration in shallow groundwater sample BH101 (580 µg/L), adjacent to the student accommodation area.

Deep Groundwater

- PFOS was detected above SAC in one deep groundwater sample (BH201), adjacent to shallow well BH101. In all other deep samples, PFOS was not detected above the laboratory reporting limit.
- PFOA was detected above the laboratory reporting limit but below SAC in BH201. No other PFOA detection was reported.

Total PFAS were reported above the laboratory reporting limit in BH201, GW128 and MW03_d.

QA/QC

Groundwater duplicate and triplicate results are within the adopted acceptance criteria of 30-50% (AS4482.1) relative percent difference (RPD) with the exception of the following:

- PFHxA between primary sample GW128 and duplicate DW2
- Total PFAS between primary sample BH111 and triplicate TW4.

RPD exceedances between primary and duplicate samples are attributed to the low concentrations detected (<5 times the LOR).

PFAS was not detected in equipment rinsate samples, indicating the sampling equipment decontamination process was sufficient to prevent cross contamination between samples.

PFAS were not detected in the field reagent blank samples, indicating sample collection, transport and analytical procedures were sufficient to prevent cross contamination.

Results of quality control checks undertaken by the laboratory are listed below table.

Report #	Analysis Within Holding Time	Surrogate Recovery	Lab. Duplicate RPD %	Lab Matrix Spike Recovery	Lab. Control Sample	Lab Method Blank
1283145-W-V2	✓	✓	✓	✓	✓	✓
1283157-W-V2	✓	X	✓	✓	✓	✓
1284389-W-V3	✓	✓	✓	✓	✓	✓
1288837-W-V3	✓	X	✓	✓	✓	✓
1285891-W	✓	✓	✓	✓	✓	✓
1286742-W	✓	X	✓	✓	✓	✓
✓ = Pass X = Fail						

Quality Assurance Criteria	Quality Control Criteria
Holding Times	**Accuracy**
PFAS: 28 Days	Surrogate, matrix spike, control sample 70-130%. Surrogate recovery 50–150%.
	Precision
	Method Blank Not detected Duplicate - No limit (<10xEQL), 0-50% (10-20xEQL), 0-200% (>20xEQL)

Report #: 1283157-W-V2

Some PFAS surrogate recoveries were outside of the recommended acceptance criteria. The laboratory reports “If the percent recoveries were less than the lower control limit, qualify detects for the analyte in associated samples as estimated with a negative bias J-“. Outliers were surrogates of PFAS compounds that have no SAC.

Report #: 1288837-W-V3 and 1286742-W

If the percent recoveries were greater than the upper control limit, qualify detects for the analyte in associated samples as estimated with a positive bias J+. Outliers were surrogates of PFAS compounds with no SAC.

Geo-Logix accepts the integrity of the analytical data.

7. CONCLUSIONS

The baseline groundwater condition as of October 2025 beneath the PFAS management area has been established.

Elevated concentrations of PFOS in shallow groundwater are localised, with greatest concentrations historically, and recently, being reported adjacent to student accommodation at BH101.

Impact to deeper groundwater is limited, PFOS being reported above SAC in one bore BH201. Groundwater flow direction in deeper groundwater is consistent with historical investigations (Geo-Logix, 2024)

8. RECOMMENDATIONS

Overall, the baseline assessment provides a robust snapshot of groundwater conditions within the PMA as of October 2025. To support any future regulatory, risk-based, or management decisions, it is recommended that bi-annual groundwater monitoring be undertaken to etablish temporal variability and PFAS concentration trends.

LIMITATIONS

This report should be read in full, and no executive summary, conclusion or other section of the report may be used or relied on in isolation, or taken as representative of the report as a whole. No responsibility is accepted by Geo-Logix, and any duty of care that may arise but for this statement is excluded, in relation to any use of any part of this report other than on this basis.

This report has been prepared for the sole benefit of and use by the Client. No other person may rely on the report for any purpose whatsoever except with Geo-Logix's express written consent. Any duty of care to third parties that would or may arise in respect of persons other than the Client, but for this statement, is excluded.

Geo-Logix has prepared this report with the diligence, care and skill which a reasonable person would expect from a reputable environmental consultancy and in accordance with environmental regulatory authority and industry standards, guidelines and assessment criteria applicable as at the date of this report. Industry standards and environmental criteria change frequently, and may change at any time after the date of this report.

9. REFERENCES

ANZECC & ARMECC (2000) *Australian and New Zealand Guidelines for Freshwater and Marine Water*, Australia and New Zealand Environment Conservation Council and Agriculture and Resource Management Council of Australia and New Zealand.

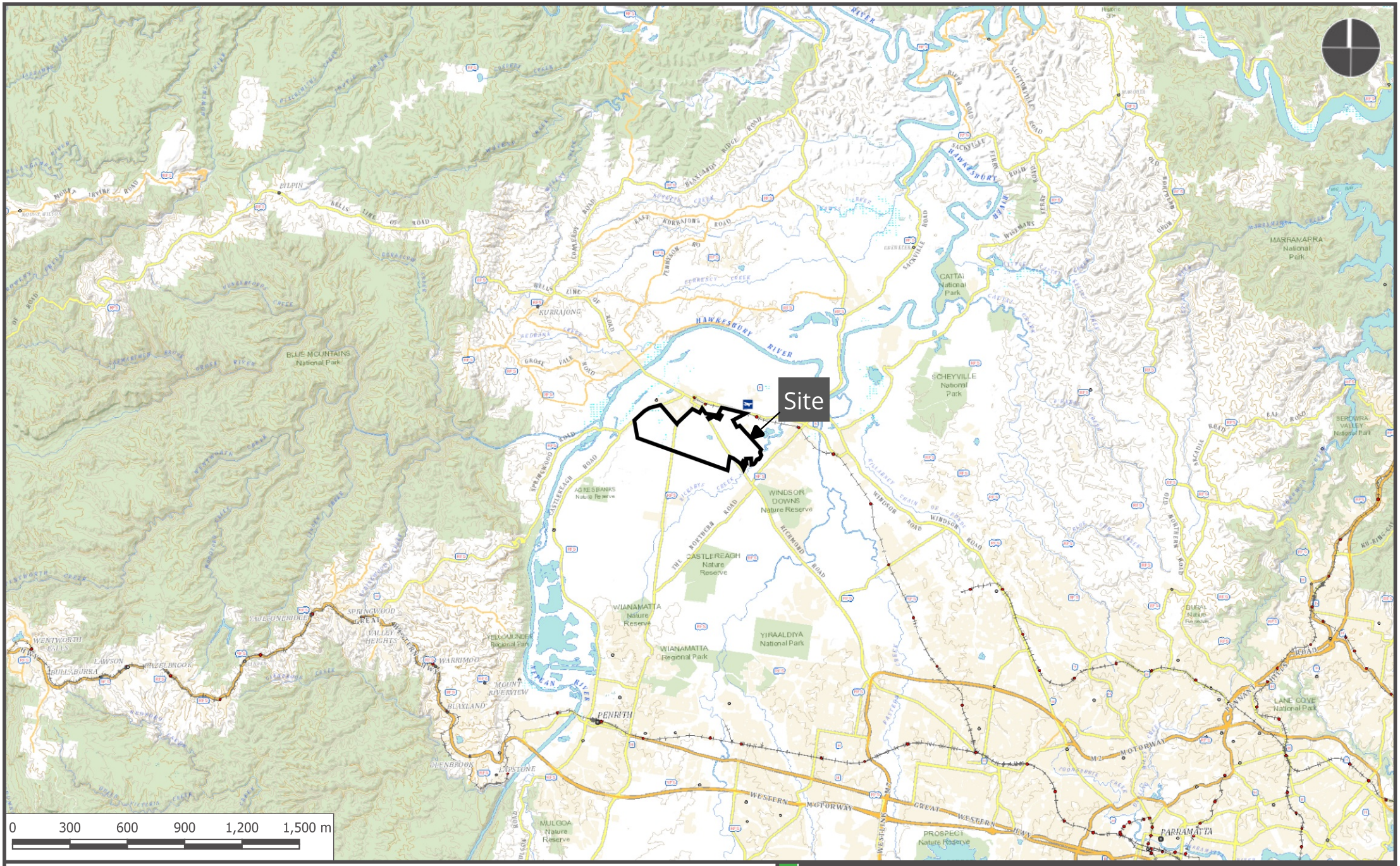
Aurecon (2022) Western Sydney University (Hawkesbury Campus) Environmental Site Assessment Report. Ref: P522692

Aurecon (2023) Western Sydney University (Hawkesbury Campus) Detailed Site Investigation – Campus Core Delineation. Report Ref: P522692.

Geo-Logix (2024) PFAS Environmental Site Assessment Report, Western Sydney University Hawkesbury Campus. Report Ref: 2301034ESAFinal_12Aug24.

PFAS NEMP (2020) *PFAS National Environmental Management Plan Version 3.0*, Heads of EPA Australia and New Zealand 2025.

FIGURES



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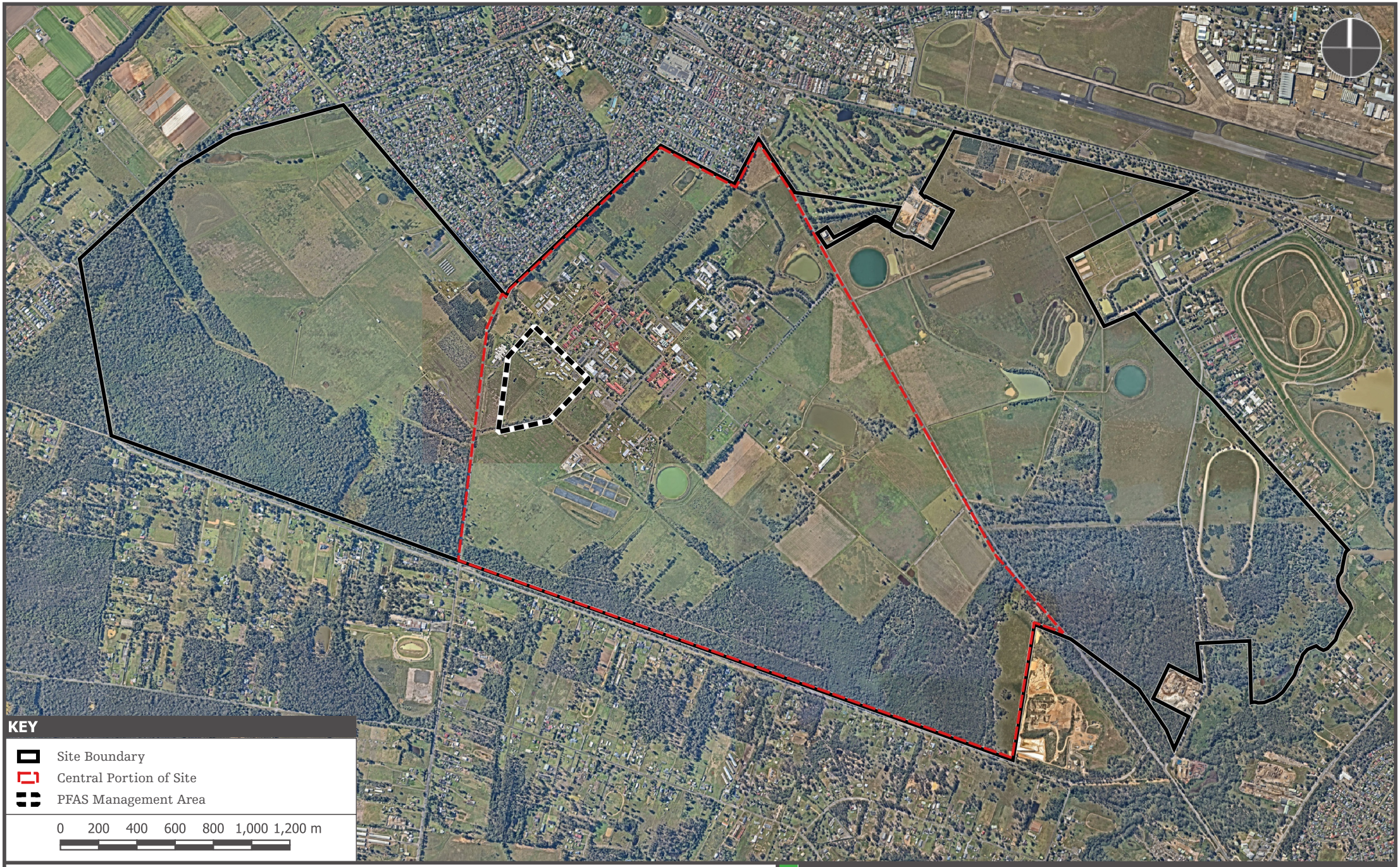
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SITE LOCATION

Groundwater Baseline Assessment
Western Sydney University - Hawkesbury Campus

Project No. 2301034g

Figure 1



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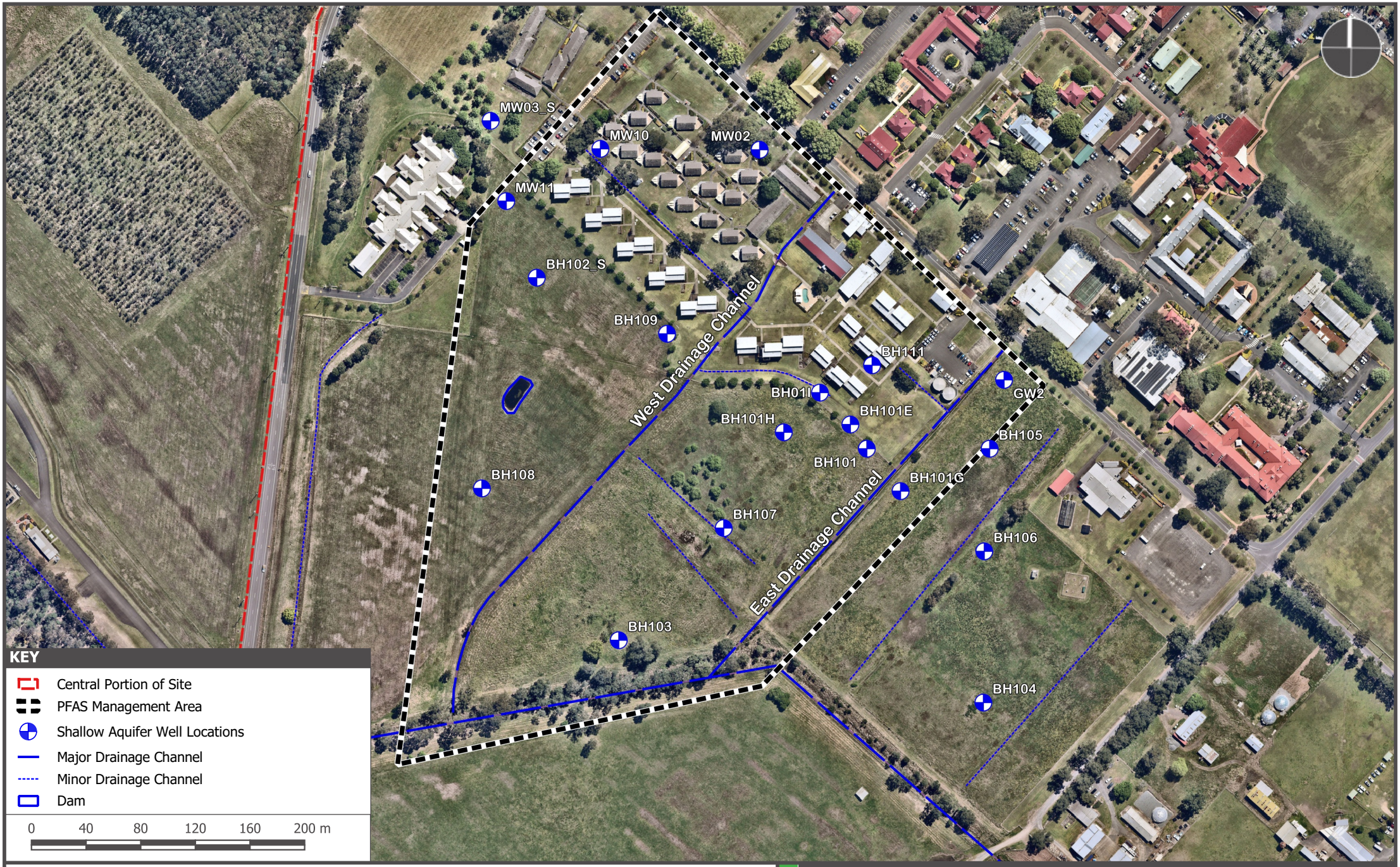
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SITE MAP

Groundwater Baseline Assessment
Western Sydney University - Hawkesbury Campus

Project No. 2301034g

Figure 2



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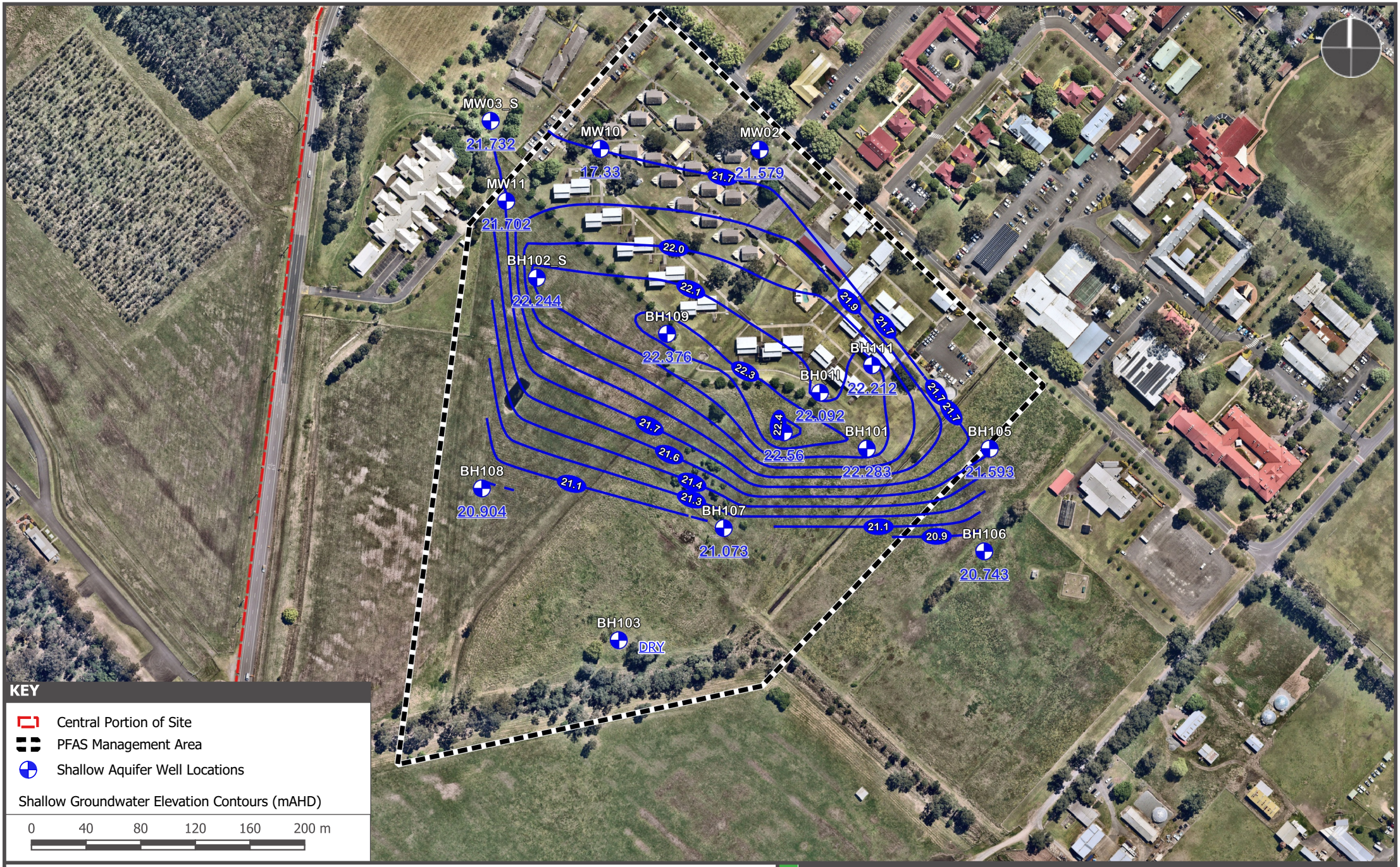
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SHALLOW AQUIFER WELL LOCATIONS

Groundwater Baseline Assessment
Western Sydney University - Hawkesbury Campus

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Figure 3



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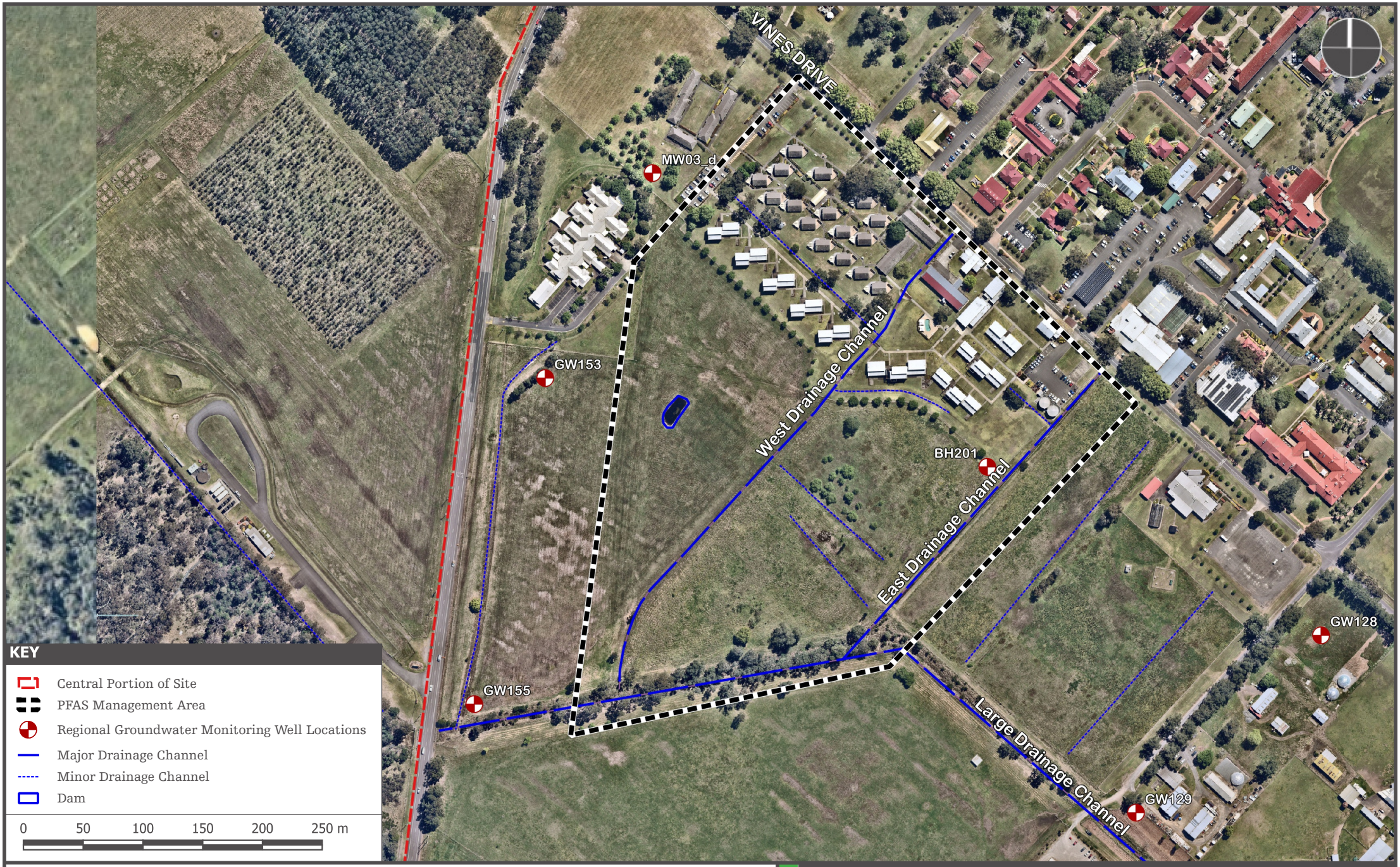
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GROUNDWATER ELEVATION MAP SHALLOW AQUIFER

Groundwater Baseline Assessment - October 2025
Western Sydney University - Hawkesbury Campus

Project No. 2301034g

Figure 3a



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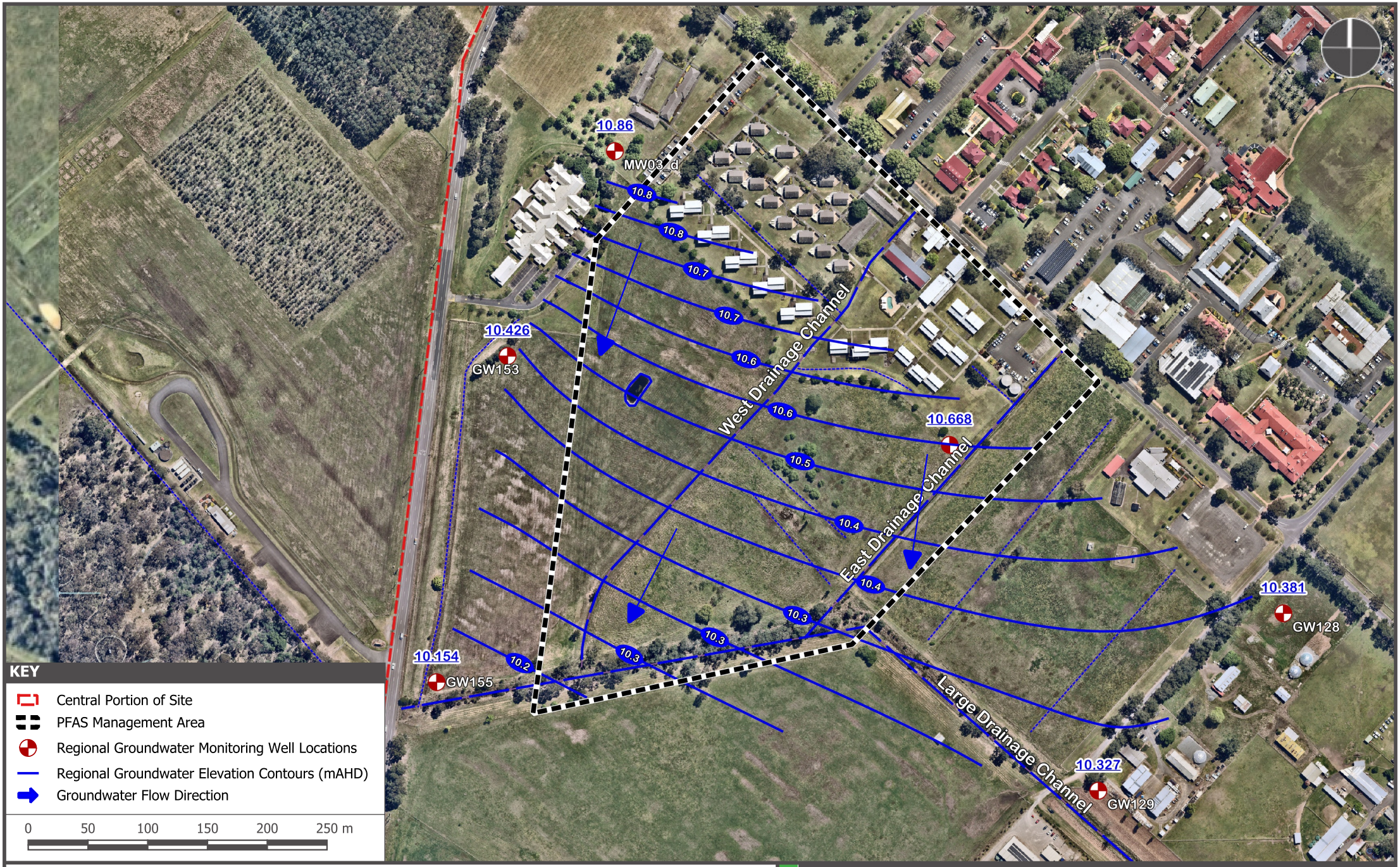
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DEEP AQUIFER WELL LOCATIONS

Groundwater Baseline Assessment
Western Sydney University - Hawkesbury Campus

Project No. 2301034g

Figure 4



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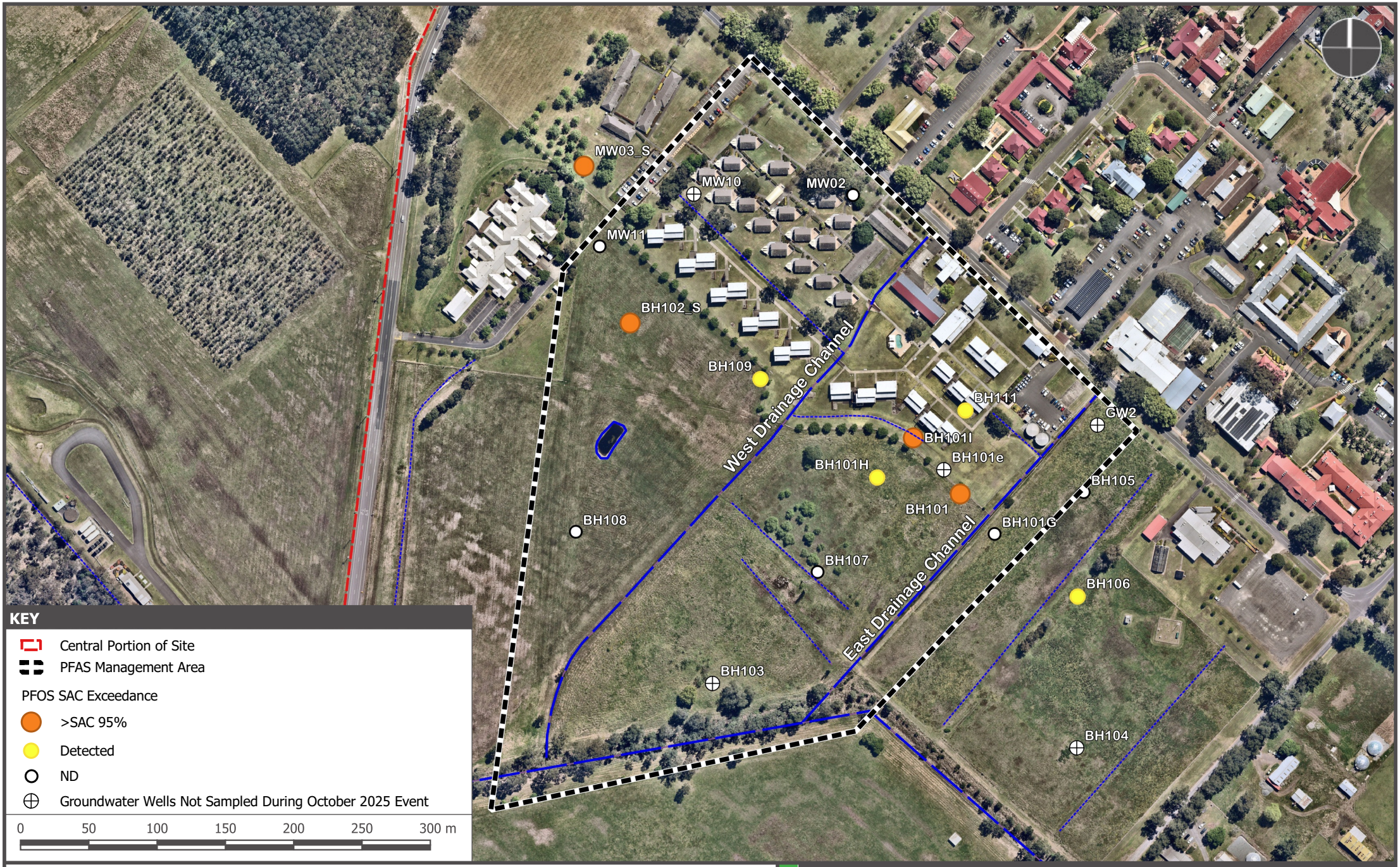
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REGIONAL GROUNDWATER ELEVATION MAP

Groundwater Baseline Assessment - October 2025
Western Sydney University - Hawkesbury Campus

Project No. 2301034g

Figure 5



KEY

- Central Portion of Site
- PFAS Management Area

PFOS SAC Exceedance

- >SAC 95%
- Detected
- ND
- Groundwater Wells Not Sampled During October 2025 Event

0 50 100 150 200 250 300 m

Geo-Logix

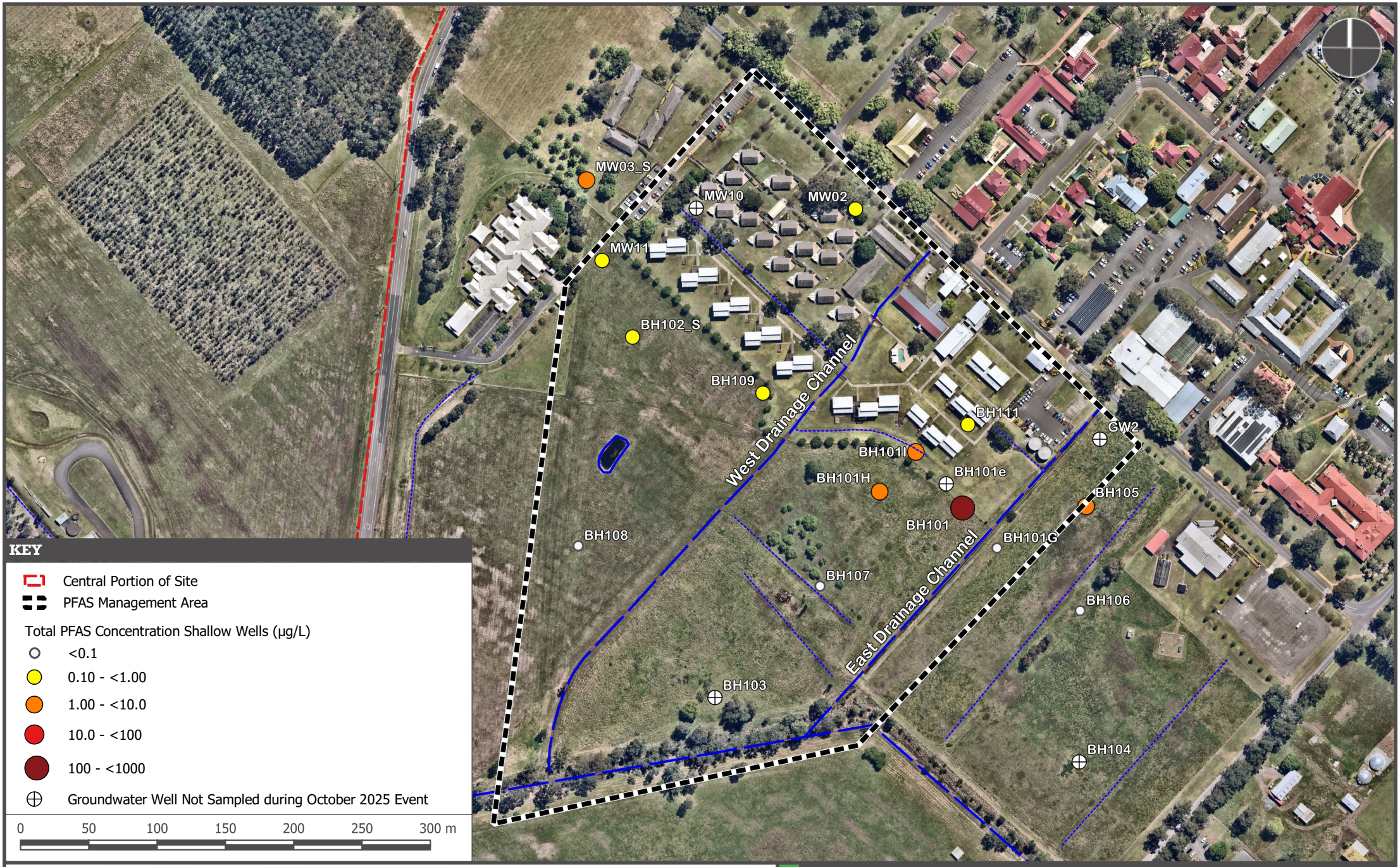
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PFOS SAC EXCEEDANCES - SHALLOW PERCHED AQUIFER

October 2025 | Groundwater Baseline Assessment
Western Sydney University - Hawkesbury Campus

Project No. 2301034g

Figure 6



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TOTAL PFAS CONCENTRATION - SHALLOW PERCHED AQUIFER

October 2025 | Groundwater Baseline Assessment
Western Sydney University - Hawkesbury Campus

Project No. 2301034g

Figure 7



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PFOS SAC EXCEEDANCES - REGIONAL GROUNDWATER

October 2025 | Groundwater Baseline Assessment
Western Sydney University - Hawkesbury Campus

Project No. 2301034g

Figure 8



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TOTAL PFAS CONCENTRATION - REGIONAL GROUNDWATER

October 2025 | Groundwater Baseline Assessment
Western Sydney University - Hawkesbury Campus

Project No. 2301034g

Figure 9

TABLES

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1						
	PFAS NEMP						
	Freshwater	Sample ID	BH101	BH101G	BH101H	BH101i	BH102
	95%	Date	23/10/2025	28/10/2025	23/10/2025	28/10/2025	21/10/2025
PFPrS	-		5.7	< 0.01	0.08	0.12	< 0.01
PFBS	-		26	< 0.01	0.14	0.37	0.03
PFPeS	-		27	< 0.01	0.08	0.27	0.03
PFHxS	-		180	< 0.01	0.77	1.9	0.4
PFHpS	-		9.1	< 0.01	< 0.01	0.02	0.02
PFOS	0.13		**250**	< 0.01	0.02	**0.25**	**0.15**
PFNS	-		0.24	< 0.01	< 0.01	< 0.01	< 0.01
PFDS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFBA	-		2.9	< 0.05	0.13	0.23	0.05
PFPeA	-		5.9	< 0.01	0.12	0.19	< 0.01
PFHxA	-		37	< 0.01	0.2	0.53	0.02
PFHpA	-		10	< 0.01	0.11	0.13	< 0.01
PFOA	220		31	< 0.01	0.09	0.1	< 0.01
PFNA	-		0.09	< 0.01	< 0.01	< 0.01	< 0.01
PFDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFUnA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFDoA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFTrDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1						
	PFAS NEMP						
	Freshwater	Sample ID	BH101	BH101G	BH101H	BH101i	BH102
	95%	Date	23/10/2025	28/10/2025	23/10/2025	28/10/2025	21/10/2025
PFTeDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
4:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
6:2 FTS	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
8:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
10:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFHxS + PFOS	-		430	< 0.01	0.79	2.15	0.55
Total PFAS	-		580	< 0.1	1.7	4.1	0.7

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1						
	PFAS NEMP						
	Freshwater	Sample ID	BH105	BH106	BH107	DW3	RPD_DW3
	95%	Date	28/10/2025	23/10/2025	23/10/2025	23/10/2025	-
PFPrS	-		0.01	< 0.01	< 0.01	< 0.01	*nc*
PFBS	-		0.04	< 0.01	< 0.01	< 0.01	*nc*
PFPeS	-		0.04	< 0.01	< 0.01	< 0.01	*nc*
PFHxS	-		1.7	0.03	< 0.01	< 0.01	*nc*
PFHpS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFOS	0.13		< 0.01	0.05	< 0.01	< 0.01	*nc*
PFNS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFDS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFBA	-		0.1	< 0.05	< 0.05	< 0.05	*nc*
PFPeA	-		0.02	< 0.01	< 0.01	< 0.01	*nc*
PFHxA	-		0.04	< 0.01	< 0.01	< 0.01	*nc*
PFHpA	-		0.03	< 0.01	< 0.01	< 0.01	*nc*
PFOA	220		0.02	< 0.01	< 0.01	< 0.01	*nc*
PFNA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFDA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFUnA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFDoA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFTrDA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1						
	PFAS NEMP						
	Freshwater	Sample ID	BH105	BH106	BH107	DW3	RPD_DW3
	95%	Date	28/10/2025	23/10/2025	23/10/2025	23/10/2025	-
PFTeDA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
MeFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
EtFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
MeFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
EtFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
MeFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
EtFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
4:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
6:2 FTS	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
8:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
10:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFHxS + PFOS	-		1.7	0.08	< 0.01	< 0.01	*nc*
Total PFAS	-		2	< 0.1	< 0.1	< 0.1	*nc*

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1					
	PFAS NEMP					
	Freshwater	Sample ID	TW3	RPD_TW3	BH108	DW1	RPD_DW1
	95%	Date	23/10/2025	-	21/10/2025	21/10/2025	-
PFPrS	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFBS	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFPeS	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFHxS	-		< 0.01	*nc*	0.01	0.01	*0%*
PFHpS	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFOS	0.13		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFNS	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFDS	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFBA	-		< 0.05	*nc*	< 0.05	< 0.05	*nc*
PFPeA	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFHxA	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFHpA	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFOA	220		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFNA	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFDA	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFUnA	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFDoA	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFTrDA	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1						
	PFAS NEMP						
	Freshwater	Sample ID	TW3	RPD_TW3	BH108	DW1	RPD_DW1
	95%	Date	23/10/2025	-	21/10/2025	21/10/2025	-
PFTeDA	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFOSA	-		< 0.05	*nc*	< 0.05	< 0.05	*nc*
MeFOSA	-		< 0.05	*nc*	< 0.05	< 0.05	*nc*
EtFOSA	-		< 0.05	*nc*	< 0.05	< 0.05	*nc*
MeFOSE	-		< 0.05	*nc*	< 0.05	< 0.05	*nc*
EtFOSE	-		< 0.05	*nc*	< 0.05	< 0.05	*nc*
MeFOSAA	-		< 0.05	*nc*	< 0.05	< 0.05	*nc*
EtFOSAA	-		< 0.05	*nc*	< 0.05	< 0.05	*nc*
4:2 FTS	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
6:2 FTS	-		< 0.05	*nc*	< 0.05	< 0.05	*nc*
8:2 FTS	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
10:2 FTS	-		< 0.01	*nc*	< 0.01	< 0.01	*nc*
PFHxS + PFOS	-		< 0.01	*nc*	0.01	0.01	*0%*
Total PFAS	-		< 0.1	*nc*	< 0.1	< 0.1	*nc*

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1					
	PFAS NEMP					
	Freshwater	Sample ID	TW1	RPD_TW1	BH109	BH111	DW4
	95%	Date	21/10/2025	-	21/10/2025	28/10/2025	28/10/2025
PFPrS	-		< 0.01	*nc*	< 0.01	0.01	< 0.01
PFBS	-		< 0.01	*nc*	0.02	0.02	0.02
PFPeS	-		< 0.01	*nc*	0.01	< 0.01	< 0.01
PFHxS	-		< 0.01	*nc*	0.14	0.04	0.04
PFHpS	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFOS	0.13		< 0.01	*nc*	0.02	0.02	0.02
PFNS	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFDS	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFBA	-		< 0.05	*nc*	< 0.05	< 0.05	< 0.05
PFPeA	-		< 0.01	*nc*	< 0.01	0.02	0.02
PFHxA	-		< 0.01	*nc*	0.01	0.02	0.02
PFHpA	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFOA	220		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFNA	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFDA	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFUnA	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFDoA	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFTrDA	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1					
	PFAS NEMP					
	Freshwater	Sample ID	TW1	RPD_TW1	BH109	BH111	DW4
	95%	Date	21/10/2025	-	21/10/2025	28/10/2025	28/10/2025
PFTeDA	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFOSA	-		< 0.05	*nc*	< 0.05	< 0.05	< 0.05
MeFOSA	-		< 0.05	*nc*	< 0.05	< 0.05	< 0.05
EtFOSA	-		< 0.05	*nc*	< 0.05	< 0.05	< 0.05
MeFOSE	-		< 0.05	*nc*	< 0.05	< 0.05	< 0.05
EtFOSE	-		< 0.05	*nc*	< 0.05	< 0.05	< 0.05
MeFOSAA	-		< 0.05	*nc*	< 0.05	< 0.05	< 0.05
EtFOSAA	-		< 0.05	*nc*	< 0.05	< 0.05	< 0.05
4:2 FTS	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
6:2 FTS	-		< 0.05	*nc*	< 0.05	< 0.05	< 0.05
8:2 FTS	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
10:2 FTS	-		< 0.01	*nc*	< 0.01	< 0.01	< 0.01
PFHxS + PFOS	-		< 0.01	*nc*	0.16	0.06	0.06
Total PFAS	-		< 0.1	*nc*	0.2	0.1	0.1

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1						
	PFAS NEMP						
	Freshwater	Sample ID	RPD_DW4	TW4	RPD_TW4	MW02	MW03-S
	95%	Date	-	28/10/2025	-	28/10/2025	28/10/2025
PFPrS	-		*nc*	0.01	*0%*	< 0.01	0.08
PFBS	-		*0%*	0.02	*0%*	0.02	0.26
PFPeS	-		*nc*	< 0.01	*nc*	< 0.01	0.24
PFHxS	-		*0%*	0.04	*0%*	0.05	1.7
PFHpS	-		*nc*	< 0.01	*nc*	< 0.01	0.03
PFOS	0.13		*0%*	0.02	*0%*	< 0.01	**0.35**
PFNS	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01
PFDS	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01
PFBA	-		*nc*	< 0.05	*nc*	< 0.05	< 0.05
PFPeA	-		*0%*	0.03	*40%*	0.04	< 0.01
PFHxA	-		*0%*	0.03	*40%*	0.05	0.05
PFHpA	-		*nc*	< 0.01	*nc*	0.03	< 0.01
PFOA	220		*nc*	< 0.01	*nc*	0.04	0.01
PFNA	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01
PFDA	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01
PFUnA	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01
PFDoA	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01
PFTrDA	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1					
	PFAS NEMP					
	Freshwater	Sample ID	RPD_DW4	TW4	RPD_TW4	MW02	MW03-S
	95%	Date	-	28/10/2025	-	28/10/2025	28/10/2025
PFTeDA	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01
PFOSA	-		*nc*	< 0.05	*nc*	< 0.05	< 0.05
MeFOSA	-		*nc*	< 0.05	*nc*	< 0.05	< 0.05
EtFOSA	-		*nc*	< 0.05	*nc*	< 0.05	< 0.05
MeFOSE	-		*nc*	< 0.05	*nc*	< 0.05	< 0.05
EtFOSE	-		*nc*	< 0.05	*nc*	< 0.05	< 0.05
MeFOSAA	-		*nc*	< 0.05	*nc*	< 0.05	< 0.05
EtFOSAA	-		*nc*	< 0.05	*nc*	< 0.05	< 0.05
4:2 FTS	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01
6:2 FTS	-		*nc*	< 0.05	*nc*	< 0.05	< 0.05
8:2 FTS	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01
10:2 FTS	-		*nc*	< 0.01	*nc*	< 0.01	< 0.01
PFHxS + PFOS	-		*0%*	0.06	*0%*	0.05	2.05
Total PFAS	-		*0%*	0.2	*67%*	0.2	2.7

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1		
	PFAS NEMP		
	Freshwater	Sample ID	MW11
	95%	Date	28/10/2025
PFPrS	-		0.04
PFBS	-		0.09
PFPeS	-		0.06
PFHxS	-		0.53
PFHpS	-		< 0.01
PFOS	0.13		< 0.01
PFNS	-		< 0.01
PFDS	-		< 0.01
PFBA	-		< 0.05
PFPeA	-		< 0.01
PFHxA	-		< 0.01
PFHpA	-		< 0.01
PFOA	220		< 0.01
PFNA	-		< 0.01
PFDA	-		< 0.01
PFUnA	-		< 0.01
PFDoA	-		< 0.01
PFTrDA	-		< 0.01

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Shallow Perched Water Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College St, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1		
	PFAS NEMP		
	Freshwater	Sample ID	MW11
	95%	Date	28/10/2025
PFTeDA	-		< 0.01
PFOSA	-		< 0.05
MeFOSA	-		< 0.05
EtFOSA	-		< 0.05
MeFOSE	-		< 0.05
EtFOSE	-		< 0.05
MeFOSAA	-		< 0.05
EtFOSAA	-		< 0.05
4:2 FTS	-		< 0.01
6:2 FTS	-		< 0.05
8:2 FTS	-		< 0.01
10:2 FTS	-		< 0.01
PFHxS + PFOS	-		0.53
Total PFAS	-		0.7

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW3 = duplicate of BH107

TW3 = triplicate of BH107

DW1 = duplicate of BH108

TW1 = triplicate of BH108

DW4 = duplicate of BH111

TW4 = triplicate of BH111

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 2 : Summary of Regional Groundwater Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College Street, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1						
	PFAS NEMP						
	Freshwater	Sample ID	BH201	GW128	DW2	TW2	RPD_DW2
	95%	Date	23/10/2025	23/10/2025	23/10/2025	23/10/2025	-
PFPrS	-		0.64	0.04	0.04	0.04	*0%*
PFBS	-		2.1	0.08	0.08	0.06	*0%*
PFPeS	-		1.6	0.04	0.04	0.04	*0%*
PFHxS	-		6.6	0.17	0.17	0.16	*0%*
PFHpS	-		0.12	< 0.01	< 0.01	< 0.01	*nc*
PFOS	0.13		**0.22**	< 0.01	< 0.01	< 0.01	*nc*
PFNS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFDS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFBA	-		0.2	< 0.05	< 0.05	< 0.05	*nc*
PFPeA	-		0.3	< 0.01	< 0.01	< 0.01	*nc*
PFHxA	-		1.6	0.01	0.02	0.01	*67%*
PFHpA	-		0.56	< 0.01	< 0.01	< 0.01	*nc*
PFOA	220		0.55	< 0.01	< 0.01	< 0.01	*nc*
PFNA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFDA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFUnA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFDoA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFTrDA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW2 = duplicate of GW128

TW2 = triplicate of GW128

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 2 : Summary of Regional Groundwater Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College Street, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1						
	PFAS NEMP						
	Freshwater	Sample ID	BH201	GW128	DW2	TW2	RPD_DW2
	95%	Date	23/10/2025	23/10/2025	23/10/2025	23/10/2025	-
PFTeDA	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
MeFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
EtFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
MeFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
EtFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
MeFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
EtFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
4:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
6:2 FTS	-		< 0.05	< 0.05	< 0.05	< 0.05	*nc*
8:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
10:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	*nc*
PFHxS + PFOS	-		6.82	0.17	0.17	0.16	*0%*
Total PFAS	-		14	0.3	0.4	0.3	*29%*

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW2 = duplicate of GW128

TW2 = triplicate of GW128

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 2 : Summary of Regional Groundwater Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College Street, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1					
	PFAS NEMP					
	Freshwater	Sample ID	RPD_TW2	GW129	GW153	GW155	MW03-D
	95%	Date	-	23/10/2025	21/10/2025	21/10/2025	21/10/2025
PFPrS	-		*0%*	0.01	< 0.01	< 0.01	0.04
PFBS	-		*29%*	< 0.01	< 0.01	< 0.01	0.12
PFPeS	-		*0%*	< 0.01	< 0.01	< 0.01	0.09
PFHxS	-		*6%*	< 0.01	< 0.01	< 0.01	0.73
PFHpS	-		*nc*	< 0.01	< 0.01	< 0.01	0.01
PFOS	0.13		*nc*	< 0.01	< 0.01	< 0.01	0.07
PFNS	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
PFDS	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
PFBA	-		*nc*	< 0.05	< 0.05	< 0.05	< 0.05
PFPeA	-		*nc*	0.02	< 0.01	< 0.01	< 0.01
PFHxA	-		*0%*	< 0.01	< 0.01	< 0.01	0.03
PFHpA	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
PFOA	220		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
PFNA	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
PFDA	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
PFUnA	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
PFDoA	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
PFTrDA	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW2 = duplicate of GW128

TW2 = triplicate of GW128

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 2 : Summary of Regional Groundwater Analytical Data - PFAS

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

2 College Street, Richmond NSW

Western Sydney University – Hawkesbury Campus

	Criteria 1					
	PFAS NEMP					
	Freshwater	Sample ID	RPD_TW2	GW129	GW153	GW155	MW03-D
	95%	Date	-	23/10/2025	21/10/2025	21/10/2025	21/10/2025
PFTeDA	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
PFOSA	-		*nc*	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSA	-		*nc*	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSA	-		*nc*	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSE	-		*nc*	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSE	-		*nc*	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSAA	-		*nc*	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSAA	-		*nc*	< 0.05	< 0.05	< 0.05	< 0.05
4:2 FTS	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
6:2 FTS	-		*nc*	< 0.05	< 0.05	< 0.05	< 0.05
8:2 FTS	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
10:2 FTS	-		*nc*	< 0.01	< 0.01	< 0.01	< 0.01
PFHxS + PFOS	-		*6%*	< 0.01	< 0.01	< 0.01	0.8
Total PFAS	-		*0%*	< 0.1	< 0.1	< 0.1	1.1

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DW2 = duplicate of GW128

TW2 = triplicate of GW128

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 3 : Summary of Groundwater Analytical Data - PFAS (QA/QC)

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

QA/QC - Blanks and Rinsates Samples

Western Sydney University – Hawkesbury Campus

	Criteria 1						
	PFAS NEMP						
	Freshwater	Sample ID	BLANK 1	BLANK 2	BLANK 3	R1	R2
	95%	Date	21/10/2025	23/10/2025	28/10/2025	21/10/2025	23/10/2025
PFPrS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFBS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFPeS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFHxS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFHpS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFOS	0.13		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFNS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFDS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFBA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
PFPeA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFHxA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFHpA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFOA	220		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFNA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFUnA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFDoA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFTrDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Bold/red indicates exceedance of assessment criteria

Total concentrations in µg/L

– = assessment criteria not available

BLANK 1 = blank sample

BLANK 2 = blank sample

BLANK 3 = blank sample

R1 = rinsate sample

R2 = rinsate sample

R3 = rinsate sample

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Table 3 : Summary of Groundwater Analytical Data - PFAS (QA/QC)

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

QA/QC - Blanks and Rinsates Samples

Western Sydney University – Hawkesbury Campus

	Criteria 1						
	PFAS NEMP						
	Freshwater	Sample ID	BLANK 1	BLANK 2	BLANK 3	R1	R2
	95%	Date	21/10/2025	23/10/2025	28/10/2025	21/10/2025	23/10/2025
PFTeDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
4:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
6:2 FTS	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
8:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
10:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFHxS + PFOS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
Total PFAS	-		< 0.1	< 0.1	< 0.1	< 0.1	< 0.1

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Bold/red indicates exceedance of assessment criteria

Total concentrations in µg/L

– = assessment criteria not available

BLANK 1 = blank sample

BLANK 2 = blank sample

BLANK 3 = blank sample

R1 = rinsate sample

R2 = rinsate sample

R3 = rinsate sample

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Table 3 : Summary of Groundwater Analytical Data - PFAS (QA/QC)

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

QA/QC - Blanks and Rinsates Samples

Western Sydney University – Hawkesbury Campus

	Criteria 1		
	PFAS NEMP		
	Freshwater	Sample ID	R3
	95%	Date	28/10/2025
PFPrS	-		< 0.01
PFBS	-		< 0.01
PFPeS	-		< 0.01
PFHxS	-		< 0.01
PFHpS	-		< 0.01
PFOS	0.13		< 0.01
PFNS	-		< 0.01
PFDS	-		< 0.01
PFBA	-		< 0.05
PFPeA	-		< 0.01
PFHxA	-		< 0.01
PFHpA	-		< 0.01
PFOA	220		< 0.01
PFNA	-		< 0.01
PFDA	-		< 0.01
PFUnA	-		< 0.01
PFDoA	-		< 0.01
PFTrDA	-		< 0.01

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Bold/red indicates exceedance of assessment criteria

Total concentrations in µg/L

– = assessment criteria not available

BLANK 1 = blank sample

BLANK 2 = blank sample

BLANK 3 = blank sample

R1 = rinsate sample

R2 = rinsate sample

R3 = rinsate sample

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Table 3 : Summary of Groundwater Analytical Data - PFAS (QA/QC)

Groundwater Baseline Assessment – Privileged and Confidential

Project No.: 2301034g

QA/QC - Blanks and Rinsates Samples

Western Sydney University – Hawkesbury Campus

	Criteria 1		
	PFAS NEMP		
	Freshwater	Sample ID	R3
	95%	Date	28/10/2025
PFTeDA	-		< 0.01
PFOSA	-		< 0.05
MeFOSA	-		< 0.05
EtFOSA	-		< 0.05
MeFOSE	-		< 0.05
EtFOSE	-		< 0.05
MeFOSAA	-		< 0.05
EtFOSAA	-		< 0.05
4:2 FTS	-		< 0.01
6:2 FTS	-		< 0.05
8:2 FTS	-		< 0.01
10:2 FTS	-		< 0.01
PFHxS + PFOS	-		< 0.01
Total PFAS	-		< 0.1

Notes:

Criteria 1 = PFAS NEMP 3.0 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Bold/red indicates exceedance of assessment criteria

Total concentrations in µg/L

– = assessment criteria not available

BLANK 1 = blank sample

BLANK 2 = blank sample

BLANK 3 = blank sample

R1 = rinsate sample

R2 = rinsate sample

R3 = rinsate sample

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

ATTACHMENT A

GROUNDWATER SAMPLE LOG

Geo-Logix

WELL ID: MW03_d

PROJECT INFORMATION

PROJECT NUMBER:	23010349.	INITIALS:	PDP - HM
DATE:	21-10-25.	WEATHER:	SUNNY.
SAMPLING METHOD:	LOW FLOW: ☐	HYDRASLEEVE: ☒	BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	12.665.	TOTAL DEPTH (mBTOC):	14.692.	STICK-UP (m):	—
DEPTH TO PSH (mBTOC):	—	THICKNESS OF PSH (m):	—	TIME:	14:00

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
1405.	—.	—	5.65	598.	-37.1	2.60.	21.4
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) ~~Soft~~ base. no odour, no sheen. Gatic.

No hydrasleeve installed after sampling

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: MW03_d	SAMPLE ID:	SAMPLE ID:
SAMPLE TIME: 14 pm	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: pfas.	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: GW155.

PROJECT INFORMATION

PROJECT NUMBER: 2301034g.	**INITIALS:** HM. PDP.
DATE: 21.10.25.	**WEATHER:** SUNNY.

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☒ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC): 13.432.	**TOTAL DEPTH (mBTOC):** 15.649.	**STICK-UP (m):** 0.63
DEPTH TO PSH (mBTOC): –	**THICKNESS OF PSH (m):** –	**TIME:** 10:30

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
10:35	–	–	6.19	7497	-94.5	5.14	20.6
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) NONUIENT. Light brown/ clear... Sediments at bottom of Hydrosleeve. No odour/ no sheen. NOT HYDRASLEEVE INSTALLED AFTER.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: GW155	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 10:30 am	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 1	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS.	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: GW153..

PROJECT INFORMATION

PROJECT NUMBER:	2301034.	INITIALS:	HN - PDP
DATE:	24-10-25.	WEATHER:	SUNNY
SAMPLING METHOD:	LOW FLOW: ☐	HYDRASLEEVE: ☑	BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	13.558	TOTAL DEPTH (mBTOC):	16.793	STICK-UP (m):	0.755.
DEPTH TO PSH (mBTOC):	-	THICKNESS OF PSH (m):	-	TIME:	10:00

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
10:10			5.93	9740	-76.8	1.48	21.5
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) MONUMENT. clear, brown sediments at the bottom. Soft base. NO odour. no sheen. NOT HYDRASLEEVE INSTALLED AFTER.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: GW153	SAMPLE ID:	SAMPLE ID:
SAMPLE TIME: 10:10	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PFAS	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: MW11 (?). check w/ well is it

take sample! next time

PROJECT INFORMATION

PROJECT NUMBER:	2301034g.	**INITIALS:**	PDP. HM.
DATE:	21.10.2025	**WEATHER:**	Sunny

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	2.428	**TOTAL DEPTH (mBTOC):**	6.952	**STICK-UP (m):**	1.07.
DEPTH TO PSH (mBTOC):	-	**THICKNESS OF PSH (m):**	—	**TIME:**	9:40 AM.

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR)

Soft base, only well casing, no monument, No Orange Cap.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING NO SAMPLING.

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID:	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME:	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS:	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS:	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Need orange cap, survey, monument, padlock.

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: BH109.

PROJECT INFORMATION

PROJECT NUMBER: 23010348	INITIALS: PDP-HN.
DATE: 21-10-25	WEATHER: SUNNY.
SAMPLING METHOD: LOW FLOW: ☒ HYDRASLEEVE: ☐ BAILER: ☐	

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC): 1.843	TOTAL DEPTH (mBTOC): 4.635.	STICK-UP (m): 0.96.
DEPTH TO PSH (mBTOC): —	THICKNESS OF PSH (m): —	TIME: 11:55.

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
12:00	2.087.	0.5 L	6.62	781	-89.6	0.05.	18.1
12:05.	2.345	1.5 L	6.56.	772	-112.9	0.09	18.5.
12:10.	2.460.	2.5 L	6.56	767	-157.9.	0.05.	19.
12:15.	2.573	2.8 L.	6.57	775.	-184.5.	0.09	20.
12:19	2.615	2.9 L.	6.58.	780	-198.0	0.16	20.5
lost 5 min slow pump speed.			6.58	783	-204.3	0.19	20.5
12:20.	2.	3 L.					
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

pump really slow.

1.3 - 4.5 screened interval.

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) monument. clear/light brown. no odour/ no sheen. leave tubing inside well after sampling. No orange cap on well.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<200% RECHARGE AFTER 24 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE X	TRIPLICATE X
SAMPLE ID: BH109	SAMPLE ID:	SAMPLE ID:
SAMPLE TIME: 12:20.	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PFAS.	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

Need padlock, orange cap, re survey.

GROUNDWATER SAMPLE LOG

Geo-Logix

WELL ID: BH108

PROJECT INFORMATION

PROJECT NUMBER: 2301034g.	INITIALS: HT - POP
DATE: 21-10-25	WEATHER: SUNNY.

SAMPLING METHOD: LOW FLOW: ☒ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC): 1.979	TOTAL DEPTH (mBTOC): 4.275.	STICK-UP (m): 0.655
DEPTH TO PSH (mBTOC): —	THICKNESS OF PSH (m): —	TIME: 10:55

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
11:15.	2.291	1L	6.44.	363.4.	-128.0	0.31	18.1
11:20	2.566	2L	6.44	362.3	-146.3	0.30	18.0
11:25.	2.744	2.8L	6.43	355.3	-154.3	0.31	17.9
11:30	2.890	3L.	6.45.	360.3	-155.9	0.32	18.1
11:35.	2.890	3.3L	6.45.	365.0	-156.5	0.33	18.2
pump not working properly, turn off in some speeds.							

TOTAL PURGE VOLUME (L): APPROX. SAMPLE PURGE RATE (LPM):

OBSERVATIONS DURING SAMPLING (1.1-3.6 screened interval).

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) MONUMENT. soft box, light brown mud on box lip. no light brown. clear, no odour - no sheen. pump low rate. rising inside well after sampling.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (~~WATER DOES NOT RECOVER~~) ☒

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING) 2.890

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: BH108.	SAMPLE ID: DW1	SAMPLE ID: ~~TB~~TW1
SAMPLE TIME: 11:35	SAMPLE TIME: 11:40	SAMPLE TIME: 11:45.
NO. CONTAINERS: 1	NO. CONTAINERS 1	NO. CONTAINERS 1
ANALYSIS: PFAS.	ANALYSIS: PFAS	ANALYSIS: PFAS.

ORIGINAL FIELD RECORD

Need padlock / ee survey.

BH102_d SWL = 2.172. GATIC cover.
T.D = 5.703.

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: BH102.

PROJECT INFORMATION

PROJECT NUMBER: 2301634g.	INITIALS: HM - POP.
DATE: 21-10-25.	WEATHER: SUNNY
SAMPLING METHOD: LOW FLOW: ☑ HYDRASLEEVE: ☐ BAILER: ☐	

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC): 1.995.	TOTAL DEPTH (mBTOC): 5.219.	STICK-UP (m): 0.90
DEPTH TO PSH (mBTOC): —	THICKNESS OF PSH (m): —	TIME: 12:40

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
12:55.	2.598	12	6.31	525.	-138.2	0.06.	17.3
13:00	3.332.	2.3.	6.32.	506.	-169.8.	0.08.	17.8.
13:03	3.637	2.5.	6.34	508.	-165.0	0.20	18.0
13:05	3.713	2.6	6.38	510.	-161.8.	0.25.	18.5. — slowest speed.
13:07	3.908.	2.7.	6.45.	507.	-160.0.	0.50.	18.9
13:10	4.260	4.	6.54	489.	-152.9.	1.50.	17.8.
post note. see what happen. all parameters change. back to slow speed.							
13:15.	4.521	4.2.	6.55	513.	-141.5	1.60	19.6.
13:20	4.505	4.7	6.55.	517.	-141.6	1.61	19.5.
13:25	4.504	4.9	6.54	518	-145.7	1.70	19.6.
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

screen interval not known. sample x 4m approx.

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) movement. clear. no odour. no sheen.

start pumping fast speed, no recharging then: slow pump. 1/2 speed.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

13:20 start slowing to recharge. take sample.

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING) 4.504.

ORIGINAL	DUPLICATE X	TRIPLICATE X.
SAMPLE ID: BH102.	SAMPLE ID:	SAMPLE ID:
SAMPLE TIME: 13:25	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PFAS.	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

- Need padlock
- TO be surveyed AGAIN.

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: BH103

PROJECT INFORMATION

PROJECT NUMBER:	23010348	INITIALS:	PDP / HM
DATE:	23-10-25.	WEATHER:	SUNNY
SAMPLING METHOD:	LOW FLOW: ☒	HYDRASLEEVE: ☐	BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC): DRY.	TOTAL DEPTH (mBTOC): 5.506.	STICK-UP (m): 1.05
DEPTH TO PSH (mBTOC): —	THICKNESS OF PSH (m): —	TIME: 10.25

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
WELL WAS DRY. IP STOP WORKING... &							
		WHEN DECON. MEASURE FINAL DEPHT W/ MEASURING					
		TAPE. (DRY AFTER measuring).					
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) MONUMENT. NEED NEW PADLOCK.

clay (light brown. moist. high plasticity on IP).

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: BH103. DRY	SAMPLE ID:	SAMPLE ID:
SAMPLE TIME:	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 4	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PFAS	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: BH101

PROJECT INFORMATION

PROJECT NUMBER:	2301034g.	**INITIALS:**	POP-HT
DATE:	23-10-25	**WEATHER:**	SUNNY.

SAMPLING METHOD: LOW FLOW: ☒ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC): 2.351	**TOTAL DEPTH (mBTOC):** 5.917.	**STICK-UP (m):** 1.075.
DEPTH TO PSH (mBTOC): –	**THICKNESS OF PSH (m):** –	**TIME:** 13:00

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
13:05.	2.637	1L	6.05.	1718	-12.9	0.98	17.5.
13:10	3.064	3.5.	6.07	1649	-124.1	1.13	17.5.
13:15	3.171	4.5.	6.07	1582	-141.0	1.15	17.5
13:20	3.249	5.8	6.05	1508	-137.7	1.26	17.7.
13:25	3.299	6	6.05	1499	-132.3	1.25	17.7.
13:30	3.316.	6.5.	6.02	1498	-139.7	1.17	18.
13:35	3.325	6.8	6.02	1501	-139.9	1.20	18.
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) NO ORANGE CAP. clear/cloudy NO ODOUR / NO SLEEVE. LEAVE TUBING IN. MONUMENT.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING) 3.325.

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: BH101	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 13:35	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 1	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS.	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: BH201

PROJECT INFORMATION

PROJECT NUMBER:	2301034g.	INITIALS:	POP.
DATE:	23-10-25.	WEATHER:	CLOUDY.
SAMPLING METHOD:	LOW FLOW: ☐	HYDRASLEEVE: ☒	BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC): 13.632.	TOTAL DEPTH (mBTOC): 20.630	STICK-UP (m): 0.62.
DEPTH TO PSH (mBTOC): —	THICKNESS OF PSH (m): —	TIME: 13:00.

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
13pm	—	—	6.18.	1094	-139.1	1.32	20.2
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) clear, no odour no sheen. nonvent.

new hydrasleeve not installed after sampling

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: BH201	SAMPLE ID: X	SAMPLE ID: X.
SAMPLE TIME: 13:00	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PCAS.	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: GW129

PROJECT INFORMATION

PROJECT NUMBER:	2301034	INITIALS:	PDP.
DATE:	23-10-25.	WEATHER:	SUNNY

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☒ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	12.394	TOTAL DEPTH (mBTOC):		STICK-UP (m):	—
DEPTH TO PSH (mBTOC):	—	THICKNESS OF PSH (m):	—	TIME:	9:45.

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
9:50	—	—	5.78	4722	-254.7	4.65	20.1
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) Smell like pond water. light brown/green. no sheen. bore in good condition. new hydrasleeve not installed after sampling.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: GW129.	SAMPLE ID: —	SAMPLE ID: —
SAMPLE TIME: 9:45	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PFAS.	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

GROUNDWATER SAMPLE LOG

Geo-Logix

WELL ID: GW128

PROJECT INFORMATION

PROJECT NUMBER:	2301034g.	**INITIALS:**	POP
DATE:	23-10-25	**WEATHER:**	SUNNY.

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☒ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	12.408	**TOTAL DEPTH (mBTOC):**	14.851	**STICK-UP (m):**	—
DEPTH TO PSH (mBTOC):	—	**THICKNESS OF PSH (m):**	—	**TIME:**	9:30

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
9:35.	—	—	5.46	585	-47.1	5.04	20.7
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) GW128 + DW2 w/ 1° HYDRASLEEVE. clear, no odour, no sheen. TW2. w/ 2° hydrosleeve - light brown / dead ants on sampler.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

new hydrasleeve not installed after sampling.

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING) 12.408. to refill HYDRASLEEVE.

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: GW128.	**SAMPLE ID:** DW2	**SAMPLE ID:** TW2
SAMPLE TIME: 9:30	**SAMPLE TIME:** 9:35	**SAMPLE TIME:** 9:40.
NO. CONTAINERS: 1	**NO. CONTAINERS** 1	**NO. CONTAINERS** 1
ANALYSIS: PFAS	**ANALYSIS:** PFAS	**ANALYSIS:** PFAS

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: BH106

PROJECT INFORMATION

PROJECT NUMBER:	2301034 9	INITIALS:	HM | PDP
DATE:	23·10·25	WEATHER:	Sunny

SAMPLING METHOD: LOW FLOW: [X] HYDRASLEEVE: [ ] BAILER: [ ]

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	2·332	TOTAL DEPTH (mBTOC):	5·120	STICK-UP (m):	0·97
DEPTH TO PSH (mBTOC):	—	THICKNESS OF PSH (m):	—	TIME:	9:45 am

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
10:00	2·516	1 L	6·33	577	-11.0	0·72	18°c
10·05	2.778	2L	6·32	532	-15.9	0·79	18·2°c
10:10	2.991	3·1L	6·31	513	-23·1	0.83	18°c
10:15	3·193	4.0L	6.34	529	-39.1	0.52	18·8°c
10:20	3.294	4.5 L	6·31	525	-49.6	0·47	20·2c
10:25	3·371	4·8L	6·31	518	-54.0	0·45	20·5°c
10:30	3·451	5·0L	6·32	516	-59·7	0·41	20.3c
10:35	3·502	5·3 L	6·32	519	-66·9	0·38	21.0°c
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

Screen interval : 1·2 - 4·2

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) Stick up doesn't match with dp report

Monument, water is dirty No odour, - IP was clean. NO Sheen

RECHARGE BEHAVIOUR: FAST RECHARGING [ ] SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) [ ]

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE X	TRIPLICATE X
SAMPLE ID: BH106	SAMPLE ID:	SAMPLE ID:
SAMPLE TIME:	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PFAS	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

GROUNDWATER SAMPLE LOG

Geo-Logix

WELL ID: BH107

PROJECT INFORMATION

PROJECT NUMBER:	2301034g	**INITIALS:**	HM
DATE:	23·10·25	**WEATHER:**	Sunny

SAMPLING METHOD: LOW FLOW: ☒ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	2·254	**TOTAL DEPTH (mBTOC):**	4·941	**STICK-UP (m):**	0.997
DEPTH TO PSH (mBTOC):	–	**THICKNESS OF PSH (m):**	–	**TIME:**	~~29/10/24~~ 11:10am

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
11:30	2.559	1 L	6.06	277.9	-40.3	0.02	17°c
11:35	2.875	3.1 L	6.02	269.1	-48.5	0.07	17.1C
11:40	3.251	5.2L	5.92	261.7	-53.0	0.61	17°C.
11:43	3.421	6 L	5.93	270.9	-56.3	0.56	17°C.
11:46	3.615	7 L	5.95	278.0	-57.6	0.53	17 °C.
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) DP Stick up data is 0.489, it is wrong

No odour, water is brown, No Sheen,

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: BH107	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 11.50 am	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 1	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: BH101H

PROJECT INFORMATION

PROJECT NUMBER:	230103497	**INITIALS:**	HM/PDP
DATE:	23·10·25	**WEATHER:**	Sunny

SAMPLING METHOD: LOW FLOW: ☒ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	2·957	**TOTAL DEPTH (mBTOC):**	5.377	**STICK-UP (m):**	0.940
DEPTH TO PSH (mBTOC):	–	**THICKNESS OF PSH (m):**	–	**TIME:**	12:20 pm

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
12:25	2.225	1L	6·48	1135	-24·7	0·49	17·2°C
12:30	2·556	3L	6·42	1113	-27·2	0·66	17·4°C
12:35	2·964	5·5L	6·33	1109	-33·1	0·74	17·7°C
12:40	3·191	6·8L	6·32	1068	-37·5	0·62	17·7°C
12:45	3·542	8L	6·35	1071	-42·8	0·57	17·5°C
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) Stick up doesn't match with DP/Screen: 1·5–4·5

clear no odour | no sheen.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: BH101H	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 12:45 pm	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 1	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: MW10

PROJECT INFORMATION

PROJECT NUMBER:	2301034g.	**INITIALS:**	PDP - HN.
DATE:	28-10-25.	**WEATHER:**	cloudy
SAMPLING METHOD:	LOW FLOW: ☒	HYDRASLEEVE: ☐	BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	6.771.	**TOTAL DEPTH (mBTOC):**	6.982.	**STICK-UP (m):**	–
DEPTH TO PSH (mBTOC):	–	**THICKNESS OF PSH (m):**	–	**TIME:**	10:00 AM

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
10:10	6.901	<200mL	5.54	903	-83.6	9.24	18.2
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) no odour. MW0 (SANDY clay) light brown. couldn't sample. NOT ENOUGHT WATER. NO odour. No sheen

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS). ☐

No recharge in 30'. close. Well. TRY LATER.

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: ~~MW10~~	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME:	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: ~~4~~ 0	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: ~~PFAS.~~	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: ~~XXXX~~ BH101i

PROJECT INFORMATION

PROJECT NUMBER: 230184g.	INITIALS: PDP - HN
DATE: 28-10-25	WEATHER: Cloudy

SAMPLING METHOD: LOW FLOW: ☒ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC): 1.011.	TOTAL DEPTH (mBTOC): 1.430.	STICK-UP (m): —
DEPTH TO PSH (mBTOC): —	THICKNESS OF PSH (m): —	TIME: 11:00.

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
11:05	1.255	0.7	6.24	2150.	-194.3	0.45	19.7
11:10	1.290	0.8	6.32	2162	-195.6	1.36.	19.6
11:15	1.3~~0~~05	~~XX~~ TAKE SAMPLE AS NOT ENOUGHT WATER ON WELL TO CONTINUE PUMPING OUT WATER.					
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) Gatic. Round base, sounds like concrete.
maybe decomm? take sample. NO recharge after 15 min.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING) ~~1.3~~ 1.305.

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: BH101i	SAMPLE ID:	SAMPLE ID:
SAMPLE TIME: 11:15.	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PFAS.	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: BH111

PROJECT INFORMATION

PROJECT NUMBER: .	INITIALS: HM/PDP
DATE: 28·10·25	WEATHER:

SAMPLING METHOD: LOW FLOW: ☒ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC): 1.109	TOTAL DEPTH (mBTOC): 4.986	STICK-UP (m): –
DEPTH TO PSH (mBTOC): –	THICKNESS OF PSH (m): –	TIME: 11·40am

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
11:45	2.065	.2	6.73	1413	-168.3	1.29	18.4
11:50	2.0[illegible]70	2.8.	6.72	1416	-171.7	1.35	18.4
11:55.	2.0[illegible]70	3	6.72	1410	-174.0	1.34	18.5.
12:00	2.485	4.5.	6.81	1411	-175.8	1.68	18.8
12:05.	2.486.	5L	6.82.	1410	-176.2	1.69	18.8
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) soft base. cloudy. no odour no sheen.

Conc. "fast recharge" if compared w/ other wells.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☒

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING) 2.486

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: BH111	SAMPLE ID: DW3	SAMPLE ID: TW3
SAMPLE TIME: 12:05	SAMPLE TIME: 12:10	SAMPLE TIME: 12:15
NO. CONTAINERS: 1	NO. CONTAINERS 1	NO. CONTAINERS 1
ANALYSIS: PFAS.	ANALYSIS: PFAS	ANALYSIS: PFAS.

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: BH101g.

PROJECT INFORMATION

PROJECT NUMBER:	2301034g	**INITIALS:**	PDP-YM.
DATE:	28-10-25	**WEATHER:**	RAINY.
SAMPLING METHOD:	LOW FLOW: ☒	HYDRASLEEVE: ☐	BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	2.503	**TOTAL DEPTH (mBTOC):**	5.505.	**STICK-UP (m):**	0.96
DEPTH TO PSH (mBTOC):	—	**THICKNESS OF PSH (m):**	—	**TIME:**	12:50 pm

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
1:00	3.047	1.9 L	6.72	1001	-150.4	4.26	17.
1:05	3.291	3L	6.67	397	-147.2	4.26	17.
1:10	3.552	4L	6.70.	962	-145.8	4.24	17.1
1:15.	3.715.	4.6	6.75	899	-145.0	4.28	17.2
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) TUBING INside, need to TO remove it. (1°). TO WE use a wire (without sleeve).

then measure GW level / start sampling. monument.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING) 3.715.

ORIGINAL	DUPLICATE X	TRIPLICATE X
SAMPLE ID: 1:15 BH101g.	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 1	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: PCAS.	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS:	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

GROUNDWATER SAMPLE LOG

WELL ID: MW03-S

Geo-Logix

PROJECT INFORMATION

PROJECT NUMBER:	**INITIALS:** HM
DATE: 28·10·25	**WEATHER:** Cloudy

SAMPLING METHOD: LOW FLOW: ☑ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC): 1.710	**TOTAL DEPTH (mBTOC):** 5.912	**STICK-UP (m):**
DEPTH TO PSH (mBTOC): —	**THICKNESS OF PSH (m):** —	**TIME:** 9:45

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
9:50	2.221	1	5.76	426.5	-97.1	1.75	17.8
9:55	2.694	2	5.84	438.6	-87.4	8.12	17.5
10:00	3.072	2.6	5.82	435.9	-110.2	8.26	17.6
10:05	3.065	3.5	5.83	437.1	-117.3	7.96	17.5
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) NO odour - no sheen Gotic empty bolt cover with soft box.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING) 3.065

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: MW03-S	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 10:10 am	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 1	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

GROUNDWATER SAMPLE LOG

Geo-Logix

WELL ID: MW02

PROJECT INFORMATION

PROJECT NUMBER:		INITIALS:	HM
DATE:	28.10.25	WEATHER:	Cloudy
SAMPLING METHOD:	LOW FLOW: ☑	HYDRASLEEVE: ☐	BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	1.313	TOTAL DEPTH (mBTOC):	2.254	STICK-UP (m):	–
DEPTH TO PSH (mBTOC):	–	THICKNESS OF PSH (m):	–	TIME:	10:35

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
10:45 10:~~45~~	1.568	0.8	6.61	903	-105.2	0.50	17.8
10:50	1.807	1.5	6.66	852	-143.1	0.64	18.0
10:55	2031	2.8	6.67	850	-149.6	0.58	17.7
11:00	2.134	3.7	6.67	854	-151.4	0.61	17.6
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) Gotic no odour - no sheen Clear / cloudy

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING) 2.134

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: MW02	SAMPLE ID:	SAMPLE ID:
SAMPLE TIME: 11:00	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PFAS	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: BH105

PROJECT INFORMATION

PROJECT NUMBER:	2301034g	INITIALS:	HM
DATE:	28/10/25	WEATHER:	Cloudy / Rainy

SAMPLING METHOD: LOW FLOW: ☒ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	2.091	TOTAL DEPTH (mBTOC):	4.715	STICK-UP (m):	
DEPTH TO PSH (mBTOC):	-	THICKNESS OF PSH (m):	-	TIME:	12:08 PM

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (MS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
12:15	2.224	0.5	6.91	1252	-13.3	4.19	17.3
12:20	2.364	1.1	6.91	1275	-13.6	4.90	17.2
12:25	2.471	1.6L	6.89	1272	-15.1	3.69	17.2
12:30	2.629	2.8	6.87	1278	-15.9	3.71	17.1
12:35	2.756	3.5	6.87	1280	-15.6	3.75	17.0
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) base is Soft clear, no odour, no sheen

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☒

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING) 2.756

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: BH105	SAMPLE ID:	SAMPLE ID:
SAMPLE TIME: 12:40 PM	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PFAS	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

GROUNDWATER SAMPLE LOG

Geo-Logix

WELL ID: MW11.

PROJECT INFORMATION

PROJECT NUMBER:	2301034g.	INITIALS:	PDP - HM
DATE:	28-10-25	WEATHER:	CLOUDY

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☒

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	2.732.	TOTAL DEPTH (mBTOC):	6.957	STICK-UP (m):	1.07
DEPTH TO PSH (mBTOC):	–	THICKNESS OF PSH (m):	–	TIME:	9:30

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
9:30	–	–	5.77	583.1	-199.8	3.17	16.2
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

called MW11 as not found in "original" location
water depth as MW11.
location as MW11 mapped previous reports.

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) stick up, no monument / no orange gatic cap.
cloudy. no odour, no sheen. IT WAS RAINING before sampling + NO water cap.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

looks like it never has monument / cap. no guide of re presentative sample.

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: MW11	SAMPLE ID:	SAMPLE ID:
SAMPLE TIME: 9:30 am	SAMPLE TIME:	SAMPLE TIME:
NO. CONTAINERS: 1	NO. CONTAINERS	NO. CONTAINERS
ANALYSIS: PFAS.	ANALYSIS:	ANALYSIS:

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER LEVEL GAUGING

SHALLOW WELLS.

PROJECT INFORMATION	
PROJECT:	2301034 g
PROJECT NO:	RICHMOND.
DATE:	27-10-25.
INITIALS:	PDP - HM.

WELL ID	Standing Water Level (mBTOC)	Total Well Depth (mBTOC)	Time	Stick Up (m)
MW03-S	1.708	5.912	9:20	—
MW10	6.761	7.014	9:10	—
MW02	1.299	2.300	13:00	—
MW11 (?)	2.680	6.952	14:00	1.07
BH111	0.979	4.986	12:10	-
BH103	DRY	5.506	10:20	1.05
BH108	2.976	4.275	13:40	0.655
BH107	3.164	4.941	10:25	0.99
BH101h	1.971	5.377	10:40	0.94
BH109	1.872	4.635	13:30	0.96
BH105	2.015	4.715	11:00	0.98
BH101g	—	5.505	—	TUBING INSIDE. COULDNT REMOVE IT. 0.96 STICK UP
BH106	3.062	5.120	10:50	0.97
BH102-S	2.069	5.219	13:50	0.90
BH101	2.379	5.917	12:45	1.075
BH101i	0.964	1.430	12:20	—
BH101e	—	—	—	BROKEN (CATIC/CASING).
GW2	—	—	—	NOT FOUND
BH104	—	—	—	"

Geo-Logix

GROUNDWATER LEVEL GAUGING

Deep Wells.

PROJECT INFORMATION	
PROJECT:	23010349.
PROJECT NO:	RICHMOND
DATE:	27-10-25.
INITIALS:	HM - PDP

(mBTOC).

WELL ID	Standing Water Level (TOTAL WELL DEEP)	SWL (Total Well Depth)	Time	Stick Up (m)
GW128	14.845	12.321	11:15.	
GW129	14.328	12.305	11:25	
GW153	17.028	13.485	13:10	
GW155	15.602	13.345	13:20	
MW03_d	14.879	12.586	9:15	
BH201	20.630	13.507	11:00	

ATTACHMENT B

Certificate of Service and Calibration

Interface Meter

Heron H.Oil

Company Name	WAM Scientific
Office Address	26 Bungarra Crescent, Chipping Norton NSW 2170
Phone Number	+61 405 241 484
Contact Name	William Pak
Instrument	Heron H.Oil Interface Meter (30m)
Serial Number	01-09487
Client Name	Paula Del Prete (Geo-Logix)
Project Number	PO6959
Comments	-

Instrument Check			
Item	Test	Test Passed	Comments
9V Battery	Klein Tools MM300 Multimeter	✓	Battery voltage reading above 7.9V
Battery Box	Check	✓	No damage
Face and Back Plates	Check	✓	No damage
Thumb Screws	Check	✓	Rubber ends intact
Tape Hangar/Protector	Check	✓	No damage
On/Off Button	Operation	✓	Button is functional
Buzzer	Operation	✓	Intermittent tone in H_2O, solid tone in product
LED Signal Light	Operation	✓	LED light functional – green and red
Probe	Operation/Check	✓	Decontaminated, cleaned and tested
Tape	Condition/Check	✓	Decontaminated and cleaned, no damage
Connection	Check	✓	Probe and link connected correctly and tightly
PCB	Operation	✓	Unit is fully functional
Electronics Panel	Orientation	✓	Correctly aligned

Instrument Readings		
Product	Buzzer	LED Light
H_2O	Intermittent	Blinking – Red
Petroleum	Solid	Steady – Red

Declaration
WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The interface meter was decontaminated, cleaned and tested with a mixture of tap water and petrol, shielded from ambient light.

Checked By	William Pak
Calibration Date	09/10/2025
Calibration Due	09/04/2026

WAM SCIENTIFIC
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26 Bungarra Crescent, CHIPPING NORTON NSW 2170
www.wamscientific.com.au

Certificate of Service and Calibration

Water Quality Meter

YSI Professional Plus

Company Name	WAM Scientific
Office Address	26 Bungarra Crescent, Chipping Norton NSW 2170
Phone Number	+61 405 241 484
Contact Name	William Pak
Instrument	YSI Pro Quatro Water Quality Meter w/ 4m Quatro Cable
Serial Number	21A102653
Client Name	Paula Del Prete, Harsha Muthu (Geo-Logix)
Project Number	6949
Comments	-

Instrument Check

Item	Test	Test Passed	Comments
2 x Alkaline C-size Batteries	Klein Tools MM300 Multimeter	✓	Both batteries reading above 2.9V
Battery Saver Function	Operation	✓	Automatically turns off after 60 minutes if idle
Unit Display	Operation	✓	Screen visible, no damage
Keypad	Operation	✓	Responsive, no damage
Connection Port and Cable	Condition/Check	✓	Clean, no damage
Monitor Housing	Condition/Check	✓	No damage
Firmware	Version	✓	4.0.0
pH Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
pH millivolts for pH 7.00	Calibration	✓	pH 7.00 calibration range between 0 mV ± 50 mV
pH millivolts for pH 4.00	Calibration	✓	pH 4 mV range +165 to +180 from 7 buffer mV value
pH slope	Calibration	✓	Range between 55 to 60 mV/pH (ideal value 59 mV)
Response time < 90 seconds	Calibration	✓	Responds to correct value within 90 seconds
ORP Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
ORP Reading	Calibration	✓	Within ± 80 mV of reference Zobell Reading
Response time < 90 seconds	Calibration	✓	Responds to correct value within 90 seconds
Conductivity/Temp Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
Conductivity Cell	Calibration	✓	Conductivity cell constant 5.0 ± 1.0 in GLP file
Clean Sensor Readings	Calibration	✓	Clean sensor reads less than 3 uS/cm in dry air
Dissolved Oxygen Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
DO Cap	Condition/Calibration	✓	1.25 mil PE membrane (yellow membrane)
DO Sensor in Use	Condition	✓	Polarographic DO sensor
DO Sensor Value	Calibration	✓	(min 4.31 uA - max 8.00 uA) Avg 6.15 uA

Instrument Readings

Parameter	Standard Used	Reference No.	Calibration Value	Observed	Actual	Units
Temperature	Centre 370 Thermometer	Room Temp.	19.3	20.3	19.3	°C
pH	pH 4.00	417183	4.01	4.14	4.01	pH
pH	pH 7.00	419528	7.00	7.09	7.00	pH
ORP	Zobell A & B	420448/418958	241.5	264.3	241.5	mV
Conductivity	2760 µs/cm at 25°C	399819	2760	2651	2760	µs/cm
Zero Dissolved O_2	$NaSO_3$ in Distilled H_2O	426184	0.0	3.0	0.0	%
100% Dissolved O_2	100% Air Saturated H_2O	Fresh Air	100.0	112.1	100.0	%

Declaration

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The calibration data supplied was obtained in accordance with manufacturer's specifications using solutions of known values.

Calibrated By	William Pak
Calibration Date	16/10/2025
Calibration Due	16/04/2026

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Certificate of Service and Calibration

Water Quality Meter

YSI Professional Plus

Company Name	WAM Scientific
Office Address	26 Bungarra Crescent, Chipping Norton NSW 2170
Phone Number	+61 405 241 484
Contact Name	William Pak
Instrument	YSI Pro Quatro Water Quality Meter w/ 4m Quatro Cable
Serial Number	21A102653
Client Name	Paula Del Prete, Harsha Muthu (Geo-Logix)
Project Number	PO6959
Comments	-

Instrument Check

Item	Test	Test Passed	Comments
2 x Alkaline C-size Batteries	Klein Tools MM300 Multimeter	✓	Both batteries reading above 2.9V
Battery Saver Function	Operation	✓	Automatically turns off after 60 minutes if idle
Unit Display	Operation	✓	Screen visible, no damage
Keypad	Operation	✓	Responsive, no damage
Connection Port and Cable	Condition/Check	✓	Clean, no damage
Monitor Housing	Condition/Check	✓	No damage
Firmware	Version	✓	4.0.0
pH Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
pH millivolts for pH 7.00	Calibration	✓	pH 7.00 calibration range between 0 mV ± 50 mV
pH millivolts for pH 4.00	Calibration	✓	pH 4 mV range +165 to +180 from 7 buffer mV value
pH slope	Calibration	✓	Range between 55 to 60 mV/pH (ideal value 59 mV)
Response time < 90 seconds	Calibration	✓	Responds to correct value within 90 seconds
ORP Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
ORP Reading	Calibration	✓	Within ± 80 mV of reference Zobell Reading
Response time < 90 seconds	Calibration	✓	Responds to correct value within 90 seconds
Conductivity/Temp Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
Conductivity Cell	Calibration	✓	Conductivity cell constant 5.0 ± 1.0 in GLP file
Clean Sensor Readings	Calibration	✓	Clean sensor reads less than 3 uS/cm in dry air
Dissolved Oxygen Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
DO Cap	Condition/Calibration	✓	1.25 mil PE membrane (yellow membrane)
DO Sensor in Use	Condition	✓	Polarographic DO sensor
DO Sensor Value	Calibration	✓	(min 4.31 uA - max 8.00 uA) Avg 6.15 uA

Instrument Readings

Parameter	Standard Used	Reference No.	Calibration Value	Observed	Actual	Units
Temperature	Centre 370 Thermometer	Room Temp.	19.3	20.3	19.3	°C
pH	pH 4.00	417183	4.01	4.14	4.01	pH
pH	pH 7.00	419528	7.00	7.09	7.00	pH
ORP	Zobell A & B	420448/418958	241.5	264.3	241.5	mV
Conductivity	2760 µs/cm at 25°C	399819	2760	2651	2760	µs/cm
Zero Dissolved O_2	$NaSO_3$ in Distilled H_2O	426184	0.0	3.0	0.0	%
100% Dissolved O_2	100% Air Saturated H_2O	Fresh Air	100.0	112.1	100.0	%

Declaration

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The calibration data supplied was obtained in accordance with manufacturer's specifications using solutions of known values.

Calibrated By	William Pak
Calibration Date	16/10/2025
Calibration Due	16/04/2026

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Certificate of Service and Calibration

Water Quality Meter

YSI Professional Plus

Company Name	WAM Scientific
Office Address	26 Bungarra Crescent, Chipping Norton NSW 2170
Phone Number	+61 405 241 484
Contact Name	William Pak
Instrument	YSI Pro Quatro Water Quality Meter w/ 4m Quatro Cable
Serial Number	23A107177
Client Name	Paula Del Prete, Harsha Muthu (Geo-Logix)
Project Number	6949
Comments	-

Instrument Check

Item	Test	Test Passed	Comments
2 x Alkaline C-size Batteries	Klein Tools MM300 Multimeter	✓	Both batteries reading above 2.9V
Battery Saver Function	Operation	✓	Automatically turns off after 60 minutes if idle
Unit Display	Operation	✓	Screen visible, no damage
Keypad	Operation	✓	Responsive, no damage
Connection Port and Cable	Condition/Check	✓	Clean, no damage
Monitor Housing	Condition/Check	✓	No damage
Firmware	Version	✓	4.0.0
pH Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
pH millivolts for pH 7.00	Calibration	✓	pH 7.00 calibration range between 0 mV ± 50 mV
pH millivolts for pH 4.00	Calibration	✓	pH 4 mV range +165 to +180 from 7 buffer mV value
pH slope	Calibration	✓	Range between 55 to 60 mV/pH (ideal value 59 mV)
Response time < 90 seconds	Calibration	✓	Responds to correct value within 90 seconds
ORP Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
ORP Reading	Calibration	✓	Within ± 80 mV of reference Zobell Reading
Response time < 90 seconds	Calibration	✓	Responds to correct value within 90 seconds
Conductivity/Temp Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
Conductivity Cell	Calibration	✓	Conductivity cell constant 5.0 ± 1.0 in GLP file
Clean Sensor Readings	Calibration	✓	Clean sensor reads less than 3 uS/cm in dry air
Dissolved Oxygen Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
DO Cap	Condition/Calibration	✓	1.25 mil PE membrane (yellow membrane)
DO Sensor in Use	Condition	✓	Polarographic DO sensor
DO Sensor Value	Calibration	✓	(min 4.31 uA - max 8.00 uA) Avg 6.15 uA

Instrument Readings

Parameter	Standard Used	Reference No.	Calibration Value	Observed	Actual	Units
Temperature	Centre 370 Thermometer	Room Temp.	19.7	21.0	19.7	°C
pH	pH 4.00	417183	4.01	4.10	4.01	pH
pH	pH 7.00	419528	7.00	7.06	7.00	pH
ORP	Zobell A & B	420448/418958	240.7	253.7	240.7	mV
Conductivity	2760 µs/cm at 25°C	399819	2760	2974	2760	µs/cm
Zero Dissolved O_2	$NaSO_3$ in Distilled H_2O	426184	0.0	0.2	0.0	%
100% Dissolved O_2	100% Air Saturated H_2O	Fresh Air	100.0	99.8	100.0	%

Declaration

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The calibration data supplied was obtained in accordance with manufacturer's specifications using solutions of known values.

Calibrated By	William Pak
Calibration Date	16/10/2025
Calibration Due	16/04/2026

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Certificate of Service and Calibration

Water Quality Meter

YSI Professional Plus

Company Name	WAM Scientific
Office Address	26 Bungarra Crescent, Chipping Norton NSW 2170
Phone Number	+61 405 241 484
Contact Name	William Pak
Instrument	YSI Pro Quatro Water Quality Meter w/ 4m Quatro Cable
Serial Number	23A107177
Client Name	Paula Del Prete, Harsha Muthu (Geo-Logix)
Project Number	PO6959
Comments	-

Instrument Check

Item	Test	Test Passed	Comments
2 x Alkaline C-size Batteries	Klein Tools MM300 Multimeter	✓	Both batteries reading above 2.9V
Battery Saver Function	Operation	✓	Automatically turns off after 60 minutes if idle
Unit Display	Operation	✓	Screen visible, no damage
Keypad	Operation	✓	Responsive, no damage
Connection Port and Cable	Condition/Check	✓	Clean, no damage
Monitor Housing	Condition/Check	✓	No damage
Firmware	Version	✓	4.0.0
pH Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
pH millivolts for pH 7.00	Calibration	✓	pH 7.00 calibration range between 0 mV ± 50 mV
pH millivolts for pH 4.00	Calibration	✓	pH 4 mV range +165 to +180 from 7 buffer mV value
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Conductivity Cell	Calibration	✓	Conductivity cell constant 5.0 ± 1.0 in GLP file
Clean Sensor Readings	Calibration	✓	Clean sensor reads less than 3 uS/cm in dry air
Dissolved Oxygen Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
DO Cap	Condition/Calibration	✓	1.25 mil PE membrane (yellow membrane)
DO Sensor in Use	Condition	✓	Polarographic DO sensor
DO Sensor Value	Calibration	✓	(min 4.31 uA - max 8.00 uA) Avg 6.15 uA

Instrument Readings

Parameter	Standard Used	Reference No.	Calibration Value	Observed	Actual	Units
Temperature	Centre 370 Thermometer	Room Temp.	19.7	21.0	19.7	°C
pH	pH 4.00	417183	4.01	4.10	4.01	pH
pH	pH 7.00	419528	7.00	7.06	7.00	pH
ORP	Zobell A & B	420448/418958	240.7	253.7	240.7	mV
Conductivity	2760 µs/cm at 25°C	399819	2760	2974	2760	µs/cm
Zero Dissolved O_2	$NaSO_3$ in Distilled H_2O	426184	0.0	0.2	0.0	%
100% Dissolved O_2	100% Air Saturated H_2O	Fresh Air	100.0	99.8	100.0	%

Declaration

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Calibrated By	William Pak
Calibration Date	16/10/2025
Calibration Due	16/04/2026

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ATTACHMENT C

04/02/2026

RE: Monitoring Well Locations Richmond

Hi Paula,

Please find below the table of values for the monitoring wells at WSU Richmond measurements are in AHD and MGA2020,

Name	Easting	Northing	RL	Desc.
BH101i	290862.914	6278231	23.146	LID
BH101i	290862.84	6278231	23.056	PIPE
BH101	290897.307	6278190	24.662	PIPE
BH101	290897.222	6278190	23.611	ESL
BH201	290902.035	6278186	23.514	ESL
BH201	290902.426	6278186	24.175	PIPE
BH101G	290921.897	6278159	23.143	ESL
BH101G	290921.892	6278159	23.886	PIPE
BH101H	290836.414	6278202	23.576	ESL
BH101H	290836.491	6278202	24.531	PIPE
BH102_s	290655.734	6278315	23.425	ESL
BH102_s	290655.651	6278315	24.313	PIPE
BH103	290715.749	6278050	23.04	ESL
BH103	290715.509	6278050	24.053	PIPE
BH105	290987.101	6278190	22.823	ESL
BH105	290987.048	6278190	23.598	PIPE
BH106	290982.964	6278115	22.85	ESL
BH106	290983.139	6278115	23.805	PIPE
BH107	290792.521	6278132	23.221	ESL
BH107	290792.468	6278132	24.237	PIPE
BH108	290615.582	6278161	23.193	ESL
BH108	290615.619	6278161	23.88	PIPE
BH109	290751.148	6278274	23.274	ESL
BH109	290751.071	6278274	24.248	PIPE
BH111	290901.034	6278251	23.277	LID
BH111	290901.076	6278251	23.191	PIPE
MW11	290633.64	6278371	23.601	ESL
MW11	290633.423	6278371	24.382	PIPE

Best regards
Glen Watson
Director
Onsite Surveying Pty Ltd

ATTACHMENT D

Geo-Logix P/L
Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

NATA Accredited
Accreditation Number 1261
Site Number 18217

Accredited for compliance with ISO/IEC 17025 – Testing
NATA is a signatory to the ILAC Mutual Recognition Arrangement for the mutual recognition of the equivalence of testing, medical testing, calibration, inspection, proficiency testing scheme providers and reference materials producers reports and certificates.

Attention: **Paula Del Prete**

Report **1283145-W-V2**
Project name RICHMOND
Project ID 2301034G
Received Date Oct 22, 2025

Client Sample ID			**BH102**	**BH108**	**BH109**	**GW153**
Sample Matrix			**Water**	**Water**	**Water**	**Water**
Eurofins Sample No.			**S25-Oc0067273**	**S25-Oc0067274**	**S25-Oc0067275**	**S25-Oc0067276**
Date Sampled			**Oct 21, 2025**	**Oct 21, 2025**	**Oct 21, 2025**	**Oct 21, 2025**
Test/Reference	LOR	Unit				
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	0.05	< 0.05	< 0.05	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.02	< 0.01	0.01	< 0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	101	100	100	95
13C5-PFPeA (surr.)	1	%	105	97	100	100
13C5-PFHxA (surr.)	1	%	118	103	110	106
13C4-PFHpA (surr.)	1	%	114	100	104	94
13C8-PFOA (surr.)	1	%	111	111	106	103
13C5-PFNA (surr.)	1	%	102	99	98	98
13C6-PFDA (surr.)	1	%	103	98	103	95
13C2-PFUnDA (surr.)	1	%	113	97	98	89
13C2-PFDoDA (surr.)	1	%	106	95	90	88
13C2-PFTeDA (surr.)	1	%	106	101	97	88
Perfluoroalkyl sulfonamido substances						
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	105	90	89	87
D3-N-MeFOSA (surr.)	1	%	90	91	96	84

Client Sample ID			BH102	BH108	BH109	GW153
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0067273	S25-Oc0067274	S25-Oc0067275	S25-Oc0067276
Date Sampled			Oct 21, 2025	Oct 21, 2025	Oct 21, 2025	Oct 21, 2025
Test/Reference	LOR	Unit				
Perfluoroalkyl sulfonamido substances						
D5-N-EtFOSA (surr.)	1	%	100	96	96	83
D7-N-MeFOSE (surr.)	1	%	91	88	89	82
D9-N-EtFOSE (surr.)	1	%	96	87	85	81
D5-N-EtFOSAA (surr.)	1	%	101	87	83	88
D3-N-MeFOSAA (surr.)	1	%	102	96	87	91
Perfluoroalkyl sulfonic acids (PFSAs)						
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	0.03	< 0.01	0.02	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	[N09]0.03	< 0.01	[N09]0.01	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]0.40	[N09]0.01	[N09]0.14	< 0.01
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	[N09]0.02	< 0.01	< 0.01	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	[N09]0.15	< 0.01	[N09]0.02	< 0.01
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	108	109	111	114
18O2-PFHxS (surr.)	1	%	100	100	109	104
13C8-PFOS (surr.)	1	%	105	95	104	96
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)						
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	107	103	101	94
13C2-6:2 FTSA (surr.)	1	%	101	95	92	85
13C2-8:2 FTSA (surr.)	1	%	97	80	88	92
13C2-10:2 FTSA (surr.)	1	%	79	97	79	84
PFASs Summations						
Sum (PFHxS + PFOS)*	0.01	ug/L	0.55	0.01	0.16	< 0.01
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	0.65	< 0.05	0.19	< 0.05
Sum of PFASs (n=30)*	0.1	ug/L	0.7	< 0.1	0.2	< 0.1

Client Sample ID			GW155	MW03-D	DW1	R1
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0067277	S25-Oc0067278	S25-Oc0067279	S25-Oc0067280
Date Sampled			Oct 21, 2025	Oct 21, 2025	Oct 21, 2025	Oct 21, 2025
Test/Reference	LOR	Unit				
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	< 0.01	0.03	< 0.01	< 0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01

Client Sample ID			GW155	MW03-D	DW1	R1
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0067277	S25-Oc0067278	S25-Oc0067279	S25-Oc0067280
Date Sampled			Oct 21, 2025	Oct 21, 2025	Oct 21, 2025	Oct 21, 2025
Test/Reference	LOR	Unit				
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	95	101	99	99
13C5-PFPeA (surr.)	1	%	103	116	104	101
13C5-PFHxA (surr.)	1	%	110	106	112	105
13C4-PFHpA (surr.)	1	%	99	103	102	114
13C8-PFOA (surr.)	1	%	98	113	106	109
13C5-PFNA (surr.)	1	%	98	101	100	96
13C6-PFDA (surr.)	1	%	96	99	93	104
13C2-PFUnDA (surr.)	1	%	95	101	85	103
13C2-PFDoDA (surr.)	1	%	99	99	73	98
13C2-PFTeDA (surr.)	1	%	114	106	59	86
Perfluoroalkyl sulfonamido substances						
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	91	90	93	97
D3-N-MeFOSA (surr.)	1	%	91	88	76	91
D5-N-EtFOSA (surr.)	1	%	97	95	85	91
D7-N-MeFOSE (surr.)	1	%	87	91	84	93
D9-N-EtFOSE (surr.)	1	%	91	93	85	88
D5-N-EtFOSAA (surr.)	1	%	87	98	85	97
D3-N-MeFOSAA (surr.)	1	%	88	97	87	97
Perfluoroalkyl sulfonic acids (PFSAs)						
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	< 0.01	0.12	< 0.01	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	< 0.01	0.04	< 0.01	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	< 0.01	[N09]0.09	< 0.01	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	< 0.01	[N09]0.73	[N09]0.01	< 0.01
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01	[N09]0.01	< 0.01	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	< 0.01	[N09]0.07	< 0.01	< 0.01
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	112	114	115	111
18O2-PFHxS (surr.)	1	%	99	103	100	105
13C8-PFOS (surr.)	1	%	101	103	85	99

Client Sample ID			GW155	MW03-D	DW1	R1
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0067277	S25-Oc0067278	S25-Oc0067279	S25-Oc0067280
Date Sampled			Oct 21, 2025	Oct 21, 2025	Oct 21, 2025	Oct 21, 2025
Test/Reference	LOR	Unit				
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)						
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	89	93	102	91
13C2-6:2 FTSA (surr.)	1	%	106	106	85	109
13C2-8:2 FTSA (surr.)	1	%	95	85	80	83
13C2-10:2 FTSA (surr.)	1	%	93	93	92	87
PFASs Summations						
Sum (PFHxS + PFOS)*	0.01	ug/L	< 0.01	0.8	0.01	< 0.01
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	< 0.05	0.95	< 0.05	< 0.05
Sum of PFASs (n=30)*	0.1	ug/L	< 0.1	1.1	< 0.1	< 0.1

Client Sample ID			BLANK 1
Sample Matrix			Water
Eurofins Sample No.			S25-Oc0067281
Date Sampled			Oct 21, 2025
Test/Reference	LOR	Unit	
Perfluoroalkyl carboxylic acids (PFCAs)			
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	< 0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01
13C4-PFBA (surr.)	1	%	97
13C5-PFPeA (surr.)	1	%	105
13C5-PFHxA (surr.)	1	%	116
13C4-PFHpA (surr.)	1	%	104
13C8-PFOA (surr.)	1	%	104
13C5-PFNA (surr.)	1	%	101
13C6-PFDA (surr.)	1	%	106
13C2-PFUnDA (surr.)	1	%	100
13C2-PFDoDA (surr.)	1	%	95
13C2-PFTeDA (surr.)	1	%	85

Client Sample ID			BLANK 1
Sample Matrix			Water
Eurofins Sample No.			S25-Oc0067281
Date Sampled			Oct 21, 2025
Test/Reference	LOR	Unit	
Perfluoroalkyl sulfonamido substances			
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05
13C8-FOSA (surr.)	1	%	92
D3-N-MeFOSA (surr.)	1	%	100
D5-N-EtFOSA (surr.)	1	%	93
D7-N-MeFOSE (surr.)	1	%	93
D9-N-EtFOSE (surr.)	1	%	92
D5-N-EtFOSAA (surr.)	1	%	97
D3-N-MeFOSAA (surr.)	1	%	99
Perfluoroalkyl sulfonic acids (PFSAs)			
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	< 0.01
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	< 0.01
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01
13C3-PFBS (surr.)	1	%	105
18O2-PFHxS (surr.)	1	%	104
13C8-PFOS (surr.)	1	%	96
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01
13C2-4:2 FTSA (surr.)	1	%	90
13C2-6:2 FTSA (surr.)	1	%	112
13C2-8:2 FTSA (surr.)	1	%	97
13C2-10:2 FTSA (surr.)	1	%	86
PFASs Summations			
Sum (PFHxS + PFOS)*	0.01	ug/L	< 0.01
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	< 0.05
Sum of PFASs (n=30)*	0.1	ug/L	< 0.1

Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description	Testing Site	Extracted	Holding Time
Per- and Polyfluoroalkyl Substances (PFASs)			
Perfluoroalkyl carboxylic acids (PFCAs)	Sydney	Oct 27, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonamido substances	Sydney	Oct 27, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonic acids (PFSAs)	Sydney	Oct 27, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)	Sydney	Oct 27, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			

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Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: RICHMOUND
Project ID: 2301034G

Order No.: PO6956
Report #: 1283145
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Oct 22, 2025 2:46 PM
Due: Oct 29, 2025
Priority: 5 Day
Contact Name: Paula Del Prete

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail						Per- and Polyfluoroalkyl Substances (PFASs)
Sydney Laboratory - NATA # 1261 Site # 18217						X
External Laboratory						
No	**Sample ID**	**Sample Date**	**Sampling Time**	**Matrix**	**LAB ID**	
1	BH102	Oct 21, 2025		Water	S25-Oc0067273	X
2	BH108	Oct 21, 2025		Water	S25-Oc0067274	X
3	BH109	Oct 21, 2025		Water	S25-Oc0067275	X
4	GW153	Oct 21, 2025		Water	S25-Oc0067276	X
5	GW155	Oct 21, 2025		Water	S25-Oc0067277	X
6	MW03-D	Oct 21, 2025		Water	S25-Oc0067278	X
7	W1	Oct 21, 2025		Water	S25-Oc0067279	X
8	R1	Oct 21, 2025		Water	S25-Oc0067280	X
9	BLANK 1	Oct 21, 2025		Water	S25-Oc0067281	X
Test Counts						9

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Internal Quality Control Review and Glossary

General

1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follow guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013. They are included in this QC report where applicable. Additional QC data may be available on request.
2. Unless otherwise stated, all soil/sediment/solid results are reported on a dry weight basis.
3. Unless otherwise stated, all biota/food results are reported on a wet weight basis on the edible portion.
4. For CEC results where the sample's origin is unknown or environmentally contaminated, the results should be used advisedly.
5. Actual LORs are matrix dependent. Quoted LORs may be raised where sample extracts are diluted due to interferences.
6. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds where annotated.
7. SVOC analysis on waters is performed on homogenised, unfiltered samples unless noted otherwise.
8. Samples were analysed on an 'as received' basis.
9. Information identified in this report with **blue** colour indicates data provided by customers that may have an impact on the results.
10. This report replaces any interim results previously issued.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours before sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the sampling date; therefore, compliance with these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether, the holding time is seven days; however, for all other VOCs, such as BTEX or C6-10 TRH, the holding time is 14 days.

Units

mg/kg: milligrams per kilogram	**mg/L:** milligrams per litre	**ppm:** parts per million
µg/L: micrograms per litre	**ppb:** parts per billion	**%:** Percentage
org/100 mL: Organisms per 100 millilitres	**NTU:** Nephelometric Turbidity Units	**MPN/100 mL:** Most Probable Number of organisms per 100 millilitres
CFU: Colony Forming Unit	**Colour:** Pt-Co Units (CU)	

Terms

APHA	American Public Health Association
CEC	Cation Exchange Capacity
COC	Chain of Custody
CP	Client Parent - QC was performed on samples pertaining to this report
CRM	Certified Reference Material (ISO17034) - reported as percent recovery.
Dry	Where moisture has been determined on a solid sample, the result is expressed on a dry weight basis.
Duplicate	A second piece of analysis from the same sample and reported in the same units as the result to show comparison.
LOR	Limit of Reporting.
LCS	Laboratory Control Sample - reported as percent recovery.
Method Blank	In the case of solid samples, these are performed on laboratory-certified clean sands and in the case of water samples, these are performed on de-ionised water.
NCP	Non-Client Parent - QC performed on samples not pertaining to this report, QC represents the sequence or batch that client samples were analysed within.
RPD	Relative Percent Difference between two Duplicate pieces of analysis.
SPIKE	Addition of the analyte to the sample and reported as percentage recovery.
SRA	Sample Receipt Advice
Surr - Surrogate	The addition of a similar compound to the analyte target is reported as percentage recovery. See below for acceptance criteria.
TBTO	Tributyltin oxide (*bis*-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment; however, free tributyltin was measured, and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits.
TCLP	Toxicity Characteristic Leaching Procedure
TEQ	Toxic Equivalency Quotient or Total Equivalence
QSM	US Department of Defense Quality Systems Manual Version 6.0
US EPA	United States Environmental Protection Agency
WA DWER	Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should only be used as a guide and may be different when site-specific Sampling Analysis and Quality Plan (SAQP) have been implemented.

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is ≤30%; however, the following acceptance guidelines are equally applicable:

Results <10 times the LOR:	No Limit
Results between 10-20 times the LOR:	RPD must lie between 0-50%
Results >20 times the LOR:	RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range, not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 – 150%, VOC recoveries 50 – 150%

PFAS field samples containing surrogate recoveries above the QC limit designated in QSM 6.0, where no positive PFAS results have been reported or reviewed, and no data was affected.

QC Data General Comments

1. Where a result is reported as less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown are not data from your samples.
3. pH and Free Chlorine analysed in the laboratory - Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time.Analysis will begin as soon as possible after sample receipt.
4. Recovery Data (Spikes & Surrogates) - where chromatographic interference does not allow the determination of recovery, the term "INT" appears against that analyte.
5. For Matrix Spikes and LCS results, a dash "-" in the report means that the specific analyte was not added to the QC sample.
6. Duplicate RPDs are calculated from raw analytical data; thus, it is possible to have two sets of data.

Quality Control Results

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Method Blank							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	ug/L	< 0.05			0.05	Pass	
Perfluoropentanoic acid (PFPeA)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanoic acid (PFHxA)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanoic acid (PFHpA)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanoic acid (PFOA)	ug/L	< 0.01			0.01	Pass	
Perfluorononanoic acid (PFNA)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanoic acid (PFDA)	ug/L	< 0.01			0.01	Pass	
Perfluoroundecanoic acid (PFUnDA)	ug/L	< 0.01			0.01	Pass	
Perfluorododecanoic acid (PFDoDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotridecanoic acid (PFTrDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotetradecanoic acid (PFTeDA)	ug/L	< 0.01			0.01	Pass	
Method Blank							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	ug/L	< 0.05			0.05	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	ug/L	< 0.05			0.05	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	ug/L	< 0.05			0.05	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	ug/L	< 0.05			0.05	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	ug/L	< 0.05			0.05	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	ug/L	< 0.05			0.05	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	ug/L	< 0.05			0.05	Pass	
Method Blank							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	ug/L	< 0.01			0.01	Pass	
Perfluorononanesulfonic acid (PFNS)	ug/L	< 0.01			0.01	Pass	
Perfluoropropanesulfonic acid (PFPrS)	ug/L	< 0.01			0.01	Pass	
Perfluoropentanesulfonic acid (PFPeS)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanesulfonic acid (PFHxS)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanesulfonic acid (PFOS)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanesulfonic acid (PFDS)	ug/L	< 0.01			0.01	Pass	
Method Blank							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	ug/L	< 0.05			0.05	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	ug/L	< 0.01			0.01	Pass	
LCS - % Recovery							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	%	89			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	%	96			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	%	103			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	%	97			50-150	Pass	
Perfluorooctanoic acid (PFOA)	%	97			50-150	Pass	
Perfluorononanoic acid (PFNA)	%	92			50-150	Pass	
Perfluorodecanoic acid (PFDA)	%	103			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	%	97			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	%	109			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	%	113			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	%	100			50-150	Pass	

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
LCS - % Recovery							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	%	92			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	%	100			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	%	98			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	%	94			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	%	99			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	%	88			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	%	102			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	%	86			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	%	102			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	%	79			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	%	77			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	%	88			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	%	105			50-150	Pass	
Perfluorooctanesulfonic acid (PFOS)	%	98			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	%	96			50-150	Pass	
LCS - % Recovery							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	%	95			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	%	94			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	%	99			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	%	98			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Spike - % Recovery									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1					
Perfluorobutanoic acid (PFBA)	S25-Oc0058537	NCP	%	98			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	S25-Oc0058537	NCP	%	116			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	S25-Oc0058537	NCP	%	96			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	S25-Oc0058537	NCP	%	104			50-150	Pass	
Perfluorooctanoic acid (PFOA)	S25-Oc0058537	NCP	%	105			50-150	Pass	
Perfluorononanoic acid (PFNA)	S25-Oc0058537	NCP	%	96			50-150	Pass	
Perfluorodecanoic acid (PFDA)	S25-Oc0058537	NCP	%	99			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	S25-Oc0058537	NCP	%	98			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	S25-Oc0058537	NCP	%	103			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	S25-Oc0058537	NCP	%	103			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S25-Oc0058537	NCP	%	101			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonamido substances				Result 1					
Perfluorooctane sulfonamide (FOSA)	S25-Oc0058537	NCP	%	114			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S25-Oc0058537	NCP	%	113			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S25-Oc0058537	NCP	%	90			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S25-Oc0058537	NCP	%	100			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S25-Oc0058537	NCP	%	101			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S25-Oc0058537	NCP	%	100			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S25-Oc0058537	NCP	%	108			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1					
Perfluorobutanesulfonic acid (PFBS)	S25-Oc0058537	NCP	%	96			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	S25-Oc0058537	NCP	%	103			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S25-Oc0058537	NCP	%	93			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S25-Oc0058537	NCP	%	77			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S25-Oc0058537	NCP	%	93			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S25-Oc0058537	NCP	%	103			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	S25-Oc0058537	NCP	%	104			50-150	Pass	
Spike - % Recovery									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1					
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S25-Oc0058537	NCP	%	111			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S25-Oc0058537	NCP	%	106			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S25-Oc0058537	NCP	%	101			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S25-Oc0058537	NCP	%	94			50-150	Pass	
Test	**Lab Sample ID**	**QA Source**	**Units**	**Result 1**			**Acceptance Limits**	**Pass Limits**	**Qualifying Code**
Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluorobutanoic acid (PFBA)	S25-Oc0062941	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Perfluoropentanoic acid (PFPeA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanoic acid (PFHxA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroheptanoic acid (PFHpA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanoic acid (PFOA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanoic acid (PFNA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanoic acid (PFDA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroundecanoic acid (PFUnDA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorododecanoic acid (PFDoDA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotridecanoic acid (PFTrDA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
Perfluorooctane sulfonamide (FOSA)	S25-Oc0062941	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S25-Oc0062941	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S25-Oc0062941	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S25-Oc0062941	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S25-Oc0062941	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S25-Oc0062941	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S25-Oc0062941	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1	Result 2	RPD			
Perfluorobutanesulfonic acid (PFBS)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanesulfonic acid (PFNS)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanesulfonic acid (PFOS)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanesulfonic acid (PFDS)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1	Result 2	RPD			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S25-Oc0062941	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S25-Oc0062941	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Comments

Report 1283145-W-V2 (amendment to report 1283145-W) has been issued with corrected project name from (RICHMOUND) to (RICHMOND) as per COC.

Sample Integrity

Custody Seals Intact (if used)	N/A
Attempt to Chill was evident	Yes
Sample correctly preserved	Yes
Appropriate sample containers have been used	Yes
Sample containers for volatile analysis received with minimal headspace	Yes
Samples received within HoldingTime	Yes
Some samples have been subcontracted	No

Qualifier Codes/Comments

Code	Description
N09	Quantification of linear and branched isomers has been conducted as a single total response using the relative response factor for the corresponding linear/branched standard.
N11	Isotope dilution is used for calibration of each native compound for which an exact labelled analogue is available (Isotope Dilution Quantitation). The isotopically labelled analogues allow identification and recovery correction of the concentration of the associated native PFAS compounds.
N15	Where the native PFAS compound does not have labelled analogue then the quantification is made using the Extracted Internal Standard Analyte with the closest retention time to the analyte and no recovery correction has been made (Internal Standard Quantitation).

Authorised by:

Nileshni Goundar	Analytical Services Manager
Roopesh Rangarajan	Senior Analyst-PFAS



Glenn Jackson
Managing Director

Final Report – this report replaces any previously issued Report

- Indicates Not Requested

* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here.

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eurofins | Environment Testing

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NATA# 2377
Site# 2370 & 2554

Sample Receipt Advice

Company name: Geo-Logix P/L
Contact name: Paula Del Prete
Project name: RICHMOUND
Project ID: 2301034G
Turnaround time: 5 Day
Date/Time received Oct 22, 2025 2:46 PM
Eurofins reference 1283145

Sample Information

✓ A detailed list of analytes logged into our LIMS, is included in the attached summary table.

✓ Sample Temperature of chilled sample on the batch as recorded by Eurofins Sample Receipt : 12.3 degrees Celsius.

✓ All samples have been received as described on the above COC.

✓ COC has been completed correctly.

✓ Attempt to chill was evident.

✓ Appropriately preserved sample containers have been used.

✓ All samples were received in good condition.

✓ Samples have been provided with adequate time to commence analysis in accordance with the relevant holding times.

✓ Appropriate sample containers have been used.

✓ Sample containers for volatile analysis received with zero headspace.

X Split sample sent to requested external lab.

X Some samples have been subcontracted.

N/A Custody Seals intact (if used).

Notes

Contact

If you have any questions with respect to these samples, please contact your Analytical Services Manager:

Bonnie Pu on phone : or by email: Bonnie.Pu@eurofinsanz.com

Results will be delivered electronically via email to Paula Del Prete - pdelprete@geo-logix.com.au.

Note: A copy of these results will also be delivered to the general Geo-Logix P/L email address.

Geo-Logix Pty Ltd
Building Q2, Level 3
2309/4 Daydream St
Warriewood, NSW 2102
ABN: 86 116 892 936
P: (02) 9979 1722
F: (02) 9979 1222

CHAIN OF CUSTODY

Page 1 of 1

Project Manager: Paula Del Prete, Harsha Muthu
Purchase Order No: PO6956.
Contact email: pdelprete@geo-logix.com.au / hmuthu@geo-logix.com.au
Quote Reference: ~
Project Name: Richmond
Send Invoice to: accounts@geo-logix.com.au
Project Number: 2301034g Date Submitted: 22-10-25
TAT required: STD

ANALYSIS REQUIRED

Lab ID	Sample ID	Date	Matrix: soil	water	air	paint, filters	other	Comments	COMPOSITE	TRH - C6 - C10	TRH - C10 - C40	VOCs	BTEXN	PAHs	PCBs	OCPs	OPPs	Phenols	Metals - M8	Metals - Lead	Metals - Specify **	TCLP	Asbestos (ID only)	Asbestos (WA DOH)	Foreign Materials	Conductivity (EC)	pH	PFAS - 30 compound		Hold	SUITE
	BH102	21-10-25		X				PFAS bottle																				X		
	BH108	"						"																				X		
	BH109	"						"																				X		
	GW153	"						"																				X		
	GW155.	"						"																				X		
	MW03-d	"						"																				X		
	DW1	"						"																				X		
	R1	"		↓				"																				X		
	Blank 1	"		X				"																				X		

Eurofins MGT Suite Codes

B1	TRH/BTEXN
B1A	TRH/MAH
B2	TRH/BTEXN/Pb
B2A	TRH/MAH/Pb
B3	PAH/Phenols
B4	TRH/BTEXN/PAH
B4A	TRH/BTEXN/PAH/Phenols
B5	TRH/BTEXN/M7
B6	TRH/BTEXN/M8
B7	TRH/BTEXN/PAH/M8
B7A	TRH/BTEXN/PAH/Phenols/M8
B8	TRH/VOC/PAH/M8
B9	TRH/BTEXN/PAH/OCP/M8
B10	TRH/BTEXN/PAH/OCP/OPP/M8
B11	Na/K/Ca/Mg/Cl/SO_4/CO_3/HCO_3/NH_3/NO_3
B11A	B11/Alkalinity
B11B	B11/EC/TDS
B12	TRH/BTEXN/Oxygenates/Ethanol
B12A	TRH/BTEXN/Oxygenates
B13	OCP/PCB
B14	OCP/OPP
B15	OCP/OPP/PCB
B16	TDS/SO_4/CH_4/Alk/BOD/COD/HPC/CUB
B17	SO_4/NO_3/Fe++/HPC/CUB
B18	Cl-/SO_4/pH
B19	N/P/K
B20	CEC/%ESP/Ca/Mg/Na/K

Metals(circle)** As, Cd, Cr, Cu, Ni, Pb, Zn, Hg, Cr^{6+}, Cr^{3+}, Fe^{2+}, Fe^{3+}, Be, B, Al, V, Mn, Fe, Co, Se, Sr, Sn, Mo, Ag, Ba, Tl, Bi, Sb

Chain of Custody

Relinquished by: paula del prete Date/Time: 22/10 Signature: CDP
Received by: Hargreet Singh Date/Time: 22/10 2:46pm Signature:

Received by P 16:25
20/10/25
12.3°C Chill

1293145

Nileshni Goundar

From: Ted Lilly <tlilly@geo-logix.com.au>
Sent: Tuesday, 28 October 2025 3:46 PM
To: Nileshni Goundar
Subject: RE: Eurofins Test Results, Invoice - Report 1283145 : Site RICHMOUND (2301034G)

Unverified Sender: The sender of this email has not been verified. Review the content of the message carefully and verify the identity of the sender before acting on this email: replying, opening attachments or clicking links.

Hi Neena,

Can we get the site name corrected on this one as well.

Should be Richmond.

Cheers,

Ted Lilly | Principal Engineer
Unit 2309/4 Daydream St, Warriewood NSW 2102
T: 02 9979 1722 | M: 0412 708 868 | W: www.geo-logix.com.au

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From: Nileshni Goundar <EET-ELVIS@eurofinsanz.com>
Sent: Tuesday, 28 October 2025 3:41 PM
To: Paula Del Prete <pdelprete@geo-logix.com.au>
Cc: Ted Lilly <tlilly@geo-logix.com.au>; Harsha Muthu <hmuthu@geo-logix.com.au>
Subject: Eurofins Test Results, Invoice - Report 1283145 : Site RICHMOUND (2301034G)

Please find the attached report.

Kind Regards,
Nileshni (Neena) Goundar
Assistant Analytical Services Manager
*Please note my work hours are **2pm-10pm,** anything outside of that please contact your ASM for anything urgent.*

Eurofins Environment Testing Australia Pty Ltd
179 Magowar Road
Girraween, NSW, 2145

Email: Nileshni.Goundar@eurofinsanz.com
Website: www.eurofins.com.au/environmental-testing

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Geo-Logix P/L
Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

NATA Accredited
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Site Number 1254

Accredited for compliance with ISO/IEC 17025 – Testing
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Attention: **Paula Del Prete**

Report **1283157-W-V2**
Project name RICHMOND
Project ID 2301034G
Received Date Oct 22, 2025

Client Sample ID			**TW1**
Sample Matrix			**Water**
Eurofins Sample No.			**S25-Oc0067315**
Date Sampled			**Oct 21, 2025**
Test/Reference	LOR	Unit	
Perfluoroalkyl carboxylic acids (PFCAs)			
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	< 0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01
13C4-PFBA (surr.)	1	%	92
13C5-PFPeA (surr.)	1	%	103
13C5-PFHxA (surr.)	1	%	117
13C4-PFHpA (surr.)	1	%	113
13C8-PFOA (surr.)	1	%	115
13C5-PFNA (surr.)	1	%	112
13C6-PFDA (surr.)	1	%	97
13C2-PFUnDA (surr.)	1	%	91
13C2-PFDoDA (surr.)	1	%	69
13C2-PFTeDA (surr.)	1	%	[J-]34
Perfluoroalkyl sulfonamido substances			
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05
13C8-FOSA (surr.)	1	%	100
D3-N-MeFOSA (surr.)	1	%	65

Client Sample ID			TW1
Sample Matrix			Water
Eurofins Sample No.			S25-Oc0067315
Date Sampled			Oct 21, 2025
Test/Reference	LOR	Unit	
Perfluoroalkyl sulfonamido substances			
D5-N-EtFOSA (surr.)	1	%	59
D7-N-MeFOSE (surr.)	1	%	68
D9-N-EtFOSE (surr.)	1	%	51
D5-N-EtFOSAA (surr.)	1	%	107
D3-N-MeFOSAA (surr.)	1	%	91
Perfluoroalkyl sulfonic acids (PFSAs)			
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	< 0.01
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	< 0.01
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01
13C3-PFBS (surr.)	1	%	120
18O2-PFHxS (surr.)	1	%	107
13C8-PFOS (surr.)	1	%	94
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01
13C2-4:2 FTSA (surr.)	1	%	98
13C2-6:2 FTSA (surr.)	1	%	128
13C2-8:2 FTSA (surr.)	1	%	111
13C2-10:2 FTSA (surr.)	1	%	71
PFASs Summations			
Sum (PFHxS + PFOS)*	0.01	ug/L	< 0.01
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	< 0.05
Sum of PFASs (n=30)*	0.1	ug/L	< 0.1

Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description	Testing Site	Extracted	Holding Time
Per- and Polyfluoroalkyl Substances (PFASs)			
Perfluoroalkyl carboxylic acids (PFCAs)	Melbourne	Oct 24, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonamido substances	Melbourne	Oct 24, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonic acids (PFSAs)	Melbourne	Oct 24, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)	Melbourne	Oct 24, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			

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Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: RICHMOND
Project ID: 2301034G

Order No.: PO6955
Report #: 1283157
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Oct 22, 2025 2:46 PM
Due: Oct 29, 2025
Priority: 5 Day
Contact Name: Paula Del Prete

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail						Per- and Polyfluoroalkyl Substances (PFASs)
Melbourne Laboratory - NATA # 1261 Site # 1254						X
External Laboratory						
No	**Sample ID**	**Sample Date**	**Sampling Time**	**Matrix**	**LAB ID**	
1	TW1	Oct 21, 2025		Water	S25-Oc0067315	X
Test Counts						1

eurofins | Environment Testing

Internal Quality Control Review and Glossary

General

1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follow guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013. They are included in this QC report where applicable. Additional QC data may be available on request.
2. Unless otherwise stated, all soil/sediment/solid results are reported on a dry weight basis.
3. Unless otherwise stated, all biota/food results are reported on a wet weight basis on the edible portion.
4. For CEC results where the sample's origin is unknown or environmentally contaminated, the results should be used advisedly.
5. Actual LORs are matrix dependent. Quoted LORs may be raised where sample extracts are diluted due to interferences.
6. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds where annotated.
7. SVOC analysis on waters is performed on homogenised, unfiltered samples unless noted otherwise.
8. Samples were analysed on an 'as received' basis.
9. Information identified in this report with **blue** colour indicates data provided by customers that may have an impact on the results.
10. This report replaces any interim results previously issued.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours before sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the sampling date; therefore, compliance with these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether, the holding time is seven days; however, for all other VOCs, such as BTEX or C6-10 TRH, the holding time is 14 days.

Units

mg/kg: milligrams per kilogram	**mg/L:** milligrams per litre	**ppm:** parts per million
µg/L: micrograms per litre	**ppb:** parts per billion	**%:** Percentage
org/100 mL: Organisms per 100 millilitres	**NTU:** Nephelometric Turbidity Units	**MPN/100 mL:** Most Probable Number of organisms per 100 millilitres
CFU: Colony Forming Unit	**Colour:** Pt-Co Units (CU)	

Terms

APHA	American Public Health Association
CEC	Cation Exchange Capacity
COC	Chain of Custody
CP	Client Parent - QC was performed on samples pertaining to this report
CRM	Certified Reference Material (ISO17034) - reported as percent recovery.
Dry	Where moisture has been determined on a solid sample, the result is expressed on a dry weight basis.
Duplicate	A second piece of analysis from the same sample and reported in the same units as the result to show comparison.
LOR	Limit of Reporting.
LCS	Laboratory Control Sample - reported as percent recovery.
Method Blank	In the case of solid samples, these are performed on laboratory-certified clean sands and in the case of water samples, these are performed on de-ionised water.
NCP	Non-Client Parent - QC performed on samples not pertaining to this report, QC represents the sequence or batch that client samples were analysed within.
RPD	Relative Percent Difference between two Duplicate pieces of analysis.
SPIKE	Addition of the analyte to the sample and reported as percentage recovery.
SRA	Sample Receipt Advice
Surr - Surrogate	The addition of a similar compound to the analyte target is reported as percentage recovery. See below for acceptance criteria.
TBTO	Tributyltin oxide (*bis*-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment; however, free tributyltin was measured, and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits.
TCLP	Toxicity Characteristic Leaching Procedure
TEQ	Toxic Equivalency Quotient or Total Equivalence
QSM	US Department of Defense Quality Systems Manual Version 6.0
US EPA	United States Environmental Protection Agency
WA DWER	Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should only be used as a guide and may be different when site-specific Sampling Analysis and Quality Plan (SAQP) have been implemented.

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is ≤30%; however, the following acceptance guidelines are equally applicable:

Results <10 times the LOR:	No Limit
Results between 10-20 times the LOR:	RPD must lie between 0-50%
Results >20 times the LOR:	RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range, not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 – 150%, VOC recoveries 50 – 150%

PFAS field samples containing surrogate recoveries above the QC limit designated in QSM 6.0, where no positive PFAS results have been reported or reviewed, and no data was affected.

QC Data General Comments

1. Where a result is reported as less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown are not data from your samples.
3. pH and Free Chlorine analysed in the laboratory - Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time.Analysis will begin as soon as possible after sample receipt.
4. Recovery Data (Spikes & Surrogates) - where chromatographic interference does not allow the determination of recovery, the term "INT" appears against that analyte.
5. For Matrix Spikes and LCS results, a dash "-" in the report means that the specific analyte was not added to the QC sample.
6. Duplicate RPDs are calculated from raw analytical data; thus, it is possible to have two sets of data.

Quality Control Results

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Method Blank							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	ug/L	< 0.05			0.05	Pass	
Perfluoropentanoic acid (PFPeA)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanoic acid (PFHxA)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanoic acid (PFHpA)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanoic acid (PFOA)	ug/L	< 0.01			0.01	Pass	
Perfluorononanoic acid (PFNA)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanoic acid (PFDA)	ug/L	< 0.01			0.01	Pass	
Perfluoroundecanoic acid (PFUnDA)	ug/L	< 0.01			0.01	Pass	
Perfluorododecanoic acid (PFDoDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotridecanoic acid (PFTrDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotetradecanoic acid (PFTeDA)	ug/L	< 0.01			0.01	Pass	
Method Blank							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	ug/L	< 0.05			0.05	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	ug/L	< 0.05			0.05	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	ug/L	< 0.05			0.05	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	ug/L	< 0.05			0.05	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	ug/L	< 0.05			0.05	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	ug/L	< 0.05			0.05	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	ug/L	< 0.05			0.05	Pass	
Method Blank							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	ug/L	< 0.01			0.01	Pass	
Perfluorononanesulfonic acid (PFNS)	ug/L	< 0.01			0.01	Pass	
Perfluoropropanesulfonic acid (PFPrS)	ug/L	< 0.01			0.01	Pass	
Perfluoropentanesulfonic acid (PFPeS)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanesulfonic acid (PFHxS)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanesulfonic acid (PFOS)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanesulfonic acid (PFDS)	ug/L	< 0.01			0.01	Pass	
Method Blank							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	ug/L	< 0.05			0.05	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	ug/L	< 0.01			0.01	Pass	
LCS - % Recovery							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	%	71			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	%	76			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	%	77			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	%	77			50-150	Pass	
Perfluorooctanoic acid (PFOA)	%	73			50-150	Pass	
Perfluorononanoic acid (PFNA)	%	77			50-150	Pass	
Perfluorodecanoic acid (PFDA)	%	80			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	%	71			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	%	82			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	%	65			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	%	77			50-150	Pass	

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
LCS - % Recovery							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	%	87			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	%	75			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	%	84			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	%	90			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	%	84			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	%	77			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	%	92			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	%	80			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	%	87			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	%	74			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	%	84			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	%	79			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	%	81			50-150	Pass	
Perfluorooctanesulfonic acid (PFOS)	%	85			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	%	74			50-150	Pass	
LCS - % Recovery							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	%	82			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	%	84			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	%	78			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	%	78			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Spike - % Recovery									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1					
Perfluorobutanoic acid (PFBA)	M25-Oc0075775	NCP	%	88			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	M25-Oc0075775	NCP	%	88			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	M25-Oc0075775	NCP	%	90			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	M25-Oc0075775	NCP	%	88			50-150	Pass	
Perfluorooctanoic acid (PFOA)	M25-Oc0075775	NCP	%	82			50-150	Pass	
Perfluorononanoic acid (PFNA)	M25-Oc0075775	NCP	%	84			50-150	Pass	
Perfluorodecanoic acid (PFDA)	M25-Oc0075775	NCP	%	86			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	M25-Oc0075775	NCP	%	85			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	M25-Oc0075775	NCP	%	90			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	M25-Oc0075775	NCP	%	60			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	M25-Oc0075775	NCP	%	89			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonamido substances				Result 1					
Perfluorooctane sulfonamide (FOSA)	M25-Oc0075775	NCP	%	91			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	M25-Oc0075775	NCP	%	111			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	M25-Oc0075775	NCP	%	79			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	M25-Oc0075775	NCP	%	99			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	M25-Oc0075775	NCP	%	94			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	M25-Oc0075775	NCP	%	85			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	M25-Oc0075775	NCP	%	91			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1					
Perfluorobutanesulfonic acid (PFBS)	M25-Oc0075775	NCP	%	92			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	M25-Oc0075775	NCP	%	91			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	M25-Oc0075775	NCP	%	86			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	M25-Oc0075775	NCP	%	99			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	M25-Oc0075775	NCP	%	91			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	M25-Oc0075775	NCP	%	99			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	M25-Oc0075775	NCP	%	87			50-150	Pass	
Spike - % Recovery									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1					
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	M25-Oc0075775	NCP	%	96			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	M25-Oc0075775	NCP	%	93			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	M25-Oc0075775	NCP	%	88			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	M25-Oc0075775	NCP	%	104			50-150	Pass	
Test	**Lab Sample ID**	**QA Source**	**Units**	**Result 1**			**Acceptance Limits**	**Pass Limits**	**Qualifying Code**
Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluorobutanoic acid (PFBA)	M25-Oc0068688	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Perfluoropentanoic acid (PFPeA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanoic acid (PFHxA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroheptanoic acid (PFHpA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanoic acid (PFOA)	M25-Oc0074399	NCP	ug/L	0.02	0.03	1.0	30%	Pass	
Perfluorononanoic acid (PFNA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanoic acid (PFDA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroundecanoic acid (PFUnDA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorododecanoic acid (PFDoDA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotridecanoic acid (PFTrDA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotetradecanoic acid (PFTeDA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
Perfluorooctane sulfonamide (FOSA)	M25-Oc0068688	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	M25-Oc0068688	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	M25-Oc0068688	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	M25-Oc0068688	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	M25-Oc0068688	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	M25-Oc0068688	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	M25-Oc0068688	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1	Result 2	RPD			
Perfluorobutanesulfonic acid (PFBS)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanesulfonic acid (PFNS)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropropanesulfonic acid (PFPrS)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropentanesulfonic acid (PFPeS)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanesulfonic acid (PFHxS)	M25-Oc0074399	NCP	ug/L	0.16	0.16	3.0	30%	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanesulfonic acid (PFOS)	M25-Oc0074399	NCP	ug/L	0.02	0.02	11	30%	Pass	
Perfluorodecanesulfonic acid (PFDS)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1	Result 2	RPD			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	M25-Oc0074399	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	M25-Oc0068688	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Comments

Report 1283157-W-V2 (amendment to report 1283157-W) has been issued with corrected Project ID from (RICHMOUND) to (RICHMOND) as per COC.

Sample Integrity

Custody Seals Intact (if used)	N/A
Attempt to Chill was evident	Yes
Sample correctly preserved	Yes
Appropriate sample containers have been used	Yes
Sample containers for volatile analysis received with minimal headspace	Yes
Samples received within HoldingTime	Yes
Some samples have been subcontracted	No

Qualifier Codes/Comments

Code	Description
J-	If the percent recoveries were less than the lower control limit, qualify detects for the analyte in associated samples as estimated with a negative bias J-.
N11	Isotope dilution is used for calibration of each native compound for which an exact labelled analogue is available (Isotope Dilution Quantitation). The isotopically labelled analogues allow identification and recovery correction of the concentration of the associated native PFAS compounds.
N15	Where the native PFAS compound does not have labelled analogue then the quantification is made using the Extracted Internal Standard Analyte with the closest retention time to the analyte and no recovery correction has been made (Internal Standard Quantitation).

Authorised by:

Nileshni Goundar	Analytical Services Manager
Dylan Jones	Senior Analyst-PFAS



Glenn Jackson
Managing Director

Final Report – this report replaces any previously issued Report

- Indicates Not Requested

* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here.

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eurofins | Environment Testing

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NATA# 2377
Site# 2370 & 2554

Sample Receipt Advice

Company name:	Geo-Logix P/L
Contact name:	Paula Del Prete
Project name:	RICHMOUND
Project ID:	2301034G
Turnaround time:	5 Day
Date/Time received	Oct 22, 2025 2:46 PM
Eurofins reference	1283157

Sample Information

✓	A detailed list of analytes logged into our LIMS, is included in the attached summary table.
✓	Sample Temperature of chilled sample on the batch as recorded by Eurofins Sample Receipt : 12.3 degrees Celsius.
✓	All samples have been received as described on the above COC.
✓	COC has been completed correctly.
✓	Attempt to chill was evident.
✓	Appropriately preserved sample containers have been used.
✓	All samples were received in good condition.
✓	Samples have been provided with adequate time to commence analysis in accordance with the relevant holding times.
✓	Appropriate sample containers have been used.
✓	Sample containers for volatile analysis received with zero headspace.
X	Split sample sent to requested external lab.
X	Some samples have been subcontracted.
N/A	Custody Seals intact (if used).

Notes

Contact

If you have any questions with respect to these samples, please contact your Analytical Services Manager:

Bonnie Pu on phone : or by email: Bonnie.Pu@eurofinsanz.com

Results will be delivered electronically via email to Paula Del Prete - pdelprete@geo-logix.com.au.

Note: A copy of these results will also be delivered to the general Geo-Logix P/L email address.

SEND TO EUROFINS MELBOURNE.

Geo-Logix Pty Ltd
Building Q2, Level 3
2309/4 Daydream St
Warriewood, NSW 2102
ABN: 86 116 892 936
P: (02) 9979 1722
F: (02) 9979 1222

CHAIN OF CUSTODY

Page 1 of 1

Project Manager: Paula Del Prete, Harsha Muthu
Purchase Order No: PO6955
Contact email: pdelprete@geo-logix.com.au / hmuthu@geo-logix.com.au
Quote Reference: -
Project Name: Richmond
Send Invoice to: accounts@geo-logix.com.au
Project Number: 2301034g **Date Submitted:** 22-10-25
TAT required: STD

ANALYSIS REQUIRED

Lab ID	Sample ID	Date	Matrix: soil	water	air	paint, filters	other	Comments	COMPOSITE	TRH - C6 - C10	TRH - C10 - C40	VOCs	BTEXN	PAHs	PCBs	OCPs	OPPs	Phenols	Metals - M8	Metals - Lead	Metals - Specify **	TCLP	Asbestos (ID only)	Asbestos (WA DOH)	Foreign Materials	Conductivity (EC)	pH	PFAS - 30 compound		Hold	SUITE
	TW1	21-10-25		X				PFAS BOTTLE																				X			

Eurofins MGT Suite Codes

B1	TRH/BTEXN
B1A	TRH/MAH
B2	TRH/BTEXN/Pb
B2A	TRH/MAH/Pb
B3	PAH/Phenols
B4	TRH/BTEXN/PAH
B4A	TRH/BTEXN/PAH/Phenols
B5	TRH/BTEXN/M7
B6	TRH/BTEXN/M8
B7	TRH/BTEXN/PAH/M8
B7A	TRH/BTEXN/PAH/Phenols/M8
B8	TRH/VOC/PAH/M8
B9	TRH/BTEXN/PAH/OCP/M8
B10	TRH/BTEXN/PAH/OCP/OPP/M8
B11	Na/K/Ca/Mg/Cl/SO_4/CO_3/HCO_3/NH_3/NO_3
B11A	B11/Alkalinity
B11B	B11/EC/TDS
B12	TRH/BTEXN/Oxygenates/Ethanol
B12A	TRH/BTEXN/Oxygenates
B13	OCP/PCB
B14	OCP/OPP
B15	OCP/OPP/PCB
B16	TDS/SO_4/CH_4/Alk/BOD/COD/HPC/CUB
B17	SO_4/NO_3/Fe++/HPC/CUB
B18	Cl-/SO_4/pH
B19	N/P/K
B20	CEC/%ESP/Ca/Mg/Na/K

Metals(circle)** As, Cd, Cr, Cu, Ni, Pb, Zn, Hg, Cr^{6+}, Cr^{3+}, Fe^{2+}, Fe^{3+}, Be, B, Al, V, Mn, Fe, Co, Se, Sr, Sn, Mo, Ag, Ba, Tl, Bi, Sb

Chain of Custody

Relinquished by: paula del prete Date/Time: 22/10 Signature: PDP Received by: Maree Date/Time: 27/10 Signature:

Oath 2:46pm

129357

Nileshni Goundar

From: Ted Lilly <tlilly@geo-logix.com.au>
Sent: Monday, 27 October 2025 4:52 PM
To: Nileshni Goundar; Paula Del Prete
Cc: Harsha Muthu
Subject: RE: Eurofins Test Results, Invoice - Report 1283157 : Site RICHMOUND (2301034G)

Unverified Sender: The sender of this email has not been verified. Review the content of the message carefully and verify the identity of the sender before acting on this email: replying, opening attachments or clicking links.

Hi Nileshni,

Can we change the project name to Richmond.

Cheers,

Ted Lilly | Principal Engineer
Unit 2309/4 Daydream St, Warriewood NSW 2102
T: 02 9979 1722 | M: 0412 708 868 | W: www.geo-logix.com.au

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From: Nileshni Goundar <EET-ELVIS@eurofinsanz.com>
Sent: Monday, 27 October 2025 4:27 PM
To: Paula Del Prete <pdelprete@geo-logix.com.au>
Cc: Ted Lilly <tlilly@geo-logix.com.au>; Harsha Muthu <hmuthu@geo-logix.com.au>
Subject: Eurofins Test Results, Invoice - Report 1283157 : Site RICHMOUND (2301034G)

Please find the attached report.

Kind Regards,
Nileshni (Neena) Goundar
Assistant Analytical Services Manager
*Please note my work hours are **2pm-10pm,** anything outside of that please contact your ASM for anything urgent.*

Eurofins Environment Testing Australia Pty Ltd
179 Magowar Road
Girraween, NSW, 2145

Email: Nileshni.Goundar@eurofinsanz.com
Website: www.eurofins.com.au/environmental-testing

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Warriewood
NSW 2102

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NATA is a signatory to the ILAC Mutual Recognition Arrangement for the mutual recognition of the equivalence of testing, medical testing, calibration, inspection, proficiency testing scheme providers and reference materials producers reports and certificates.

Attention: **Paula Del Prete**

Report **1284389-W-V3**
Project name RICHMOND
Project ID 2301034G
Received Date Oct 24, 2025

Client Sample ID			**GW128**	**GW129**	**BH106**	**BH201**
Sample Matrix			**Water**	**Water**	**Water**	**Water**
Eurofins Sample No.			**S25-Oc0076880**	**S25-Oc0076881**	**S25-Oc0076882**	**S25-Oc0076883**
Date Sampled			**Oct 23, 2025**	**Oct 23, 2025**	**Oct 23, 2025**	**Oct 23, 2025**
Test/Reference	LOR	Unit				
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	0.20
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	< 0.01	0.02	< 0.01	0.30
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.01	< 0.01	< 0.01	1.6
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	[N09]0.56
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	[N09]0.55
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	89	89	101	78
13C5-PFPeA (surr.)	1	%	93	85	105	90
13C5-PFHxA (surr.)	1	%	116	100	112	93
13C4-PFHpA (surr.)	1	%	102	103	107	93
13C8-PFOA (surr.)	1	%	105	96	119	89
13C5-PFNA (surr.)	1	%	108	101	110	94
13C6-PFDA (surr.)	1	%	86	99	104	91
13C2-PFUnDA (surr.)	1	%	104	87	106	92
13C2-PFDoDA (surr.)	1	%	115	74	94	93
13C2-PFTeDA (surr.)	1	%	107	82	72	95
Perfluoroalkyl sulfonamido substances						
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	94	77	100	85
D3-N-MeFOSA (surr.)	1	%	94	50	96	86

Client Sample ID			GW128	GW129	BH106	BH201
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0076880	S25-Oc0076881	S25-Oc0076882	S25-Oc0076883
Date Sampled			Oct 23, 2025	Oct 23, 2025	Oct 23, 2025	Oct 23, 2025
Test/Reference	LOR	Unit				
Perfluoroalkyl sulfonamido substances						
D5-N-EtFOSA (surr.)	1	%	94	48	101	88
D7-N-MeFOSE (surr.)	1	%	90	59	90	81
D9-N-EtFOSE (surr.)	1	%	90	60	90	83
D5-N-EtFOSAA (surr.)	1	%	92	59	73	77
D3-N-MeFOSAA (surr.)	1	%	87	67	86	87
Perfluoroalkyl sulfonic acids (PFSAs)						
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	0.08	< 0.01	< 0.01	2.1
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	0.04	0.01	< 0.01	0.64
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	[N09]0.04	< 0.01	< 0.01	[N09]1.6
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]0.17	< 0.01	[N09]0.03	[N09]6.6
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	[N09]0.12
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	< 0.01	< 0.01	[N09]0.05	[N09]0.22
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	97	106	117	85
18O2-PFHxS (surr.)	1	%	102	108	121	81
13C8-PFOS (surr.)	1	%	101	101	106	92
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)						
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	96	INT	127	86
13C2-6:2 FTSA (surr.)	1	%	100	INT	109	81
13C2-8:2 FTSA (surr.)	1	%	90	122	115	88
13C2-10:2 FTSA (surr.)	1	%	101	85	94	88
PFASs Summations						
Sum (PFHxS + PFOS)*	0.01	ug/L	0.17	< 0.01	0.08	6.82
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	0.26	< 0.05	0.08	12.13
Sum of PFASs (n=30)*	0.1	ug/L	0.3	< 0.1	< 0.1	14

Client Sample ID			BH101	BH107	BH101H	R2
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0076884	S25-Oc0076885	S25-Oc0076886	S25-Oc0076887
Date Sampled			Oct 23, 2025	Oct 23, 2025	Oct 23, 2025	Oct 23, 2025
Test/Reference	LOR	Unit				
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	2.9	< 0.05	0.13	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	5.9	< 0.01	0.12	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	37	< 0.01	0.20	< 0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	[N09]10	< 0.01	[N09]0.11	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	[N09]31	< 0.01	[N09]0.09	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	0.09	< 0.01	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01

Client Sample ID			BH101	BH107	BH101H	R2
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0076884	S25-Oc0076885	S25-Oc0076886	S25-Oc0076887
Date Sampled			Oct 23, 2025	Oct 23, 2025	Oct 23, 2025	Oct 23, 2025
Test/Reference	LOR	Unit				
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	77	88	84	87
13C5-PFPeA (surr.)	1	%	71	94	90	86
13C5-PFHxA (surr.)	1	%	96	107	105	96
13C4-PFHpA (surr.)	1	%	72	111	94	100
13C8-PFOA (surr.)	1	%	83	110	98	103
13C5-PFNA (surr.)	1	%	84	100	103	90
13C6-PFDA (surr.)	1	%	63	96	94	95
13C2-PFUnDA (surr.)	1	%	93	101	91	96
13C2-PFDoDA (surr.)	1	%	96	89	91	95
13C2-PFTeDA (surr.)	1	%	111	86	85	97
Perfluoroalkyl sulfonamido substances						
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	86	90	84	102
D3-N-MeFOSA (surr.)	1	%	86	87	82	96
D5-N-EtFOSA (surr.)	1	%	88	92	92	100
D7-N-MeFOSE (surr.)	1	%	78	83	79	89
D9-N-EtFOSE (surr.)	1	%	82	81	81	85
D5-N-EtFOSAA (surr.)	1	%	60	67	78	88
D3-N-MeFOSAA (surr.)	1	%	69	76	80	89
Perfluoroalkyl sulfonic acids (PFSAs)						
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	26	< 0.01	0.14	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	0.24	< 0.01	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	5.7	< 0.01	0.08	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	[N09]27	< 0.01	[N09]0.08	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]180	< 0.01	[N09]0.77	< 0.01
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	[N09]9.1	< 0.01	< 0.01	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	[N09]250	< 0.01	[N09]0.02	< 0.01
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	93	101	98	99
18O2-PFHxS (surr.)	1	%	90	100	97	100
13C8-PFOS (surr.)	1	%	89	98	96	95

Client Sample ID			BH101	BH107	BH101H	R2
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0076884	S25-Oc0076885	S25-Oc0076886	S25-Oc0076887
Date Sampled			Oct 23, 2025	Oct 23, 2025	Oct 23, 2025	Oct 23, 2025
Test/Reference	LOR	Unit				
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)						
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	104	90	107	85
13C2-6:2 FTSA (surr.)	1	%	90	86	88	87
13C2-8:2 FTSA (surr.)	1	%	79	90	86	80
13C2-10:2 FTSA (surr.)	1	%	88	95	96	92
PFASs Summations						
Sum (PFHxS + PFOS)*	0.01	ug/L	430	< 0.01	0.79	< 0.01
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	542.8	< 0.05	1.58	< 0.05
Sum of PFASs (n=30)*	0.1	ug/L	580	< 0.1	1.7	< 0.1

Client Sample ID			DW2	DW3
Sample Matrix			Water	Water
Eurofins Sample No.			S25-Oc0076888	S25-Oc0076889
Date Sampled			Oct 23, 2025	Oct 23, 2025
Test/Reference	LOR	Unit		
Perfluoroalkyl carboxylic acids (PFCAs)				
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.02	< 0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	84	89
13C5-PFPeA (surr.)	1	%	93	97
13C5-PFHxA (surr.)	1	%	95	105
13C4-PFHpA (surr.)	1	%	87	98
13C8-PFOA (surr.)	1	%	106	102
13C5-PFNA (surr.)	1	%	95	97
13C6-PFDA (surr.)	1	%	97	97
13C2-PFUnDA (surr.)	1	%	93	97
13C2-PFDoDA (surr.)	1	%	104	84
13C2-PFTeDA (surr.)	1	%	94	89

Client Sample ID			DW2	DW3
Sample Matrix			Water	Water
Eurofins Sample No.			S25-Oc0076888	S25-Oc0076889
Date Sampled			Oct 23, 2025	Oct 23, 2025
Test/Reference	LOR	Unit		
Perfluoroalkyl sulfonamido substances				
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	92	88
D3-N-MeFOSA (surr.)	1	%	98	87
D5-N-EtFOSA (surr.)	1	%	103	91
D7-N-MeFOSE (surr.)	1	%	91	83
D9-N-EtFOSE (surr.)	1	%	86	81
D5-N-EtFOSAA (surr.)	1	%	84	70
D3-N-MeFOSAA (surr.)	1	%	88	76
Perfluoroalkyl sulfonic acids (PFSAs)				
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	0.08	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	0.04	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	[N09]0.04	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]0.17	< 0.01
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	101	101
18O2-PFHxS (surr.)	1	%	100	105
13C8-PFOS (surr.)	1	%	95	93
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	87	95
13C2-6:2 FTSA (surr.)	1	%	104	83
13C2-8:2 FTSA (surr.)	1	%	85	81
13C2-10:2 FTSA (surr.)	1	%	94	93
PFASs Summations				
Sum (PFHxS + PFOS)*	0.01	ug/L	0.17	< 0.01
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	0.27	< 0.05
Sum of PFASs (n=30)*	0.1	ug/L	0.4	< 0.1

Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description	Testing Site	Extracted	Holding Time
Per- and Polyfluoroalkyl Substances (PFASs)			
Perfluoroalkyl carboxylic acids (PFCAs)	Sydney	Oct 31, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonamido substances	Sydney	Oct 31, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonic acids (PFSAs)	Sydney	Oct 31, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)	Sydney	Oct 31, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			

eurofins

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Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: RICHMOND
Project ID: 2301034G

Order No.: PO6958
Report #: 1284389
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Oct 24, 2025 5:45 PM
Due: Oct 31, 2025
Priority: 5 Day
Contact Name: Paula Del Prete

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail						Per- and Polyfluoroalkyl Substances (PFASs)
Sydney Laboratory - NATA # 1261 Site # 18217						X
External Laboratory						
No	**Sample ID**	**Sample Date**	**Sampling Time**	**Matrix**	**LAB ID**	
1	GW128	Oct 23, 2025		Water	S25-Oc0076880	X
2	GW129	Oct 23, 2025		Water	S25-Oc0076881	X
3	BH106	Oct 23, 2025		Water	S25-Oc0076882	X
4	BH201	Oct 23, 2025		Water	S25-Oc0076883	X
5	BH101	Oct 23, 2025		Water	S25-Oc0076884	X
6	BH107	Oct 23, 2025		Water	S25-Oc0076885	X
7	BH101H	Oct 23, 2025		Water	S25-Oc0076886	X
8	R2	Oct 23, 2025		Water	S25-Oc0076887	X
9	DW2	Oct 23, 2025		Water	S25-Oc0076888	X
10	DW3	Oct 23, 2025		Water	S25-Oc0076889	X
Test Counts						10

Internal Quality Control Review and Glossary

General

1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follow guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013. They are included in this QC report where applicable. Additional QC data may be available on request.
2. Unless otherwise stated, all soil/sediment/solid results are reported on a dry weight basis.
3. Unless otherwise stated, all biota/food results are reported on a wet weight basis on the edible portion.
4. For CEC results where the sample's origin is unknown or environmentally contaminated, the results should be used advisedly.
5. Actual LORs are matrix dependent. Quoted LORs may be raised where sample extracts are diluted due to interferences.
6. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds where annotated.
7. SVOC analysis on waters is performed on homogenised, unfiltered samples unless noted otherwise.
8. Samples were analysed on an 'as received' basis.
9. Information identified in this report with **blue** colour indicates data provided by customers that may have an impact on the results.
10. This report replaces any interim results previously issued.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours before sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the sampling date; therefore, compliance with these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether, the holding time is seven days; however, for all other VOCs, such as BTEX or C6-10 TRH, the holding time is 14 days.

Units

mg/kg: milligrams per kilogram	**mg/L:** milligrams per litre	**ppm:** parts per million
µg/L: micrograms per litre	**ppb:** parts per billion	**%:** Percentage
org/100 mL: Organisms per 100 millilitres	**NTU:** Nephelometric Turbidity Units	**MPN/100 mL:** Most Probable Number of organisms per 100 millilitres
CFU: Colony Forming Unit	**Colour:** Pt-Co Units (CU)	

Terms

APHA	American Public Health Association
CEC	Cation Exchange Capacity
COC	Chain of Custody
CP	Client Parent - QC was performed on samples pertaining to this report
CRM	Certified Reference Material (ISO17034) - reported as percent recovery.
Dry	Where moisture has been determined on a solid sample, the result is expressed on a dry weight basis.
Duplicate	A second piece of analysis from the same sample and reported in the same units as the result to show comparison.
LOR	Limit of Reporting.
LCS	Laboratory Control Sample - reported as percent recovery.
Method Blank	In the case of solid samples, these are performed on laboratory-certified clean sands and in the case of water samples, these are performed on de-ionised water.
NCP	Non-Client Parent - QC performed on samples not pertaining to this report, QC represents the sequence or batch that client samples were analysed within.
RPD	Relative Percent Difference between two Duplicate pieces of analysis.
SPIKE	Addition of the analyte to the sample and reported as percentage recovery.
SRA	Sample Receipt Advice
Surr - Surrogate	The addition of a similar compound to the analyte target is reported as percentage recovery. See below for acceptance criteria.
TBTO	Tributyltin oxide (*bis*-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment; however, free tributyltin was measured, and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits.
TCLP	Toxicity Characteristic Leaching Procedure
TEQ	Toxic Equivalency Quotient or Total Equivalence
QSM	US Department of Defense Quality Systems Manual Version 6.0
US EPA	United States Environmental Protection Agency
WA DWER	Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should only be used as a guide and may be different when site-specific Sampling Analysis and Quality Plan (SAQP) have been implemented.

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is ≤30%; however, the following acceptance guidelines are equally applicable:

Results <10 times the LOR:	No Limit
Results between 10-20 times the LOR:	RPD must lie between 0-50%
Results >20 times the LOR:	RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range, not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 – 150%, VOC recoveries 50 – 150%

PFAS field samples containing surrogate recoveries above the QC limit designated in QSM 6.0, where no positive PFAS results have been reported or reviewed, and no data was affected.

QC Data General Comments

1. Where a result is reported as less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown are not data from your samples.
3. pH and Free Chlorine analysed in the laboratory - Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time.Analysis will begin as soon as possible after sample receipt.
4. Recovery Data (Spikes & Surrogates) - where chromatographic interference does not allow the determination of recovery, the term "INT" appears against that analyte.
5. For Matrix Spikes and LCS results, a dash "-" in the report means that the specific analyte was not added to the QC sample.
6. Duplicate RPDs are calculated from raw analytical data; thus, it is possible to have two sets of data.

Quality Control Results

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Method Blank							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	ug/L	< 0.05			0.05	Pass	
Perfluoropentanoic acid (PFPeA)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanoic acid (PFHxA)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanoic acid (PFHpA)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanoic acid (PFOA)	ug/L	< 0.01			0.01	Pass	
Perfluorononanoic acid (PFNA)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanoic acid (PFDA)	ug/L	< 0.01			0.01	Pass	
Perfluoroundecanoic acid (PFUnDA)	ug/L	< 0.01			0.01	Pass	
Perfluorododecanoic acid (PFDoDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotridecanoic acid (PFTrDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotetradecanoic acid (PFTeDA)	ug/L	< 0.01			0.01	Pass	
Method Blank							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	ug/L	< 0.05			0.05	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	ug/L	< 0.05			0.05	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	ug/L	< 0.05			0.05	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	ug/L	< 0.05			0.05	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	ug/L	< 0.05			0.05	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	ug/L	< 0.05			0.05	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	ug/L	< 0.05			0.05	Pass	
Method Blank							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	ug/L	< 0.01			0.01	Pass	
Perfluorononanesulfonic acid (PFNS)	ug/L	< 0.01			0.01	Pass	
Perfluoropropanesulfonic acid (PFPrS)	ug/L	< 0.01			0.01	Pass	
Perfluoropentanesulfonic acid (PFPeS)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanesulfonic acid (PFHxS)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanesulfonic acid (PFOS)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanesulfonic acid (PFDS)	ug/L	< 0.01			0.01	Pass	
Method Blank							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	ug/L	< 0.05			0.05	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	ug/L	< 0.01			0.01	Pass	
Method Blank							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	ug/L	< 0.05			0.05	Pass	
Perfluoropentanoic acid (PFPeA)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanoic acid (PFHxA)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanoic acid (PFHpA)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanoic acid (PFOA)	ug/L	< 0.01			0.01	Pass	
Perfluorononanoic acid (PFNA)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanoic acid (PFDA)	ug/L	< 0.01			0.01	Pass	
Perfluoroundecanoic acid (PFUnDA)	ug/L	< 0.01			0.01	Pass	
Perfluorododecanoic acid (PFDoDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotridecanoic acid (PFTrDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotetradecanoic acid (PFTeDA)	ug/L	< 0.01			0.01	Pass	

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Method Blank							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	ug/L	< 0.05			0.05	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	ug/L	< 0.05			0.05	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	ug/L	< 0.05			0.05	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	ug/L	< 0.05			0.05	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	ug/L	< 0.05			0.05	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	ug/L	< 0.05			0.05	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	ug/L	< 0.05			0.05	Pass	
Method Blank							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	ug/L	< 0.01			0.01	Pass	
Perfluorononanesulfonic acid (PFNS)	ug/L	< 0.01			0.01	Pass	
Perfluoropropanesulfonic acid (PFPrS)	ug/L	< 0.01			0.01	Pass	
Perfluoropentanesulfonic acid (PFPeS)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanesulfonic acid (PFHxS)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanesulfonic acid (PFOS)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanesulfonic acid (PFDS)	ug/L	< 0.01			0.01	Pass	
Method Blank							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	ug/L	< 0.05			0.05	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	ug/L	< 0.01			0.01	Pass	
LCS - % Recovery							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	%	105			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	%	107			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	%	115			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	%	112			50-150	Pass	
Perfluorooctanoic acid (PFOA)	%	104			50-150	Pass	
Perfluorononanoic acid (PFNA)	%	110			50-150	Pass	
Perfluorodecanoic acid (PFDA)	%	118			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	%	117			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	%	112			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	%	121			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	%	115			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	%	115			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	%	116			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	%	114			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	%	126			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	%	119			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	%	111			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	%	119			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	%	110			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	%	107			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	%	106			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	%	94			50-150	Pass	

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Perfluorohexanesulfonic acid (PFHxS)	%	103			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	%	120			50-150	Pass	
Perfluorooctanesulfonic acid (PFOS)	%	101			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	%	109			50-150	Pass	
LCS - % Recovery							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	%	112			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	%	121			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	%	108			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	%	123			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	%	105			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	%	107			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	%	115			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	%	112			50-150	Pass	
Perfluorooctanoic acid (PFOA)	%	104			50-150	Pass	
Perfluorononanoic acid (PFNA)	%	110			50-150	Pass	
Perfluorodecanoic acid (PFDA)	%	118			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	%	117			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	%	112			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	%	121			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	%	115			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	%	115			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	%	116			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	%	114			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	%	126			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	%	119			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	%	111			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	%	119			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	%	110			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	%	107			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	%	106			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	%	94			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	%	103			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	%	120			50-150	Pass	
Perfluorooctanesulfonic acid (PFOS)	%	101			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	%	109			50-150	Pass	
LCS - % Recovery							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	%	112			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	%	121			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	%	108			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	%	123			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Spike - % Recovery									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1					
Perfluorobutanoic acid (PFBA)	S25-Oc0077023	NCP	%	108			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	S25-Oc0077023	NCP	%	108			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	S25-Oc0077023	NCP	%	108			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	S25-Oc0077023	NCP	%	101			50-150	Pass	
Perfluorooctanoic acid (PFOA)	S25-Oc0077023	NCP	%	123			50-150	Pass	
Perfluorononanoic acid (PFNA)	S25-Oc0077023	NCP	%	103			50-150	Pass	
Perfluorodecanoic acid (PFDA)	S25-Oc0077023	NCP	%	117			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	S25-Oc0077023	NCP	%	117			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	S25-Oc0077023	NCP	%	121			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	S25-Oc0077023	NCP	%	126			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S25-Oc0077023	NCP	%	113			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonamido substances				Result 1					
Perfluorooctane sulfonamide (FOSA)	S25-Oc0077023	NCP	%	119			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S25-Oc0077023	NCP	%	131			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S25-Oc0077023	NCP	%	109			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S25-Oc0077023	NCP	%	119			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S25-Oc0077023	NCP	%	125			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S25-Oc0077023	NCP	%	100			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S25-Oc0077023	NCP	%	112			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1					
Perfluorobutanesulfonic acid (PFBS)	S25-Oc0077023	NCP	%	107			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	S25-Oc0077023	NCP	%	109			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S25-Oc0077023	NCP	%	106			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S25-Oc0077023	NCP	%	91			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S25-Oc0077023	NCP	%	104			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S25-Oc0077023	NCP	%	118			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	S25-Oc0077023	NCP	%	107			50-150	Pass	
Spike - % Recovery									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1					
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S25-Oc0077023	NCP	%	125			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S25-Oc0077023	NCP	%	109			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S25-Oc0077023	NCP	%	98			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S25-Oc0077023	NCP	%	114			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluorobutanoic acid (PFBA)	S25-Oc0076880	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Perfluoropentanoic acid (PFPeA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanoic acid (PFHxA)	S25-Oc0076880	CP	ug/L	0.01	0.01	4.0	30%	Pass	
Perfluoroheptanoic acid (PFHpA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanoic acid (PFOA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanoic acid (PFNA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanoic acid (PFDA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroundecanoic acid (PFUnDA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorododecanoic acid (PFDoDA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotridecanoic acid (PFTrDA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
Perfluorooctane sulfonamide (FOSA)	S25-Oc0076880	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S25-Oc0076880	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S25-Oc0076880	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S25-Oc0076880	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S25-Oc0076880	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S25-Oc0076880	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S25-Oc0076880	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1	Result 2	RPD			
Perfluorobutanesulfonic acid (PFBS)	S25-Oc0076880	CP	ug/L	0.08	0.07	13	30%	Pass	
Perfluorononanesulfonic acid (PFNS)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S25-Oc0076880	CP	ug/L	0.04	0.04	11	30%	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S25-Oc0076880	CP	ug/L	0.04	0.04	12	30%	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S25-Oc0076880	CP	ug/L	0.17	0.17	<1	30%	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanesulfonic acid (PFOS)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanesulfonic acid (PFDS)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Duplicate									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1	Result 2	RPD			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S25-Oc0076880	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S25-Oc0076880	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Comments

Report 1284389-W-V2 (amendment to report 1284389-W) has been issued with an updated sample ID from BH103 to BH106

Report 1284389-W-V3 (amendment to report 1284389-W-V2) has been issued following the migration of samples TW2 and TW3 to report 1288837 as they are intralab samples per the COC.

Sample Integrity

Custody Seals Intact (if used)	N/A
Attempt to Chill was evident	Yes
Sample correctly preserved	Yes
Appropriate sample containers have been used	Yes
Sample containers for volatile analysis received with minimal headspace	Yes
Samples received within HoldingTime	Yes
Some samples have been subcontracted	No

Qualifier Codes/Comments

Code	Description
N09	Quantification of linear and branched isomers has been conducted as a single total response using the relative response factor for the corresponding linear/branched standard.
N11	Isotope dilution is used for calibration of each native compound for which an exact labelled analogue is available (Isotope Dilution Quantitation). The isotopically labelled analogues allow identification and recovery correction of the concentration of the associated native PFAS compounds.
N15	Where the native PFAS compound does not have labelled analogue then the quantification is made using the Extracted Internal Standard Analyte with the closest retention time to the analyte and no recovery correction has been made (Internal Standard Quantitation).

Authorised by:

Bonnie Pu	Analytical Services Manager
Roopesh Rangarajan	Senior Analyst-PFAS



Glenn Jackson
Managing Director

Final Report – this report replaces any previously issued Report

- Indicates Not Requested

* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here.

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eurofins | Environment Testing

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Sample Receipt Advice

Company name:	Geo-Logix P/L
Contact name:	Paula Del Prete
Project name:	RICHMOND
Project ID:	2301034G
Turnaround time:	5 Day
Date/Time received	Oct 24, 2025 5:45 PM
Eurofins reference	1284389

Sample Information

✓	A detailed list of analytes logged into our LIMS, is included in the attached summary table.
✓	Sample Temperature of chilled sample on the batch as recorded by Eurofins Sample Receipt : 4.7 degrees Celsius.
✓	All samples have been received as described on the above COC.
✓	COC has been completed correctly.
✓	Attempt to chill was evident.
✓	Appropriately preserved sample containers have been used.
✓	All samples were received in good condition.
✓	Samples have been provided with adequate time to commence analysis in accordance with the relevant holding times.
✓	Appropriate sample containers have been used.
✓	Sample containers for volatile analysis received with zero headspace.
X	Split sample sent to requested external lab.
X	Some samples have been subcontracted.
N/A	Custody Seals intact (if used).

Notes

Sample ID BH103 based on COC.
Samples received by the laboratory after 5.30pm are deemed to have been received the following working day.

Contact

If you have any questions with respect to these samples, please contact your Analytical Services Manager:

Bonnie Pu on phone : or by email: Bonnie.Pu@eurofinsanz.com

Results will be delivered electronically via email to Paula Del Prete - pdelprete@geo-logix.com.au.

Note: A copy of these results will also be delivered to the general Geo-Logix P/L email address.

1284389

Geo-Logix Pty Ltd
Building Q2, Level 3
2309/4 Daydream St
Warriewood, NSW 2102
ABN: 86 116 892 936
P: (02) 9979 1722
F: (02) 9979 1222

CHAIN OF CUSTODY

Page 1 of 1

Project Manager: Paula Del Prete, Harsha Muthu
Purchase Order No: PO6958.
Contact email: pdelprete@geo-logix.com.au / hmuthu@geo-logix.com.au
Quote Reference: —
Project Name: Richmond
Send Invoice to: accounts@geo-logix.com.au
Project Number: 2301034g Date Submitted: 24-10-25.
TAT required: STD

ANALYSIS REQUIRED

Lab ID	Sample ID	Date	Matrix: soil	water	air	paint, filters	other	Comments	COMPOSITE	TRH - C6 - C10	TRH - C10 - C40	VOCs	BTEXN	PAHs	PCBs	OCPs	OPPs	Phenols	Metals - M8	Metals - Lead	Metals - Specify **	TCLP	Asbestos (ID only)	Asbestos (WA DOH)	Foreign Materials	Conductivity (EC)	pH	PFAS - 30 compound		Hold	SUITE
	GW128	23-10-25		X				PFAS BOTTLE																				X			
	GW129	"		X				"																							
	BH103	"		X				"																							
	BH201	"		X				"																							
	BH101	"		X				"																							
	BH107	"		X				"																				↓			
	BH101A	"		X				"																				X			
	D2	"		X				"																				X			
	DW2	"		X				"																				X			
	DW3	"		X				"																				X			

Eurofins MGT Suite Codes

B1	TRH/BTEXN
B1A	TRH/MAH
B2	TRH/BTEXN/Pb
B2A	TRH/MAH/Pb
B3	PAH/Phenols
B4	TRH/BTEXN/PAH
B4A	TRH/BTEXN/PAH/Phenols
B5	TRH/BTEXN/M7
B6	TRH/BTEXN/M8
B7	TRH/BTEXN/PAH/M8
B7A	TRH/BTEXN/PAH/Phenols/M8
B8	TRH/VOC/PAH/M8
B9	TRH/BTEXN/PAH/OCP/M8
B10	TRH/BTEXN/PAH/OCP/OPP/M8
B11	Na/K/Ca/Mg/Cl/SO_4/CO_3/HCO_3/NH_3/NO_3
B11A	B11/Alkalinity
B11B	B11/EC/TDS
B12	TRH/BTEXN/Oxygenates/Ethanol
B12A	TRH/BTEXN/Oxygenates
B13	OCP/PCB
B14	OCP/OPP
B15	OCP/OPP/PCB
B16	TDS/SO_4/CH_4/Alk/BOD/COD/HPC/CUB
B17	SO_4/NO_3/Fe++/HPC/CUB
B18	Cl-/SO_4/pH
B19	N/P/K
B20	CEC/%ESP/Ca/Mg/Na/K

Metals(circle)** As, Cd, Cr, Cu, Ni, Pb, Zn, Hg, Cr^{6+}, Cr^{3+}, Fe^{2+}, Fe^{3+}, Be, B, Al, V, Mn, Fe, Co, Se, Sr, Sn, Mo, Ag, Ba, Tl, Bi, Sb

Chain of Custody

Relinquished by: CP Date/Time: _____ Signature: _____
paula del prete 24-10-25.

Received by: Mary Ann Date/Time: 24/10 Signature: _____
5:45PM

4.7° chill

Version: V1
Issued: June 2019

Geo-Logix P/L
Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

NATA Accredited
Accreditation Number 1261
Site Number 1254

Accredited for compliance with ISO/IEC 17025 – Testing
NATA is a signatory to the ILAC Mutual Recognition Arrangement for the mutual recognition of the equivalence of testing, medical testing, calibration, inspection, proficiency testing scheme providers and reference materials producers reports and certificates.

Attention: **Paula Del Prete**

Report **1288837-W-V3**
Project name RICHMOND
Project ID 2301034G
Received Date Oct 24, 2025

Client Sample ID			**TW3**	**TW2**
Sample Matrix			**Water**	**Water**
Eurofins Sample No.			**S25-Oc0076890**	**S25-Oc0076891**
Date Sampled			**Oct 23, 2025**	**Oct 23, 2025**
Test/Reference	LOR	Unit		
Perfluoroalkyl carboxylic acids (PFCAs)				
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	< 0.01	0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	110	103
13C5-PFPeA (surr.)	1	%	95	95
13C5-PFHxA (surr.)	1	%	97	93
13C4-PFHpA (surr.)	1	%	95	90
13C8-PFOA (surr.)	1	%	96	90
13C5-PFNA (surr.)	1	%	96	91
13C6-PFDA (surr.)	1	%	104	97
13C2-PFUnDA (surr.)	1	%	128	129
13C2-PFDoDA (surr.)	1	%	122	126
13C2-PFTeDA (surr.)	1	%	126	118
Perfluoroalkyl sulfonamido substances				
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	74	71
D3-N-MeFOSA (surr.)	1	%	67	70

Client Sample ID			TW3	TW2
Sample Matrix			Water	Water
Eurofins Sample No.			S25-Oc0076890	S25-Oc0076891
Date Sampled			Oct 23, 2025	Oct 23, 2025
Test/Reference	LOR	Unit		
Perfluoroalkyl sulfonamido substances				
D5-N-EtFOSA (surr.)	1	%	83	87
D7-N-MeFOSE (surr.)	1	%	55	53
D9-N-EtFOSE (surr.)	1	%	58	64
D5-N-EtFOSAA (surr.)	1	%	J+165	146
D3-N-MeFOSAA (surr.)	1	%	144	130
Perfluoroalkyl sulfonic acids (PFSAs)				
Perfluorobutanesulfonic acid (PFBS)N11	0.01	ug/L	< 0.01	0.06
Perfluorononanesulfonic acid (PFNS)N15	0.01	ug/L	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)N15	0.01	ug/L	< 0.01	0.04
Perfluoropentanesulfonic acid (PFPeS)N15	0.01	ug/L	< 0.01	0.04
Perfluorohexanesulfonic acid (PFHxS)N11	0.01	ug/L	< 0.01	0.16
Perfluoroheptanesulfonic acid (PFHpS)N15	0.01	ug/L	< 0.01	< 0.01
Perfluorooctanesulfonic acid (PFOS)N11	0.01	ug/L	< 0.01	< 0.01
Perfluorodecanesulfonic acid (PFDS)N15	0.01	ug/L	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	104	92
18O2-PFHxS (surr.)	1	%	91	88
13C8-PFOS (surr.)	1	%	103	88
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)N11	0.01	ug/L	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)N11	0.05	ug/L	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)N11	0.01	ug/L	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)N11	0.01	ug/L	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	114	82
13C2-6:2 FTSA (surr.)	1	%	91	84
13C2-8:2 FTSA (surr.)	1	%	86	84
13C2-10:2 FTSA (surr.)	1	%	86	104
PFASs Summations				
Sum (PFHxS + PFOS)*	0.01	ug/L	< 0.01	0.16
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	< 0.05	0.23
Sum of PFASs (n=30)*	0.1	ug/L	< 0.1	0.3

Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description	Testing Site	Extracted	Holding Time
Per- and Polyfluoroalkyl Substances (PFASs)			
Perfluoroalkyl carboxylic acids (PFCAs)	Melbourne	Nov 07, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonamido substances	Melbourne	Nov 07, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonic acids (PFSAs)	Melbourne	Nov 07, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)	Melbourne	Nov 07, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			

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Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: RICHMOND
Project ID: 2301034G

Order No.: PO6958
Report #: 1288837
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Oct 24, 2025 5:45 PM
Due: Nov 7, 2025
Priority: 1 Day
Contact Name: Paula Del Prete

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail						Per- and Polyfluoroalkyl Substances (PFASs)
Melbourne Laboratory - NATA # 1261 Site # 1254						X
External Laboratory						
No	**Sample ID**	**Sample Date**	**Sampling Time**	**Matrix**	**LAB ID**	
1	TW3	Oct 23, 2025		Water	S25-Oc0076890	X
2	TW2	Oct 23, 2025		Water	S25-Oc0076891	X
Test Counts						2

Internal Quality Control Review and Glossary

General

1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follow guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013. They are included in this QC report where applicable. Additional QC data may be available on request.
2. Unless otherwise stated, all soil/sediment/solid results are reported on a dry weight basis.
3. Unless otherwise stated, all biota/food results are reported on a wet weight basis on the edible portion.
4. For CEC results where the sample's origin is unknown or environmentally contaminated, the results should be used advisedly.
5. Actual LORs are matrix dependent. Quoted LORs may be raised where sample extracts are diluted due to interferences.
6. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds where annotated.
7. SVOC analysis on waters is performed on homogenised, unfiltered samples unless noted otherwise.
8. Samples were analysed on an 'as received' basis.
9. Information identified in this report with **blue** colour indicates data provided by customers that may have an impact on the results.
10. This report replaces any interim results previously issued.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours before sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the sampling date; therefore, compliance with these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether, the holding time is seven days; however, for all other VOCs, such as BTEX or C6-10 TRH, the holding time is 14 days.

Units

mg/kg: milligrams per kilogram	**mg/L:** milligrams per litre	**ppm:** parts per million
µg/L: micrograms per litre	**ppb:** parts per billion	**%:** Percentage
org/100 mL: Organisms per 100 millilitres	**NTU:** Nephelometric Turbidity Units	**MPN/100 mL:** Most Probable Number of organisms per 100 millilitres
CFU: Colony Forming Unit	**Colour:** Pt-Co Units (CU)	

Terms

APHA	American Public Health Association
CEC	Cation Exchange Capacity
COC	Chain of Custody
CP	Client Parent - QC was performed on samples pertaining to this report
CRM	Certified Reference Material (ISO17034) - reported as percent recovery.
Dry	Where moisture has been determined on a solid sample, the result is expressed on a dry weight basis.
Duplicate	A second piece of analysis from the same sample and reported in the same units as the result to show comparison.
LOR	Limit of Reporting.
LCS	Laboratory Control Sample - reported as percent recovery.
Method Blank	In the case of solid samples, these are performed on laboratory-certified clean sands and in the case of water samples, these are performed on de-ionised water.
NCP	Non-Client Parent - QC performed on samples not pertaining to this report, QC represents the sequence or batch that client samples were analysed within.
RPD	Relative Percent Difference between two Duplicate pieces of analysis.
SPIKE	Addition of the analyte to the sample and reported as percentage recovery.
SRA	Sample Receipt Advice
Surr - Surrogate	The addition of a similar compound to the analyte target is reported as percentage recovery. See below for acceptance criteria.
TBTO	Tributyltin oxide (*bis*-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment; however, free tributyltin was measured, and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits.
TCLP	Toxicity Characteristic Leaching Procedure
TEQ	Toxic Equivalency Quotient or Total Equivalence
QSM	US Department of Defense Quality Systems Manual Version 6.0
US EPA	United States Environmental Protection Agency
WA DWER	Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should only be used as a guide and may be different when site-specific Sampling Analysis and Quality Plan (SAQP) have been implemented.

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is ≤30%; however, the following acceptance guidelines are equally applicable:

Results <10 times the LOR:	No Limit
Results between 10-20 times the LOR:	RPD must lie between 0-50%
Results >20 times the LOR:	RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range, not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 – 150%, VOC recoveries 50 – 150%

PFAS field samples containing surrogate recoveries above the QC limit designated in QSM 6.0, where no positive PFAS results have been reported or reviewed, and no data was affected.

QC Data General Comments

1. Where a result is reported as less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown are not data from your samples.
3. pH and Free Chlorine analysed in the laboratory - Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time.Analysis will begin as soon as possible after sample receipt.
4. Recovery Data (Spikes & Surrogates) - where chromatographic interference does not allow the determination of recovery, the term "INT" appears against that analyte.
5. For Matrix Spikes and LCS results, a dash "-" in the report means that the specific analyte was not added to the QC sample.
6. Duplicate RPDs are calculated from raw analytical data; thus, it is possible to have two sets of data.

Quality Control Results

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Method Blank							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	ug/L	< 0.05			0.05	Pass	
Perfluoropentanoic acid (PFPeA)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanoic acid (PFHxA)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanoic acid (PFHpA)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanoic acid (PFOA)	ug/L	< 0.01			0.01	Pass	
Perfluorononanoic acid (PFNA)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanoic acid (PFDA)	ug/L	< 0.01			0.01	Pass	
Perfluoroundecanoic acid (PFUnDA)	ug/L	< 0.01			0.01	Pass	
Perfluorododecanoic acid (PFDoDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotridecanoic acid (PFTrDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotetradecanoic acid (PFTeDA)	ug/L	< 0.01			0.01	Pass	
Method Blank							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	ug/L	< 0.05			0.05	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	ug/L	< 0.05			0.05	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	ug/L	< 0.05			0.05	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	ug/L	< 0.05			0.05	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	ug/L	< 0.05			0.05	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	ug/L	< 0.05			0.05	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	ug/L	< 0.05			0.05	Pass	
Method Blank							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	ug/L	< 0.01			0.01	Pass	
Perfluorononanesulfonic acid (PFNS)	ug/L	< 0.01			0.01	Pass	
Perfluoropropanesulfonic acid (PFPrS)	ug/L	< 0.01			0.01	Pass	
Perfluoropentanesulfonic acid (PFPeS)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanesulfonic acid (PFHxS)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanesulfonic acid (PFOS)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanesulfonic acid (PFDS)	ug/L	< 0.01			0.01	Pass	
Method Blank							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	ug/L	< 0.05			0.05	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	ug/L	< 0.01			0.01	Pass	
LCS - % Recovery							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	%	84			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	%	91			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	%	95			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	%	90			50-150	Pass	
Perfluorooctanoic acid (PFOA)	%	92			50-150	Pass	
Perfluorononanoic acid (PFNA)	%	86			50-150	Pass	
Perfluorodecanoic acid (PFDA)	%	85			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	%	84			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	%	88			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	%	64			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	%	83			50-150	Pass	

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
LCS - % Recovery							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	%	101			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	%	100			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	%	73			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	%	103			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	%	94			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	%	90			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	%	104			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	%	80			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	%	83			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	%	83			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	%	91			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	%	100			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	%	103			50-150	Pass	
Perfluorooctanesulfonic acid (PFOS)	%	102			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	%	79			50-150	Pass	
LCS - % Recovery							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	%	82			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	%	81			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	%	81			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	%	109			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Spike - % Recovery									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1					
Perfluorobutanoic acid (PFBA)	S25-Oc0077023	NCP	%	108			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	S25-Oc0077023	NCP	%	108			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	S25-Oc0077023	NCP	%	108			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	S25-Oc0077023	NCP	%	101			50-150	Pass	
Perfluorooctanoic acid (PFOA)	S25-Oc0077023	NCP	%	123			50-150	Pass	
Perfluorononanoic acid (PFNA)	S25-Oc0077023	NCP	%	103			50-150	Pass	
Perfluorodecanoic acid (PFDA)	S25-Oc0077023	NCP	%	117			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	S25-Oc0077023	NCP	%	117			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	S25-Oc0077023	NCP	%	121			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	S25-Oc0077023	NCP	%	126			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S25-Oc0077023	NCP	%	113			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonamido substances				Result 1					
Perfluorooctane sulfonamide (FOSA)	S25-Oc0077023	NCP	%	119			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S25-Oc0077023	NCP	%	131			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S25-Oc0077023	NCP	%	109			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S25-Oc0077023	NCP	%	119			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S25-Oc0077023	NCP	%	125			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S25-Oc0077023	NCP	%	100			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S25-Oc0077023	NCP	%	112			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1					
Perfluorobutanesulfonic acid (PFBS)	S25-Oc0077023	NCP	%	107			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	S25-Oc0077023	NCP	%	109			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S25-Oc0077023	NCP	%	106			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S25-Oc0077023	NCP	%	91			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S25-Oc0077023	NCP	%	104			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S25-Oc0077023	NCP	%	118			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	S25-Oc0077023	NCP	%	107			50-150	Pass	
Spike - % Recovery									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1					
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S25-Oc0077023	NCP	%	125			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S25-Oc0077023	NCP	%	109			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S25-Oc0077023	NCP	%	98			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S25-Oc0077023	NCP	%	114			50-150	Pass	
Test	**Lab Sample ID**	**QA Source**	**Units**	**Result 1**			**Acceptance Limits**	**Pass Limits**	**Qualifying Code**
Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluorobutanoic acid (PFBA)	S25-Oc0083731	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Perfluoropentanoic acid (PFPeA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanoic acid (PFHxA)	S25-Oc0083731	NCP	ug/L	0.03	0.03	15	30%	Pass	
Perfluoroheptanoic acid (PFHpA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanoic acid (PFOA)	S25-Oc0083731	NCP	ug/L	0.02	0.01	9.0	30%	Pass	
Perfluorononanoic acid (PFNA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanoic acid (PFDA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroundecanoic acid (PFUnDA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorododecanoic acid (PFDoDA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotridecanoic acid (PFTrDA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
Perfluorooctane sulfonamide (FOSA)	S25-Oc0083731	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S25-Oc0083731	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S25-Oc0083731	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S25-Oc0083731	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S25-Oc0083731	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S25-Oc0083731	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S25-Oc0083731	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1	Result 2	RPD			
Perfluorobutanesulfonic acid (PFBS)	S25-Oc0083731	NCP	ug/L	0.02	0.02	1.0	30%	Pass	
Perfluorononanesulfonic acid (PFNS)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S25-Oc0083731	NCP	ug/L	0.02	0.02	5.0	30%	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S25-Oc0083731	NCP	ug/L	0.22	0.23	4.0	30%	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S25-Oc0083731	NCP	ug/L	0.01	0.01	4.0	30%	Pass	
Perfluorooctanesulfonic acid (PFOS)	S25-Oc0083731	NCP	ug/L	0.37	0.39	5.0	30%	Pass	
Perfluorodecanesulfonic acid (PFDS)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1	Result 2	RPD			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S25-Oc0083731	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S25-Oc0083731	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Comments

Report 1288837-W-V2 (amendment to report 1288837-W) has been issued with PFAS results for all samples analysed as intralab samples per COC.

Report 1288837-W-V3 (amendment to report 1288837-W-V2) has been issued with corrected sample IDs from TW2 to TW3 and TW3 to TW2.

Sample Integrity

Custody Seals Intact (if used)	N/A
Attempt to Chill was evident	Yes
Sample correctly preserved	Yes
Appropriate sample containers have been used	Yes
Sample containers for volatile analysis received with minimal headspace	Yes
Samples received within HoldingTime	Yes
Some samples have been subcontracted	No

Qualifier Codes/Comments

Code	Description
J+	If the percent recoveries were greater than the upper control limit, qualify detects for the analyte in associated samples as estimated with a positive bias J+.
N11	Isotope dilution is used for calibration of each native compound for which an exact labelled analogue is available (Isotope Dilution Quantitation). The isotopically labelled analogues allow identification and recovery correction of the concentration of the associated native PFAS compounds.
N15	Where the native PFAS compound does not have labelled analogue then the quantification is made using the Extracted Internal Standard Analyte with the closest retention time to the analyte and no recovery correction has been made (Internal Standard Quantitation).

Authorised by:

Bonnie Pu	Analytical Services Manager
Joseph Edouard	Senior Analyst-PFAS
Roopesh Rangarajan	Senior Analyst-PFAS



Glenn Jackson
Managing Director

Final Report – this report replaces any previously issued Report

- Indicates Not Requested

* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here.

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eurofins | Environment Testing

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Sample Receipt Advice

Company name:	Geo-Logix P/L
Contact name:	Paula Del Prete
Project name:	RICHMOND
Project ID:	2301034G
Turnaround time:	1 Day
Date/Time received	Oct 24, 2025 5:45 PM
Eurofins reference	1288837

Sample Information

✓	A detailed list of analytes logged into our LIMS, is included in the attached summary table.
✓	Sample Temperature of chilled sample on the batch as recorded by Eurofins Sample Receipt : 4.7 degrees Celsius.
✓	All samples have been received as described on the above COC.
✓	COC has been completed correctly.
✓	Attempt to chill was evident.
✓	Appropriately preserved sample containers have been used.
✓	All samples were received in good condition.
✓	Samples have been provided with adequate time to commence analysis in accordance with the relevant holding times.
✓	Appropriate sample containers have been used.
✓	Sample containers for volatile analysis received with zero headspace.
X	Split sample sent to requested external lab.
X	Some samples have been subcontracted.
N/A	Custody Seals intact (if used).

Notes

Samples received by the laboratory after 5.30pm are deemed to have been received the following working day.

Contact

If you have any questions with respect to these samples, please contact your Analytical Services Manager:

Bonnie Pu on phone : or by email: Bonnie.Pu@eurofinsanz.com

Results will be delivered electronically via email to Paula Del Prete - pdelprete@geo-logix.com.au.

Note: A copy of these results will also be delivered to the general Geo-Logix P/L email address.

SEND TO EUROFINS MELBOURNE.

Geo-Logix Pty Ltd
Building Q2, Level 3
2309/4 Daydream St
Warriewood, NSW 2102
ABN: 86 116 892 936
P: (02) 9979 1722
F: (02) 9979 1222

CHAIN OF CUSTODY

Page 1 of 1

Project Manager: Paula Del Prete, Harsha Muthu
Purchase Order No: PO6957.
Contact email: pdelprete@geo-logix.com.au / hmuthu@geo-logix.com.au
Quote Reference: -
Project Name: Richmond
Send Invoice to: accounts@geo-logix.com.au
Project Number: 2301034g Date Submitted: 24-10-25.
TAT required: STD

ANALYSIS REQUIRED

Lab ID	Sample ID	Date	Matrix: soil	Matrix: water	Matrix: air	Matrix: paint, filters	Matrix: other	Comments	COMPOSITE	TRH - C6 - C10	TRH - C10 - C40	VOCs	BTEXN	PAHs	PCBs	OCPs	OPPs	Phenols	Metals - M8	Metals - Lead	Metals - Specify **	TCLP	Asbestos (ID only)	Asbestos (WA DOH)	Foreign Materials	Conductivity (EC)	pH	PFAS - 30 compound		Hold	SUITE
	TW2	23-10-25		X				PFAS bottle																				X			
	TW3	"		X				"																				X			

Eurofins MGT Suite Codes

B1	TRH/BTEXN
B1A	TRH/MAH
B2	TRH/BTEXN/Pb
B2A	TRH/MAH/Pb
B3	PAH/Phenols
B4	TRH/BTEXN/PAH
B4A	TRH/BTEXN/PAH/Phenols
B5	TRH/BTEXN/M7
B6	TRH/BTEXN/M8
B7	TRH/BTEXN/PAH/M8
B7A	TRH/BTEXN/PAH/Phenols/M8
B8	TRH/VOC/PAH/M8
B9	TRH/BTEXN/PAH/OCP/M8
B10	TRH/BTEXN/PAH/OCP/OPP/M8
B11	Na/K/Ca/Mg/Cl/SO_4/CO_3/HCO_3/NH_3/NO_3
B11A	B11/Alkalinity
B11B	B11/EC/TDS
B12	TRH/BTEXN/Oxygenates/Ethanol
B12A	TRH/BTEXN/Oxygenates
B13	OCP/PCB
B14	OCP/OPP
B15	OCP/OPP/PCB
B16	TDS/SO_4/CH_4/Alk/BOD/COD/HPC/CUB
B17	SO_4/NO_3/Fe++/HPC/CUB
B18	Cl-/SO_4/pH
B19	N/P/K
B20	CEC/%ESP/Ca/Ma/Na/K

Metals(circle)** As, Cd, Cr, Cu, Ni, Pb, Zn, Hg, Cr^{6+}, Cr^{3+}, Fe^{2+}, Fe^{3+}, Be, B, Al, V, Mn, Fe, Co, Se, Sr, Sn, Mo, Ag, Ba, Tl, Bi, Sb

Chain of Custody

Relinquished by: CLP Date/Time: _____ Signature: _____ Received by: _____ Date/Time: _____ Signature: _____

Paula Del Prete 24/10/25.

Geo-Logix P/L
Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

NATA Accredited
Accreditation Number 1261
Site Number 18217

Accredited for compliance with ISO/IEC 17025 – Testing
NATA is a signatory to the ILAC Mutual Recognition Arrangement for the mutual recognition of the equivalence of testing, medical testing, calibration, inspection, proficiency testing scheme providers and reference materials producers reports and certificates.

Attention: **Paula Del Prete**

Report **1285891-W**
Project name RICHMOND
Project ID 2301034G
Received Date Oct 29, 2025

Client Sample ID			**MW03-S**	**MW11**	**MW02**	**BH101i**
Sample Matrix			**Water**	**Water**	**Water**	**Water**
Eurofins Sample No.			**S25-Oc0089229**	**S25-Oc0089230**	**S25-Oc0089231**	**S25-Oc0089232**
Date Sampled			**Oct 28, 2025**	**Oct 28, 2025**	**Oct 28, 2025**	**Oct 28, 2025**
Test/Reference	LOR	Unit				
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	0.23
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	< 0.01	< 0.01	0.04	0.19
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.05	< 0.01	0.05	0.53
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	< 0.01	[N09]0.03	[N09]0.13
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	[N09]0.01	< 0.01	[N09]0.04	[N09]0.10
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	103	104	95	104
13C5-PFPeA (surr.)	1	%	106	112	103	106
13C5-PFHxA (surr.)	1	%	111	118	115	118
13C4-PFHpA (surr.)	1	%	107	111	105	105
13C8-PFOA (surr.)	1	%	107	113	109	107
13C5-PFNA (surr.)	1	%	99	106	111	97
13C6-PFDA (surr.)	1	%	100	106	107	102
13C2-PFUnDA (surr.)	1	%	105	98	112	86
13C2-PFDoDA (surr.)	1	%	104	99	122	88
13C2-PFTeDA (surr.)	1	%	102	79	112	74
Perfluoroalkyl sulfonamido substances						
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	95	99	105	107
D3-N-MeFOSA (surr.)	1	%	96	99	105	100

Client Sample ID			MW03-S	MW11	MW02	BH101i
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0089229	S25-Oc0089230	S25-Oc0089231	S25-Oc0089232
Date Sampled			Oct 28, 2025	Oct 28, 2025	Oct 28, 2025	Oct 28, 2025
Test/Reference	LOR	Unit				
Perfluoroalkyl sulfonamido substances						
D5-N-EtFOSA (surr.)	1	%	107	101	107	106
D7-N-MeFOSE (surr.)	1	%	102	92	95	99
D9-N-EtFOSE (surr.)	1	%	97	88	96	98
D5-N-EtFOSAA (surr.)	1	%	90	86	96	91
D3-N-MeFOSAA (surr.)	1	%	96	91	100	87
Perfluoroalkyl sulfonic acids (PFSAs)						
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	0.26	0.09	0.02	0.37
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	0.08	0.04	< 0.01	0.12
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	[N09]0.24	[N09]0.06	< 0.01	[N09]0.27
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]1.7	[N09]0.53	[N09]0.05	[N09]1.9
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	[N09]0.03	< 0.01	< 0.01	[N09]0.02
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	[N09]0.35	< 0.01	< 0.01	[N09]0.25
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	109	120	106	109
18O2-PFHxS (surr.)	1	%	108	111	109	97
13C8-PFOS (surr.)	1	%	107	109	110	87
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)						
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	114	131	119	133
13C2-6:2 FTSA (surr.)	1	%	106	103	102	103
13C2-8:2 FTSA (surr.)	1	%	97	101	98	100
13C2-10:2 FTSA (surr.)	1	%	92	94	97	85
PFASs Summations						
Sum (PFHxS + PFOS)*	0.01	ug/L	2.05	0.53	0.05	2.15
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	2.37	0.62	0.23	3.7
Sum of PFASs (n=30)*	0.1	ug/L	2.7	0.7	0.2	4.1

Client Sample ID			BH111	BH105	BH101G	R3
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0089233	S25-Oc0089234	S25-Oc0089235	S25-Oc0089236
Date Sampled			Oct 28, 2025	Oct 28, 2025	Oct 28, 2025	Oct 28, 2025
Test/Reference	LOR	Unit				
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05	0.10	< 0.05	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	0.02	0.02	< 0.01	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.02	0.04	< 0.01	< 0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	[N09]0.03	< 0.01	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01	[N09]0.02	< 0.01	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01

Client Sample ID			BH111	BH105	BH101G	R3
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0089233	S25-Oc0089234	S25-Oc0089235	S25-Oc0089236
Date Sampled			Oct 28, 2025	Oct 28, 2025	Oct 28, 2025	Oct 28, 2025
Test/Reference	LOR	Unit				
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	106	97	51	89
13C5-PFPeA (surr.)	1	%	110	104	47	104
13C5-PFHxA (surr.)	1	%	131	120	66	103
13C4-PFHpA (surr.)	1	%	112	118	55	99
13C8-PFOA (surr.)	1	%	119	112	65	113
13C5-PFNA (surr.)	1	%	103	104	55	102
13C6-PFDA (surr.)	1	%	102	109	52	102
13C2-PFUnDA (surr.)	1	%	90	116	57	101
13C2-PFDoDA (surr.)	1	%	96	107	58	102
13C2-PFTeDA (surr.)	1	%	83	115	57	110
Perfluoroalkyl sulfonamido substances						
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	103	103	56	102
D3-N-MeFOSA (surr.)	1	%	96	103	53	95
D5-N-EtFOSA (surr.)	1	%	110	115	55	104
D7-N-MeFOSE (surr.)	1	%	99	93	54	100
D9-N-EtFOSE (surr.)	1	%	99	93	50	94
D5-N-EtFOSAA (surr.)	1	%	86	91	54	99
D3-N-MeFOSAA (surr.)	1	%	94	89	54	100
Perfluoroalkyl sulfonic acids (PFSAs)						
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	0.02	0.04	< 0.01	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	0.01	0.01	< 0.01	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	< 0.01	[N09]0.04	< 0.01	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]0.04	[N09]1.7	< 0.01	< 0.01
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	[N09]0.02	< 0.01	< 0.01	< 0.01
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	112	114	59	109
18O2-PFHxS (surr.)	1	%	98	108	60	116
13C8-PFOS (surr.)	1	%	97	115	62	115

Client Sample ID			BH111	BH105	BH101G	R3
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S25-Oc0089233	S25-Oc0089234	S25-Oc0089235	S25-Oc0089236
Date Sampled			Oct 28, 2025	Oct 28, 2025	Oct 28, 2025	Oct 28, 2025
Test/Reference	LOR	Unit				
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)						
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	108	142	78	94
13C2-6:2 FTSA (surr.)	1	%	101	105	59	112
13C2-8:2 FTSA (surr.)	1	%	93	94	53	111
13C2-10:2 FTSA (surr.)	1	%	82	111	63	102
PFASs Summations						
Sum (PFHxS + PFOS)*	0.01	ug/L	0.06	1.7	< 0.01	< 0.01
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	0.12	1.95	< 0.05	< 0.05
Sum of PFASs (n=30)*	0.1	ug/L	0.1	2.0	< 0.1	< 0.1

Client Sample ID			DW4	BLANK 2	BLANK 3
Sample Matrix			Water	Water	Water
Eurofins Sample No.			S25-Oc0089237	S25-Oc0089238	S25-Oc0089239
Date Sampled			Oct 28, 2025	Oct 23, 2025	Oct 28, 2025
Test/Reference	LOR	Unit			
Perfluoroalkyl carboxylic acids (PFCAs)					
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	0.02	< 0.01	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.02	< 0.01	< 0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	87	57	62
13C5-PFPeA (surr.)	1	%	111	68	66
13C5-PFHxA (surr.)	1	%	101	65	72
13C4-PFHpA (surr.)	1	%	101	61	70
13C8-PFOA (surr.)	1	%	115	69	73
13C5-PFNA (surr.)	1	%	98	67	71
13C6-PFDA (surr.)	1	%	93	70	68
13C2-PFUnDA (surr.)	1	%	107	64	69
13C2-PFDoDA (surr.)	1	%	112	63	73
13C2-PFTeDA (surr.)	1	%	98	70	70

Client Sample ID			DW4	BLANK 2	BLANK 3
Sample Matrix			Water	Water	Water
Eurofins Sample No.			S25-Oc0089237	S25-Oc0089238	S25-Oc0089239
Date Sampled			Oct 28, 2025	Oct 23, 2025	Oct 28, 2025
Test/Reference	LOR	Unit			
Perfluoroalkyl sulfonamido substances					
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	100	69	67
D3-N-MeFOSA (surr.)	1	%	100	62	65
D5-N-EtFOSA (surr.)	1	%	99	67	75
D7-N-MeFOSE (surr.)	1	%	90	61	68
D9-N-EtFOSE (surr.)	1	%	93	60	66
D5-N-EtFOSAA (surr.)	1	%	94	56	64
D3-N-MeFOSAA (surr.)	1	%	103	72	73
Perfluoroalkyl sulfonic acids (PFSAs)					
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	0.02	< 0.01	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]0.04	< 0.01	< 0.01
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	[N09]0.02	< 0.01	< 0.01
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	117	67	76
18O2-PFHxS (surr.)	1	%	115	76	80
13C8-PFOS (surr.)	1	%	110	73	80
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)					
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	103	78	71
13C2-6:2 FTSA (surr.)	1	%	105	61	74
13C2-8:2 FTSA (surr.)	1	%	101	71	81
13C2-10:2 FTSA (surr.)	1	%	97	69	63
PFASs Summations					
Sum (PFHxS + PFOS)*	0.01	ug/L	0.06	< 0.01	< 0.01
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	0.12	< 0.05	< 0.05
Sum of PFASs (n=30)*	0.1	ug/L	0.1	< 0.1	< 0.1

Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description	Testing Site	Extracted	Holding Time
Per- and Polyfluoroalkyl Substances (PFASs)			
Perfluoroalkyl carboxylic acids (PFCAs)	Sydney	Nov 03, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonamido substances	Sydney	Nov 03, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonic acids (PFSAs)	Sydney	Nov 03, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)	Sydney	Nov 03, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			

eurofins

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Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: RICHMOND
Project ID: 2301034G

Order No.: PO6961
Report #: 1285891
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Oct 29, 2025 4:40 PM
Due: Nov 5, 2025
Priority: 5 Day
Contact Name: Paula Del Prete

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail						Per- and Polyfluoroalkyl Substances (PFASs)
Sydney Laboratory - NATA # 1261 Site # 18217						X
External Laboratory						
No	**Sample ID**	**Sample Date**	**Sampling Time**	**Matrix**	**LAB ID**	
1	MW03-S	Oct 28, 2025		Water	S25-Oc0089229	X
2	MW11	Oct 28, 2025		Water	S25-Oc0089230	X
3	MW02	Oct 28, 2025		Water	S25-Oc0089231	X
4	BH101i	Oct 28, 2025		Water	S25-Oc0089232	X
5	BH111	Oct 28, 2025		Water	S25-Oc0089233	X
6	BH105	Oct 28, 2025		Water	S25-Oc0089234	X
7	BH101G	Oct 28, 2025		Water	S25-Oc0089235	X
8	R3	Oct 28, 2025		Water	S25-Oc0089236	X
9	DW4	Oct 28, 2025		Water	S25-Oc0089237	X
10	BLANK 2	Oct 23, 2025		Water	S25-Oc0089238	X
11	BLANK 3	Oct 28, 2025		Water	S25-Oc0089239	X
12	TW4	Oct 28, 2025		Water	S25-Oc0089240	X
Test Counts						12

Internal Quality Control Review and Glossary

General

1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follow guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013. They are included in this QC report where applicable. Additional QC data may be available on request.
2. Unless otherwise stated, all soil/sediment/solid results are reported on a dry weight basis.
3. Unless otherwise stated, all biota/food results are reported on a wet weight basis on the edible portion.
4. For CEC results where the sample's origin is unknown or environmentally contaminated, the results should be used advisedly.
5. Actual LORs are matrix dependent. Quoted LORs may be raised where sample extracts are diluted due to interferences.
6. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds where annotated.
7. SVOC analysis on waters is performed on homogenised, unfiltered samples unless noted otherwise.
8. Samples were analysed on an 'as received' basis.
9. Information identified in this report with **blue** colour indicates data provided by customers that may have an impact on the results.
10. This report replaces any interim results previously issued.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours before sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the sampling date; therefore, compliance with these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether, the holding time is seven days; however, for all other VOCs, such as BTEX or C6-10 TRH, the holding time is 14 days.

Units

mg/kg: milligrams per kilogram	**mg/L:** milligrams per litre	**ppm:** parts per million
µg/L: micrograms per litre	**ppb:** parts per billion	**%:** Percentage
org/100 mL: Organisms per 100 millilitres	**NTU:** Nephelometric Turbidity Units	**MPN/100 mL:** Most Probable Number of organisms per 100 millilitres
CFU: Colony Forming Unit	**Colour:** Pt-Co Units (CU)	

Terms

APHA	American Public Health Association
CEC	Cation Exchange Capacity
COC	Chain of Custody
CP	Client Parent - QC was performed on samples pertaining to this report
CRM	Certified Reference Material (ISO17034) - reported as percent recovery.
Dry	Where moisture has been determined on a solid sample, the result is expressed on a dry weight basis.
Duplicate	A second piece of analysis from the same sample and reported in the same units as the result to show comparison.
LOR	Limit of Reporting.
LCS	Laboratory Control Sample - reported as percent recovery.
Method Blank	In the case of solid samples, these are performed on laboratory-certified clean sands and in the case of water samples, these are performed on de-ionised water.
NCP	Non-Client Parent - QC performed on samples not pertaining to this report, QC represents the sequence or batch that client samples were analysed within.
RPD	Relative Percent Difference between two Duplicate pieces of analysis.
SPIKE	Addition of the analyte to the sample and reported as percentage recovery.
SRA	Sample Receipt Advice
Surr - Surrogate	The addition of a similar compound to the analyte target is reported as percentage recovery. See below for acceptance criteria.
TBTO	Tributyltin oxide (*bis*-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment; however, free tributyltin was measured, and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits.
TCLP	Toxicity Characteristic Leaching Procedure
TEQ	Toxic Equivalency Quotient or Total Equivalence
QSM	US Department of Defense Quality Systems Manual Version 6.0
US EPA	United States Environmental Protection Agency
WA DWER	Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should only be used as a guide and may be different when site-specific Sampling Analysis and Quality Plan (SAQP) have been implemented.

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is ≤30%; however, the following acceptance guidelines are equally applicable:

Results <10 times the LOR:	No Limit
Results between 10-20 times the LOR:	RPD must lie between 0-50%
Results >20 times the LOR:	RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range, not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 – 150%, VOC recoveries 50 – 150%

PFAS field samples containing surrogate recoveries above the QC limit designated in QSM 6.0, where no positive PFAS results have been reported or reviewed, and no data was affected.

QC Data General Comments

1. Where a result is reported as less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown are not data from your samples.
3. pH and Free Chlorine analysed in the laboratory - Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time.Analysis will begin as soon as possible after sample receipt.
4. Recovery Data (Spikes & Surrogates) - where chromatographic interference does not allow the determination of recovery, the term "INT" appears against that analyte.
5. For Matrix Spikes and LCS results, a dash "-" in the report means that the specific analyte was not added to the QC sample.
6. Duplicate RPDs are calculated from raw analytical data; thus, it is possible to have two sets of data.

Quality Control Results

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Method Blank							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	ug/L	< 0.05			0.05	Pass	
Perfluoropentanoic acid (PFPeA)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanoic acid (PFHxA)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanoic acid (PFHpA)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanoic acid (PFOA)	ug/L	< 0.01			0.01	Pass	
Perfluorononanoic acid (PFNA)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanoic acid (PFDA)	ug/L	< 0.01			0.01	Pass	
Perfluoroundecanoic acid (PFUnDA)	ug/L	< 0.01			0.01	Pass	
Perfluorododecanoic acid (PFDoDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotridecanoic acid (PFTrDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotetradecanoic acid (PFTeDA)	ug/L	< 0.01			0.01	Pass	
Method Blank							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	ug/L	< 0.05			0.05	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	ug/L	< 0.05			0.05	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	ug/L	< 0.05			0.05	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	ug/L	< 0.05			0.05	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	ug/L	< 0.05			0.05	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	ug/L	< 0.05			0.05	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	ug/L	< 0.05			0.05	Pass	
Method Blank							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	ug/L	< 0.01			0.01	Pass	
Perfluorononanesulfonic acid (PFNS)	ug/L	< 0.01			0.01	Pass	
Perfluoropropanesulfonic acid (PFPrS)	ug/L	< 0.01			0.01	Pass	
Perfluoropentanesulfonic acid (PFPeS)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanesulfonic acid (PFHxS)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanesulfonic acid (PFOS)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanesulfonic acid (PFDS)	ug/L	< 0.01			0.01	Pass	
Method Blank							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	ug/L	< 0.05			0.05	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	ug/L	< 0.01			0.01	Pass	
Method Blank							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	ug/L	< 0.05			0.05	Pass	
Perfluoropentanoic acid (PFPeA)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanoic acid (PFHxA)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanoic acid (PFHpA)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanoic acid (PFOA)	ug/L	< 0.01			0.01	Pass	
Perfluorononanoic acid (PFNA)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanoic acid (PFDA)	ug/L	< 0.01			0.01	Pass	
Perfluoroundecanoic acid (PFUnDA)	ug/L	< 0.01			0.01	Pass	
Perfluorododecanoic acid (PFDoDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotridecanoic acid (PFTrDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotetradecanoic acid (PFTeDA)	ug/L	< 0.01			0.01	Pass	

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Method Blank							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	ug/L	< 0.05			0.05	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	ug/L	< 0.05			0.05	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	ug/L	< 0.05			0.05	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	ug/L	< 0.05			0.05	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	ug/L	< 0.05			0.05	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	ug/L	< 0.05			0.05	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	ug/L	< 0.05			0.05	Pass	
Method Blank							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	ug/L	< 0.01			0.01	Pass	
Perfluorononanesulfonic acid (PFNS)	ug/L	< 0.01			0.01	Pass	
Perfluoropropanesulfonic acid (PFPrS)	ug/L	< 0.01			0.01	Pass	
Perfluoropentanesulfonic acid (PFPeS)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanesulfonic acid (PFHxS)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanesulfonic acid (PFOS)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanesulfonic acid (PFDS)	ug/L	< 0.01			0.01	Pass	
Method Blank							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	ug/L	< 0.05			0.05	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	ug/L	< 0.01			0.01	Pass	
LCS - % Recovery							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	%	108			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	%	121			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	%	118			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	%	117			50-150	Pass	
Perfluorooctanoic acid (PFOA)	%	122			50-150	Pass	
Perfluorononanoic acid (PFNA)	%	108			50-150	Pass	
Perfluorodecanoic acid (PFDA)	%	123			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	%	111			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	%	126			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	%	124			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	%	117			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	%	106			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	%	109			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	%	106			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	%	120			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	%	107			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	%	114			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	%	122			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	%	100			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	%	111			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	%	99			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	%	90			50-150	Pass	

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Perfluorohexanesulfonic acid (PFHxS)	%	100			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	%	114			50-150	Pass	
Perfluorooctanesulfonic acid (PFOS)	%	96			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	%	98			50-150	Pass	
LCS - % Recovery							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	%	100			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	%	124			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	%	97			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	%	121			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	%	100			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	%	106			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	%	119			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	%	122			50-150	Pass	
Perfluorooctanoic acid (PFOA)	%	122			50-150	Pass	
Perfluorononanoic acid (PFNA)	%	108			50-150	Pass	
Perfluorodecanoic acid (PFDA)	%	124			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	%	112			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	%	113			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	%	101			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	%	109			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	%	123			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	%	106			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	%	99			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	%	104			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	%	104			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	%	94			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	%	109			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	%	96			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	%	98			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	%	86			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	%	84			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	%	99			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	%	101			50-150	Pass	
Perfluorooctanesulfonic acid (PFOS)	%	98			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	%	96			50-150	Pass	
LCS - % Recovery							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	%	122			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	%	102			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	%	91			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	%	115			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Spike - % Recovery									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1					
Perfluorobutanoic acid (PFBA)	S25-Oc0094963	NCP	%	105			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	S25-Oc0094963	NCP	%	120			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	S25-Oc0094963	NCP	%	124			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	S25-Oc0094963	NCP	%	118			50-150	Pass	
Perfluorooctanoic acid (PFOA)	S25-Oc0094963	NCP	%	119			50-150	Pass	
Perfluorononanoic acid (PFNA)	S25-Oc0094963	NCP	%	113			50-150	Pass	
Perfluorodecanoic acid (PFDA)	S25-Oc0094963	NCP	%	143			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	S25-Oc0094963	NCP	%	116			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	S25-Oc0094963	NCP	%	117			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	S25-Oc0094963	NCP	%	127			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S25-Oc0094963	NCP	%	114			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonamido substances				Result 1					
Perfluorooctane sulfonamide (FOSA)	S25-Oc0094963	NCP	%	107			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S25-Oc0094963	NCP	%	122			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S25-Oc0094963	NCP	%	104			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S25-Oc0094963	NCP	%	119			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S25-Oc0094963	NCP	%	114			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S25-Oc0094963	NCP	%	107			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S25-Oc0094963	NCP	%	119			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1					
Perfluorobutanesulfonic acid (PFBS)	S25-Oc0094963	NCP	%	111			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	S25-Oc0094963	NCP	%	100			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S25-Oc0094963	NCP	%	99			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S25-Oc0094963	NCP	%	92			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S25-Oc0094963	NCP	%	108			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S25-Oc0094963	NCP	%	106			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	S25-Oc0094963	NCP	%	104			50-150	Pass	
Spike - % Recovery									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1					
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S25-Oc0094963	NCP	%	128			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S25-Oc0094963	NCP	%	125			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S25-Oc0094963	NCP	%	100			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S25-Oc0094963	NCP	%	125			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluorobutanoic acid (PFBA)	S25-No0001650	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Perfluoropentanoic acid (PFPeA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanoic acid (PFHxA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroheptanoic acid (PFHpA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanoic acid (PFOA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanoic acid (PFNA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanoic acid (PFDA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroundecanoic acid (PFUnDA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorododecanoic acid (PFDoDA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotridecanoic acid (PFTrDA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
Perfluorooctane sulfonamide (FOSA)	S25-No0001650	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S25-No0001650	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S25-No0001650	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S25-No0001650	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S25-No0001650	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S25-No0001650	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S25-No0001650	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1	Result 2	RPD			
Perfluorobutanesulfonic acid (PFBS)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanesulfonic acid (PFNS)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S25-No0001650	NCP	ug/L	0.02	0.02	13	30%	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanesulfonic acid (PFOS)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanesulfonic acid (PFDS)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Duplicate									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1	Result 2	RPD			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S25-No0001650	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S25-No0001650	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluorobutanoic acid (PFBA)	N25-Oc0090476	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Perfluoropentanoic acid (PFPeA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanoic acid (PFHxA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroheptanoic acid (PFHpA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanoic acid (PFOA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanoic acid (PFNA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanoic acid (PFDA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroundecanoic acid (PFUnDA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorododecanoic acid (PFDoDA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotridecanoic acid (PFTrDA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotetradecanoic acid (PFTeDA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
Perfluorooctane sulfonamide (FOSA)	N25-Oc0090476	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	N25-Oc0090476	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	N25-Oc0090476	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	N25-Oc0090476	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	N25-Oc0090476	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	N25-Oc0090476	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	N25-Oc0090476	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1	Result 2	RPD			
Perfluorobutanesulfonic acid (PFBS)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanesulfonic acid (PFNS)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropropanesulfonic acid (PFPrS)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropentanesulfonic acid (PFPeS)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanesulfonic acid (PFHxS)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanesulfonic acid (PFOS)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanesulfonic acid (PFDS)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Duplicate									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1	Result 2	RPD			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	N25-Oc0090476	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	N25-Oc0090476	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Comments

Sample Integrity

Custody Seals Intact (if used)	N/A
Attempt to Chill was evident	Yes
Sample correctly preserved	Yes
Appropriate sample containers have been used	Yes
Sample containers for volatile analysis received with minimal headspace	Yes
Samples received within HoldingTime	Yes
Some samples have been subcontracted	No

Qualifier Codes/Comments

Code	Description
N09	Quantification of linear and branched isomers has been conducted as a single total response using the relative response factor for the corresponding linear/branched standard.
N11	Isotope dilution is used for calibration of each native compound for which an exact labelled analogue is available (Isotope Dilution Quantitation). The isotopically labelled analogues allow identification and recovery correction of the concentration of the associated native PFAS compounds.
N15	Where the native PFAS compound does not have labelled analogue then the quantification is made using the Extracted Internal Standard Analyte with the closest retention time to the analyte and no recovery correction has been made (Internal Standard Quantitation).

Authorised by:

Nileshni Goundar	Analytical Services Manager
Roopesh Rangarajan	Senior Analyst-PFAS



Glenn Jackson
Managing Director

Final Report – this report replaces any previously issued Report

- Indicates Not Requested

* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here.

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Sample Receipt Advice

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Contact name:	Paula Del Prete
Project name:	RICHMOND
Project ID:	2301034G
Turnaround time:	5 Day
Date/Time received	Oct 29, 2025 4:40 PM
Eurofins reference	1285891

Sample Information

✓	A detailed list of analytes logged into our LIMS, is included in the attached summary table.
✓	Sample Temperature of chilled sample on the batch as recorded by Eurofins Sample Receipt : -.4 degrees Celsius.
✓	All samples have been received as described on the above COC.
✓	COC has been completed correctly.
✓	Attempt to chill was evident.
✓	Appropriately preserved sample containers have been used.
✓	All samples were received in good condition.
✓	Samples have been provided with adequate time to commence analysis in accordance with the relevant holding times.
✓	Appropriate sample containers have been used.
✓	Sample containers for volatile analysis received with zero headspace.
X	Split sample sent to requested external lab.
X	Some samples have been subcontracted.
N/A	Custody Seals intact (if used).

Notes

Contact

If you have any questions with respect to these samples, please contact your Analytical Services Manager:

Bonnie Pu on phone : or by email: Bonnie.Pu@eurofinsanz.com

Results will be delivered electronically via email to Paula Del Prete - pdelprete@geo-logix.com.au.

Note: A copy of these results will also be delivered to the general Geo-Logix P/L email address.

1285891

Geo-Logix Pty Ltd
Building Q2, Level 3
2309/4 Daydream St
Warriewood, NSW 2102
ABN: 86 116 892 936
P: (02) 9979 1722
F: (02) 9979 1222

CHAIN OF CUSTODY

Page 1 of 1

Project Manager: Paula Del Prete, Harsha Muthu
Purchase Order No: PO6961
Contact email: pdelprete@geo-logix.com.au / hmuthu@geo-logix.com.au
Quote Reference:
Project Name: Richmond
Send Invoice to: accounts@geo-logix.com.au
Project Number: 2301034g Date Submitted: 29/10/2025
TAT required: STD

ANALYSIS REQUIRED

Lab ID	Sample ID	Date	Matrix: soil	water	air	paint, filters	other	Comments	COMPOSITE	TRH - C6 - C10	TRH - C10 - C40	VOCs	BTEXN	PAHs	PCBs	OCPs	OPPs	Phenols	Metals - M8	Metals - Lead	Metals - Specify **	TCLP	Asbestos (ID only)	Asbestos (WA DOH)	Foreign Materials	Conductivity (EC)	pH	PFAS - 30 compound		Hold	SUITE
	MW03 - S	28/10/25		X				PFAS bottle																				X		
	MW11	"		X				"																				X		
	MW02	"		X				"																				X		
	BH101i	"		X				"																				X		
	BH111	"		X				"																				X		
	BH105	"		X				"																				X		
	BH101g	"		X				"																				X		
	R3	"		X				"																				X		
	DW4	"		X				"																				X		
	Blank 2	23/10/25		X				"																				X		
	Blank 3	28/10/25		X				"																				X		

Eurofins MGT Suite Codes

B1	TRH/BTEXN
B1A	TRH/MAH
B2	TRH/BTEXN/Pb
B2A	TRH/MAH/Pb
B3	PAH/Phenols
B4	TRH/BTEXN/PAH
B4A	TRH/BTEXN/PAH/Phenols
B5	TRH/BTEXN/M7
B6	TRH/BTEXN/M8
B7	TRH/BTEXN/PAH/M8
B7A	TRH/BTEXN/PAH/Phenols/M8
B8	TRH/VOC/PAH/M8
B9	TRH/BTEXN/PAH/OCP/M8
B10	TRH/BTEXN/PAH/OCP/OPP/M8
B11	Na/K/Ca/Mg/Cl/SO_4/CO_3/HCO_3/NH_3/NO_3
B11A	B11/Alkalinity
B11B	B11/EC/TDS
B12	TRH/BTEXN/Oxygenates/Ethanol
B12A	TRH/BTEXN/Oxygenates
B13	OCP/PCB
B14	OCP/OPP
B15	OCP/OPP/PCB
B16	TDS/SO_4/CH_4/Alk/BOD/COD/HPC/CUB
B17	SO_4/NO_3/Fe++/HPC/CUB
B18	Cl-/SO_4/pH
B19	N/P/K
B20	CEC/%ESP/Ca/Mg/Na/K

Metals(circle)** As, Cd, Cr, Cu, Ni, Pb, Zn, Hg, Cr^{6+}, Cr^{3+}, Fe^{2+}, Fe^{3+}, Be, B, Al, V, Mn, Fe, Co, Se, Sr, Sn, Mo, Ag, Ba, Tl, Bi, Sb

Chain of Custody

Relinquished by: Harsha Date/Time: 29/10/25 Signature:
Received by: Simon Date/Time: 29/10 Signature:
4:40 pm

-0.2
AECH

Geo-Logix P/L
Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

NATA Accredited
Accreditation Number 1261
Site Number 1254

Accredited for compliance with ISO/IEC 17025 – Testing
NATA is a signatory to the ILAC Mutual Recognition Arrangement for the mutual recognition of the equivalence of testing, medical testing, calibration, inspection, proficiency testing scheme providers and reference materials producers reports and certificates.

Attention: **Paula Del Prete**

Report **1286742-W**
Project name RICHMOND
Project ID 2301034G
Received Date Oct 29, 2025

Client Sample ID			**TW4**
Sample Matrix			**Water**
Eurofins Sample No.			**S25-Oc0095657**
Date Sampled			**Oct 28, 2025**
Test/Reference	LOR	Unit	
Perfluoroalkyl carboxylic acids (PFCAs)			
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	0.03
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.03
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01
13C4-PFBA (surr.)	1	%	108
13C5-PFPeA (surr.)	1	%	82
13C5-PFHxA (surr.)	1	%	136
13C4-PFHpA (surr.)	1	%	140
13C8-PFOA (surr.)	1	%	141
13C5-PFNA (surr.)	1	%	111
13C6-PFDA (surr.)	1	%	[J+]156
13C2-PFUnDA (surr.)	1	%	[J+]179
13C2-PFDoDA (surr.)	1	%	[J+]187
13C2-PFTeDA (surr.)	1	%	[J+]152
Perfluoroalkyl sulfonamido substances			
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05
13C8-FOSA (surr.)	1	%	121
D3-N-MeFOSA (surr.)	1	%	[J-]37

Client Sample ID			TW4
Sample Matrix			Water
Eurofins Sample No.			S25-Oc0095657
Date Sampled			Oct 28, 2025
Test/Reference	LOR	Unit	
Perfluoroalkyl sulfonamido substances			
D5-N-EtFOSA (surr.)	1	%	[J-]44
D7-N-MeFOSE (surr.)	1	%	103
D9-N-EtFOSE (surr.)	1	%	103
D5-N-EtFOSAA (surr.)	1	%	102
D3-N-MeFOSAA (surr.)	1	%	86
Perfluoroalkyl sulfonic acids (PFSAs)			
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	0.02
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]0.04
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	[N09]0.02
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01
13C3-PFBS (surr.)	1	%	131
18O2-PFHxS (surr.)	1	%	124
13C8-PFOS (surr.)	1	%	133
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01
13C2-4:2 FTSA (surr.)	1	%	[J+]178
13C2-6:2 FTSA (surr.)	1	%	99
13C2-8:2 FTSA (surr.)	1	%	111
13C2-10:2 FTSA (surr.)	1	%	123
PFASs Summations			
Sum (PFHxS + PFOS)*	0.01	ug/L	0.06
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	0.14
Sum of PFASs (n=30)*	0.1	ug/L	0.2

Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description	Testing Site	Extracted	Holding Time
Per- and Polyfluoroalkyl Substances (PFASs)			
Perfluoroalkyl carboxylic acids (PFCAs)	Melbourne	Nov 03, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonamido substances	Melbourne	Nov 03, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonic acids (PFSAs)	Melbourne	Nov 03, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)	Melbourne	Nov 03, 2025	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			

eurofins

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ABN: 50 005 085 521

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NATA# 2377
Site# 2370 & 2554

web: www.eurofins.com.au
email: EnviroSales@eurofinsanz.com

Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: RICHMOND
Project ID: 2301034G

Order No.:
Report #: 1286742
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Oct 29, 2025 4:40 PM
Due: Nov 5, 2025
Priority: 5 Day
Contact Name: Paula Del Prete

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail						Per- and Polyfluoroalkyl Substances (PFASs)
Sydney Laboratory - NATA # 1261 Site # 18217						X
External Laboratory						
No	**Sample ID**	**Sample Date**	**Sampling Time**	**Matrix**	**LAB ID**	
1	TW4	Oct 28, 2025		Water	S25-Oc0095657	X
Test Counts						1

Internal Quality Control Review and Glossary

General

1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follow guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013. They are included in this QC report where applicable. Additional QC data may be available on request.
2. Unless otherwise stated, all soil/sediment/solid results are reported on a dry weight basis.
3. Unless otherwise stated, all biota/food results are reported on a wet weight basis on the edible portion.
4. For CEC results where the sample's origin is unknown or environmentally contaminated, the results should be used advisedly.
5. Actual LORs are matrix dependent. Quoted LORs may be raised where sample extracts are diluted due to interferences.
6. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds where annotated.
7. SVOC analysis on waters is performed on homogenised, unfiltered samples unless noted otherwise.
8. Samples were analysed on an 'as received' basis.
9. Information identified in this report with **blue** colour indicates data provided by customers that may have an impact on the results.
10. This report replaces any interim results previously issued.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours before sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the sampling date; therefore, compliance with these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether, the holding time is seven days; however, for all other VOCs, such as BTEX or C6-10 TRH, the holding time is 14 days.

Units

mg/kg: milligrams per kilogram	**mg/L:** milligrams per litre	**ppm:** parts per million
µg/L: micrograms per litre	**ppb:** parts per billion	**%:** Percentage
org/100 mL: Organisms per 100 millilitres	**NTU:** Nephelometric Turbidity Units	**MPN/100 mL:** Most Probable Number of organisms per 100 millilitres
CFU: Colony Forming Unit	**Colour:** Pt-Co Units (CU)	

Terms

APHA	American Public Health Association
CEC	Cation Exchange Capacity
COC	Chain of Custody
CP	Client Parent - QC was performed on samples pertaining to this report
CRM	Certified Reference Material (ISO17034) - reported as percent recovery.
Dry	Where moisture has been determined on a solid sample, the result is expressed on a dry weight basis.
Duplicate	A second piece of analysis from the same sample and reported in the same units as the result to show comparison.
LOR	Limit of Reporting.
LCS	Laboratory Control Sample - reported as percent recovery.
Method Blank	In the case of solid samples, these are performed on laboratory-certified clean sands and in the case of water samples, these are performed on de-ionised water.
NCP	Non-Client Parent - QC performed on samples not pertaining to this report, QC represents the sequence or batch that client samples were analysed within.
RPD	Relative Percent Difference between two Duplicate pieces of analysis.
SPIKE	Addition of the analyte to the sample and reported as percentage recovery.
SRA	Sample Receipt Advice
Surr - Surrogate	The addition of a similar compound to the analyte target is reported as percentage recovery. See below for acceptance criteria.
TBTO	Tributyltin oxide (*bis*-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment; however, free tributyltin was measured, and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits.
TCLP	Toxicity Characteristic Leaching Procedure
TEQ	Toxic Equivalency Quotient or Total Equivalence
QSM	US Department of Defense Quality Systems Manual Version 6.0
US EPA	United States Environmental Protection Agency
WA DWER	Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should only be used as a guide and may be different when site-specific Sampling Analysis and Quality Plan (SAQP) have been implemented.

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is ≤30%; however, the following acceptance guidelines are equally applicable:

Results <10 times the LOR:	No Limit
Results between 10-20 times the LOR:	RPD must lie between 0-50%
Results >20 times the LOR:	RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range, not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 – 150%, VOC recoveries 50 – 150%

PFAS field samples containing surrogate recoveries above the QC limit designated in QSM 6.0, where no positive PFAS results have been reported or reviewed, and no data was affected.

QC Data General Comments

1. Where a result is reported as less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown are not data from your samples.
3. pH and Free Chlorine analysed in the laboratory - Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time.Analysis will begin as soon as possible after sample receipt.
4. Recovery Data (Spikes & Surrogates) - where chromatographic interference does not allow the determination of recovery, the term "INT" appears against that analyte.
5. For Matrix Spikes and LCS results, a dash "-" in the report means that the specific analyte was not added to the QC sample.
6. Duplicate RPDs are calculated from raw analytical data; thus, it is possible to have two sets of data.

Quality Control Results

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluorobutanoic acid (PFBA)	M25-Oc0093310	NCP	ug/L	3.0	3.1	4.0	30%	Pass	
Perfluoropentanoic acid (PFPeA)	M25-Oc0093310	NCP	ug/L	0.38	0.38	<1	30%	Pass	
Perfluorohexanoic acid (PFHxA)	M25-Oc0093310	NCP	ug/L	0.07	0.08	6.0	30%	Pass	
Perfluoroheptanoic acid (PFHpA)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanoic acid (PFOA)	M25-No0000307	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanoic acid (PFNA)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanoic acid (PFDA)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroundecanoic acid (PFUnDA)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorododecanoic acid (PFDoDA)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotridecanoic acid (PFTrDA)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotetradecanoic acid (PFTeDA)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
Perfluorooctane sulfonamide (FOSA)	M25-Oc0093310	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	M25-Oc0093310	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	M25-Oc0093310	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	M25-Oc0093310	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	M25-Oc0093310	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	M25-Oc0093310	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	M25-Oc0093310	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1	Result 2	RPD			
Perfluorobutanesulfonic acid (PFBS)	M25-Oc0093310	NCP	ug/L	0.07	0.06	9.0	30%	Pass	
Perfluorononanesulfonic acid (PFNS)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropropanesulfonic acid (PFPrS)	M25-Oc0093310	NCP	ug/L	0.91	0.87	5.0	30%	Pass	
Perfluoropentanesulfonic acid (PFPeS)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanesulfonic acid (PFHxS)	M25-No0000307	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanesulfonic acid (PFOS)	M25-No0000307	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanesulfonic acid (PFDS)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Duplicate									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1	Result 2	RPD			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	M25-No0000307	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	M25-Oc0093310	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Comments

Sample Integrity

Custody Seals Intact (if used)	N/A
Attempt to Chill was evident	Yes
Sample correctly preserved	Yes
Appropriate sample containers have been used	Yes
Sample containers for volatile analysis received with minimal headspace	Yes
Samples received within HoldingTime	Yes
Some samples have been subcontracted	No

Qualifier Codes/Comments

Code	Description
J-	If the percent recoveries were less than the lower control limit, qualify detects for the analyte in associated samples as estimated with a negative bias J-.
J+	If the percent recoveries were greater than the upper control limit, qualify detects for the analyte in associated samples as estimated with a positive bias J+.
N09	Quantification of linear and branched isomers has been conducted as a single total response using the relative response factor for the corresponding linear/branched standard.
N11	Isotope dilution is used for calibration of each native compound for which an exact labelled analogue is available (Isotope Dilution Quantitation). The isotopically labelled analogues allow identification and recovery correction of the concentration of the associated native PFAS compounds.
N15	Where the native PFAS compound does not have labelled analogue then the quantification is made using the Extracted Internal Standard Analyte with the closest retention time to the analyte and no recovery correction has been made (Internal Standard Quantitation).

Authorised by:

Nileshni Goundar	Analytical Services Manager
Dylan Jones	Senior Analyst-PFAS



Glenn Jackson
Managing Director

Final Report – this report replaces any previously issued Report

- Indicates Not Requested

* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here.

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eurofins | Environment Testing

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Sample Receipt Advice

Company name:	Geo-Logix P/L
Contact name:	Paula Del Prete
Project name:	RICHMOND
Project ID:	2301034G
Turnaround time:	5 Day
Date/Time received	Oct 29, 2025 4:40 PM
Eurofins reference	1286742

Sample Information

✓	A detailed list of analytes logged into our LIMS, is included in the attached summary table.
✓	Sample Temperature of chilled sample on the batch as recorded by Eurofins Sample Receipt : -.4 degrees Celsius.
✓	All samples have been received as described on the above COC.
✓	COC has been completed correctly.
✓	Attempt to chill was evident.
✓	Appropriately preserved sample containers have been used.
✓	All samples were received in good condition.
✓	Samples have been provided with adequate time to commence analysis in accordance with the relevant holding times.
✓	Appropriate sample containers have been used.
✓	Sample containers for volatile analysis received with zero headspace.
X	Split sample sent to requested external lab.
X	Some samples have been subcontracted.
N/A	Custody Seals intact (if used).

Notes

Contact

If you have any questions with respect to these samples, please contact your Analytical Services Manager:

Bonnie Pu on phone : or by email: Bonnie.Pu@eurofinsanz.com

Results will be delivered electronically via email to Paula Del Prete - pdelprete@geo-logix.com.au.

Note: A copy of these results will also be delivered to the general Geo-Logix P/L email address.

Geo-Logix Pty Ltd
Building Q2, Level 3
2309/4 Daydream St
Warriewood, NSW 2102
ABN: 86 116 892 936
P: (02) 9979 1722
F: (02) 9979 1222

CHAIN OF CUSTODY

Page 1 of 1

Project Manager: Paula Del Prete, Harsha Muthu
Purchase Order No: P06961
Contact email: pdelprete@geo-logix.com.au / hmuthu@geo-logix.com.au
Quote Reference:
Project Name: Richmond
Send Invoice to: accounts@geo-logix.com.au
Project Number: 2301034g **Date Submitted:** 29/10/2025
TAT required: STD

ANALYSIS REQUIRED

Lab ID	Sample ID	Date	Matrix: soil	water	air	paint, filters	other	Comments	COMPOSITE	TRH - C6 - C10	TRH - C10 - C40	VOCs	BTEXN	PAHs	PCBs	OCPs	OPPs	Phenols	Metals - M8	Metals - Lead	Metals - Specify **	TCLP	Asbestos (ID only)	Asbestos (WA DOH)	Foreign Materials	Conductivity (EC)	pH	PFAS - 30 compound		Hold	SUITE
	TW4			X				PFAS bottle																				X		

Eurofins MGT Suite Codes

B1	TRH/BTEXN
B1A	TRH/MAH
B2	TRH/BTEXN/Pb
B2A	TRH/MAH/Pb
B3	PAH/Phenols
B4	TRH/BTEXN/PAH
B4A	TRH/BTEXN/PAH/Phenols
B5	TRH/BTEXN/M7
B6	TRH/BTEXN/M8
B7	TRH/BTEXN/PAH/M8
B7A	TRH/BTEXN/PAH/Phenols/M8
B8	TRH/VOC/PAH/M8
B9	TRH/BTEXN/PAH/OCP/M8
B10	TRH/BTEXN/PAH/OCP/OPP/M8
B11	Na/K/Ca/Mg/Cl/SO_4/CO_3/HCO_3/NH_3/NO_3
B11A	B11/Alkalinity
B11B	B11/EC/TDS
B12	TRH/BTEXN/Oxygenates/Ethanol
B12A	TRH/BTEXN/Oxygenates
B13	OCP/PCB
B14	OCP/OPP
B15	OCP/OPP/PCB
B16	TDS/SO_4/CH_4/Alk/BOD/COD/HPC/CUB
B17	SO_4/NO_3/Fe++/HPC/CUB
B18	Cl-/SO_4/pH
B19	N/P/K
B20	CEC/%ESP/Ca/Ma/Na/K

Metals(circle)** As, Cd, Cr, Cu, Ni, Pb, Zn, Hg, Cr^{6+}, Cr^{3+}, Fe^{2+}, Fe^{3+}, Be, B, Al, V, Mn, Fe, Co, Se, Sr, Sn, Mo, Ag, Ba, Tl, Bi, Sb

Chain of Custody

Relinquished by: Harsha Date/Time: 29/10/25 Signature:
Received by: Simon Date/Time: 29/10 Signature:
Brendan 9:40
-0.4
CH

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