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30 July 2026

Mr Johan Haaring
Western Sydney University
Hawkesbury Campus
j.haaring@westernsydney.edu.au

SUBJECT: **WSU EMP Surface Water Sampling 2026 – Dry Event 1**

SITE: **2 College Street, Richmond NSW 2753**

Dear Johan,

Geo-Logix conducted surface water monitoring for per- and polyfluoroalkyl substances (PFAS) within the PFAS Management Area (PMA) at the Hawkesbury Campus, Richmond NSW, in accordance with the *Environmental Management Plan* (2301034hEMPDraftV02.30.3.26) (EMP).

The EMP requires four rounds of surface water monitoring, comprising two dry-weather events and two wet-weather events, to assess whether PFAS from the PMA is contributing to Hillside Dam at concentrations exceeding the relevant default guideline values (DGVs).

This letter report presents the results of the first dry-weather monitoring event undertaken as part of the EMP monitoring program. Surface water samples were collected from the nominated monitoring locations and analysed for PFAS and total suspended solids (TSS). Monitoring was undertaken in general accordance with the requirements of the EMP.

SCOPE OF WORKS

The scope of works for this assessment include:

- Site visit to geo-reference sample locations as per the EMP and assess access logistics.
- Collection of surface water samples from 12 fixed sampling sites.
- Analysis of primary and quality control samples for PFAS and Total Suspended Solids by a NATA-accredited laboratory.
- Provision of this letter report.

A summary of sample locations is presented in the table below. Sample locations are presented on Figure 2. A photographic log of sample locations has been included in Attachment A.

Sample ID	Location	Purpose
SW01	Upgradient PMA – MDC	Ambient/upgradient reference within the local drainage network
SW02	WDC discharge from PMA	Measures PFAS exported from the western drainage pathway
SW03	MDC between WDC and EDC	Assesses PFAS in MDC after WDC contribution but prior to EDC input
SW04	EDC discharge from PMA	Measures PFAS exported from the eastern drainage pathway
SW05	MDC immediately downgradient of WDC + EDC	Assesses combined PMA drainage load following confluence of the two source channels
SW06	MDC downgradient of confluence with Native Seed Centre drainage channel	Assesses any dilution or additional contribution from external drainage input
SW07	Horticulture Dam drainage channel confluence with MDC	Assesses interaction with the Horticulture Dam drainage pathway
SW08	MDC	Assesses any dilution or additional contribution from Horticulture Dam drainage pathway
SW09	MDC	Intermediate downgradient pathway monitoring location
SW10	MDC	Final pathway monitoring point immediately prior to entry into POC 1
DW3	Hillside Dam	POC 1
DW4	Horticultural Dam	Discharges into MDC

MDC: Main Drainage Channel

PMA: PFAS Management Area

EDC: PMA Eastern Drainage Channel

WDC: PMA Western Drainage Channel

POC: Point of Compliance

SAMPLING METHODOLOGY

Surface Water

Surface water samples were collected directly into laboratory-supplied, PFAS-compatible, Teflon-free sample containers. Where practical, the containers were submerged approximately 100 mm below the water surface, clear of aquatic vegetation and above the channel bed to minimise the disturbance of bottom sediments. Samples were generally collected from the centre of the flow channel. Samples from DW03 and DW04 were collected adjacent to the bank due to the size and depth of the dams, while SW10 was sampled from the bank because dense vegetation restricted access to the centre of the channel.

Surface water within the drainage channels was shallow during the monitoring event, consistent with the prevailing dry conditions. Where required, field personnel carefully entered the channel to collect samples from the main flow path while minimising disturbance to bed sediments. Field water-quality parameters were measured using a calibrated water-quality meter immersed in a separate sample container. The corresponding field sampling logs and instrument calibration certificate are provided in Attachment B and Attachment C.

Disposable nitrile gloves were changed between each sampling location to minimise the potential for cross-contamination. Following collection, sample containers were labelled, immediately placed on ice in an insulated cooler and transported under chain-of-custody documentation to Eurofins, a NATA-accredited laboratory, for analysis of PFAS and total suspended solids.

Surface water sample logs are presented in Attachment B. Equipment calibration certificates are presented in Attachment C.

Quality Assurance

Quality control (QC) sampling was undertaken in general accordance with specifications outlined in Section 7.7 and 8.8 of the EMP and PFAS NEPM 3.1. Any exceptions are commented on in the assessment results. Field QC samples were collected and included the following:

Sample Identification	Sample Type	Sample Matrix	Rate of Collection
Dup 1	Field duplicate of DW3	Surface water	1 in 10 samples
Trip 1	Field triplicate of DW3	Surface water	1 in 10 samples
BLANK1	Field Reagent Blank	Water	1 per sampling batch

The laboratory internal QC procedures are consistent with NEPM policy on laboratory analysis of contaminated waters and sediments.

Field quality assurance procedures included:

- Disposable nitrile gloves were changed between each sample location. Nitrile gloves are appropriate for use in PFAS sampling.

ASSESSMENT CRITERIA

The draft EMP specifies that surface water monitoring results are to be assessed against the DGVs for freshwater ecosystems published in the PFAS National Environmental Management Plan (NEMP) Version 3.0. Since approval of the EMP, PFAS NEMP Version 3.1 has been published. Accordingly, version 2 of the draft EMP has been issued and the results presented in this letter have been assessed against the updated DGVs in PFAS NEMP Version 3.1, reflecting the current national guidance available at the time of reporting.

The DGVs used for this monitoring program are:

- PFOS: 0.9 µg/L
- PFOA: 220 µg/L

The DGVs represent the 95% freshwater ecosystem protection guideline value.

These values are used as benchmark trigger values for interpreting PFAS concentrations in surface water. They do not represent compliance limits or acceptable concentrations. Rather they are applied as reference values to assist in assessing monitoring results within the context of the CSM.

SUMMARY OF RESULTS

Surface Water Parameters

Surface water parameters are comprised in the following table.

Sample	pH	EC (µS/cm)	DO (mg/L)	Redox (MV)	Temp (°C)
SW01	n/a*				
SW02	n/a*				
SW03	6.61	265.6	89.6	1.91	12.9
SW04	7.13	906	87.3	7.39	12.8
SW05	6.86	695	21.6	3.30	10.6
SW06	6.60	223	92.7	5.70	11.8
SW07	6.10	223.6	109.6	3.5	10.3
SW08	6.97	223.4	97.7	4.54	10.2
SW09	6.72	224	99.7	11.95	10.1
SW10	6.59	219.7	88.6	6.80	10.1
DW3	7.10	539	88.5	6.27	13.7
DW4	7.98	645	84.1	8.08	13.9

*Samples were not able to be collected from SW01 and SW02 because no surface water was present at the time of the sampling event.

Complete results are included in the surface water sample logs in Attachment B.

Laboratory Results

A summary of analytical data is presented in Tables 1 and 2, and Figures 3 and 4. Laboratory analytical reports are presented in Attachment D.

PFOS

PFOS was detected above the 95% EWQG in sample SW04 (Table 1)

PFOS concentrations in surface water drainage network samples ranged from:

- 0.04 to 1.1 µg/L

PFOS concentrations in surface water samples collected from Hillside and Horticultural Dams were 0.03 and 0.02 ug/L respectively.

PFOA

PFOA was not detected in any surface water samples in excess of the Assessment Criteria.

TSS

Total Suspended Solids ranged from < 5 to 1,400 in all samples analysed (Table 2).

TSS were analysed to support the interpretation of PFAS results.

QA/QC ASSESSMENT

All field duplicate/ triplicate results were within the adopted acceptance criteria of 30-50% (AS4482.1) relative percent difference (RPD) for surface water samples, except for a small number of results that were considered acceptable given the reported concentrations were close to the laboratory limit of reporting (LOR).

PFAS were not detected in the laboratory blank, indicating that the sample collection and handling procedures were sufficient to prevent cross-contamination (Table 1).

A summary of Laboratory QA/QC data is presented on the following table.

Report #	Analysis Within Holding Time	Surrogate Recovery	Lab. Duplicate RPD %	Lab. Matrix Spike Recovery	Lab. Control Sample	Lab. Method Blank
1384514-W	✓	X	✓	✓	✓	✓
1384515-W	✓	X	✓	✓	✓	✓
✓ = Pass X = Fail						

Quality Assurance Criteria	Quality Control Criteria
Holding Times	**Accuracy**
PFAS – 28 days	Matrix spike, control sample and surrogate recovery 50-150%
	Precision
	Method Blank - Not detected Duplicate - No limit (<10xEQL), 0-50% (10-20xEQL), 0-30% (>20xEQL)

A limited number of PFAS surrogate recoveries were outside the expected acceptance range. None related to PFOS or PFOA, the analytes for which assessment criteria were adopted, and the departures are therefore not considered to have materially affected interpretation of the results.

The TSS field duplicate result was also outside the expected acceptance range. However, the result met the laboratory's internal quality-control acceptance criteria, as indicated by qualifier Q15.

Geo-Logix considers the analytical dataset to be of acceptable quality and suitable for interpretation in the context of this investigation.

CONCLUSIONS

This letter presents the findings of the first dry-weather surface water monitoring event undertaken in accordance with the EMP. The results will be considered together with the remaining monitoring events and interpreted in the Surface Water Monitoring Report prepared at completion of the monitoring program.

Please feel free to contact Geo-Logix on (02) 9979 1722 should you have any further questions regarding this letter.

Yours sincerely,



Zahra Ginwala
BSc ENV
Environmental Geologist



Jenna Maltman
BSc MEnvMgt
Senior Scientist

FIGURES

Figure 1: Site Location

Figure 2: Surface Water Sampling Map

Figure 3: PFOS SAC Exceedances Map

Figure 4: Sum of PFAS Concentrations

TABLES

Table 1: Summary of Surface Water Analytical Data – Per- and Polyfluorinated Alkyl Substances

Table 2: Summary of Surface Water Analytical Data – Total Suspended Solids

ATTACHMENTS

Attachment A: Photographic Log

Attachment B: Sampling Logs

Attachment C: Calibration Certificates

Attachment D: Laboratory Report

LIMITATIONS

This report should be read in full, and no conclusion or other section of the report may be used or relied on in isolation or taken as representative of the report as a whole. No responsibility is accepted by Geo-Logix, and any duty of care that may arise but for this statement is excluded, in relation to any use of any part of this report other than on this basis.

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a) suitability of the Site for any specific use, or category of use, or

b) potential statutory requirements for remediation, if any, of the Site,

c) approvals, if any, that may be needed in respect of any use or category of use, or

d) level of remediation, if any, that is warranted to render the Site suitable for any specific use, or category of use, or

e) level of ongoing monitoring of Site conditions, if any, that is required in respect of any specific use, or category of use, or

f) presence, extent or absence of any substance in, on or under the Site,

other than as expressly stated in this report.

The conclusions stated in this report are based solely on the information, Scope of Works, analysis and data that are stated or expressly referred to in this report.

To the extent that the information and data relied upon to prepare this report has been conveyed to Geo-Logix by the Client or third parties orally or in the form of documents, Geo-Logix has assumed that the information and data are completely accurate and has not sought independently to verify the accuracy of the information or data. Geo-Logix assumes no responsibility or duty of care in respect of any errors or omissions in the information or data provided to it.

Without limiting the paragraph above, where laboratory tests have been carried out by others on Geo-Logix' behalf, the tests are reproduced in this report on the assumption that the tests are accurate. Geo-Logix has not sought independently to verify the accuracy of those tests and assumes no responsibility in respect of them.

Geo-Logix assumes no responsibility in respect of any changes in the condition of the Site which have occurred since the time when Geo-Logix gathered data and/or took samples from the Site on its site inspections dated **15 July 2026.**

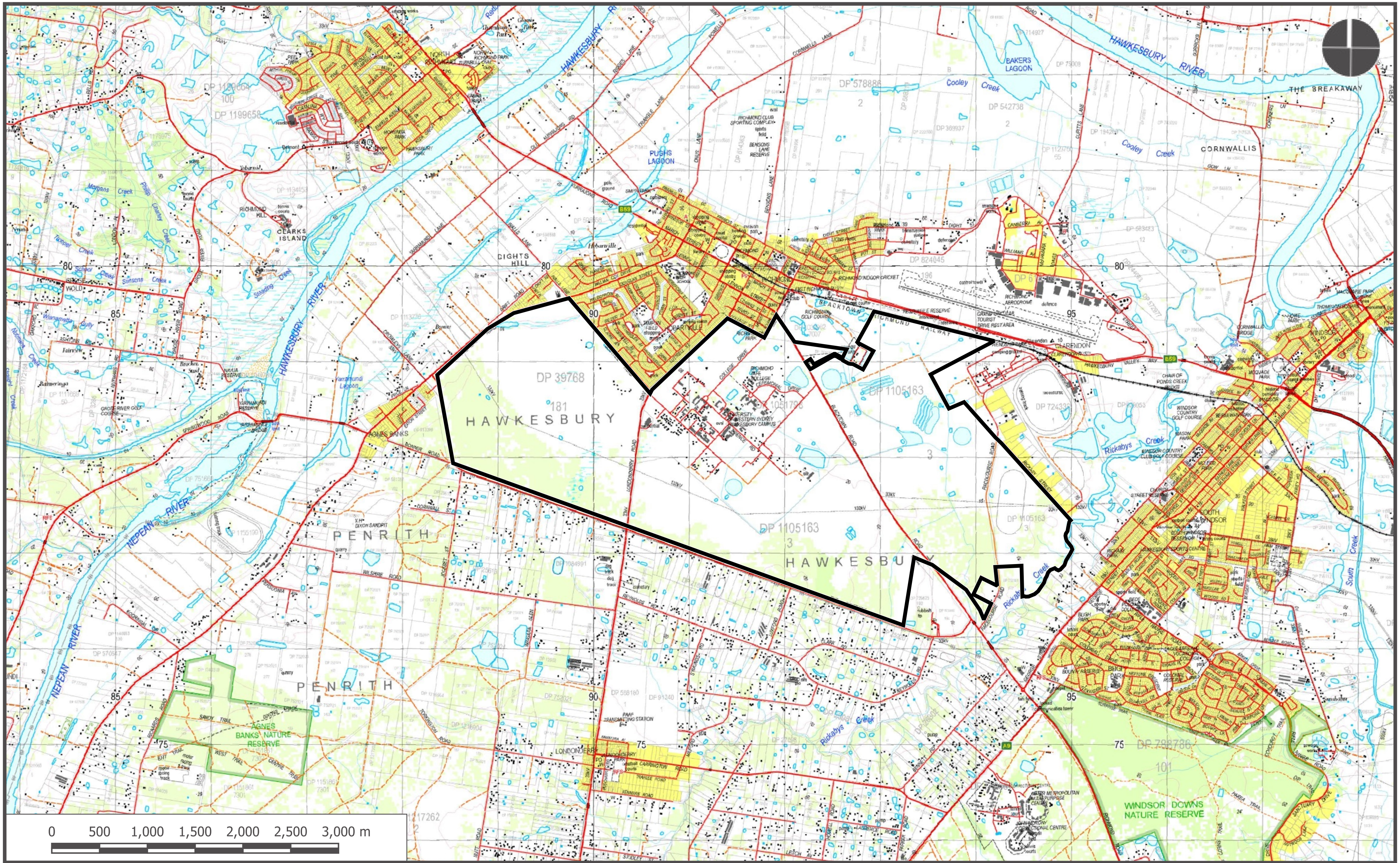
Where the Scope of Works does not include offsite investigations, Geo-Logix provides no warranty as to offsite conditions, including the extent if any to which substances in the Site may be emanating off

site, and if so whether any adjoining sites have been or may be impacted by contamination originating from the Site.

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Geo-Logix has prepared this report with the diligence, care and skill which a reasonable person would expect from a reputable environmental consultancy and in accordance with environmental regulatory authority and industry standards, guidelines and assessment criteria applicable as at the date of this report. Industry standards and environmental criteria change frequently, and may change at any time after the date of this report.

FIGURES



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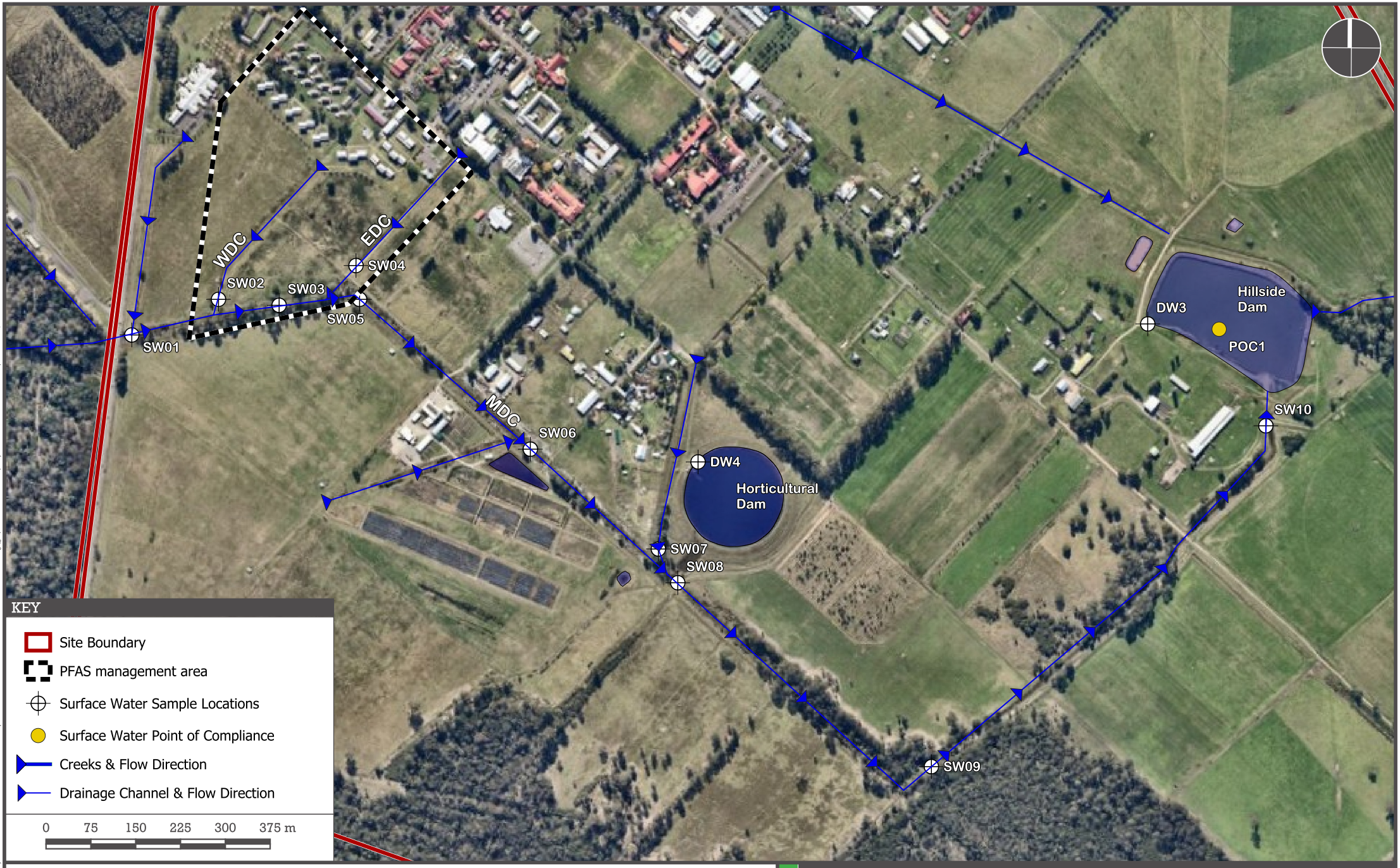
SITE LOCATION MAP

Environmental Management Plan

Western Sydney University - Hawkesbury Campus

Project No. 2301034h

Figure 1



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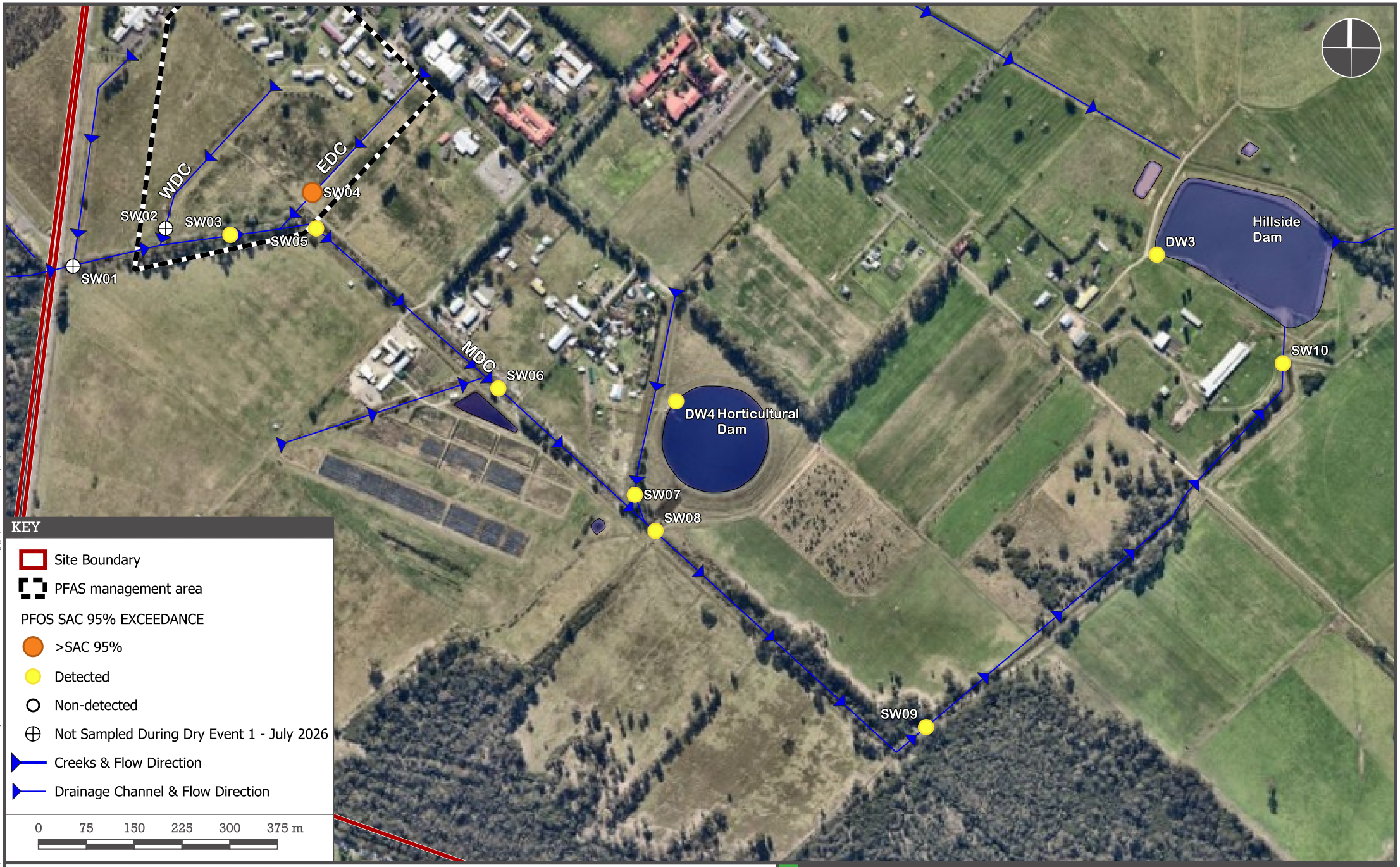
SURFACE WATER MONITORING LOCATIONS

PFAS Surface Water Monitoring - Dry Event 1 July 2026
2 College Street, Richmond NSW 2753

Project No. 2301034j

Figure 2

Filepath: Z:\Share\06_Projects\2023\NSW Projects\2301034 NSW WSU\HC Richmond PFAS\2301034j NSW WSU EMP SW & GW\3 Data\GIS\WSJ Surface Water Monitoring.qgz | Published: 2026-07-30 | Project CRS: GDA2020 / MGA zone 56 | A4 Scale: 1:8,000



Filepath: Z:\Share\06_Projects\2023\NSW Projects\2301034 NSW WSUHC Richmond PFAS\2301034j NSW WSU EMP SW & GW\3 Data\GIS\WSU Surface Water Monitoring.qgz | Published: 2026-07-30 | Project CRS: GDA2020 / MGA zone 56 | A4 Scale: 1:7,500

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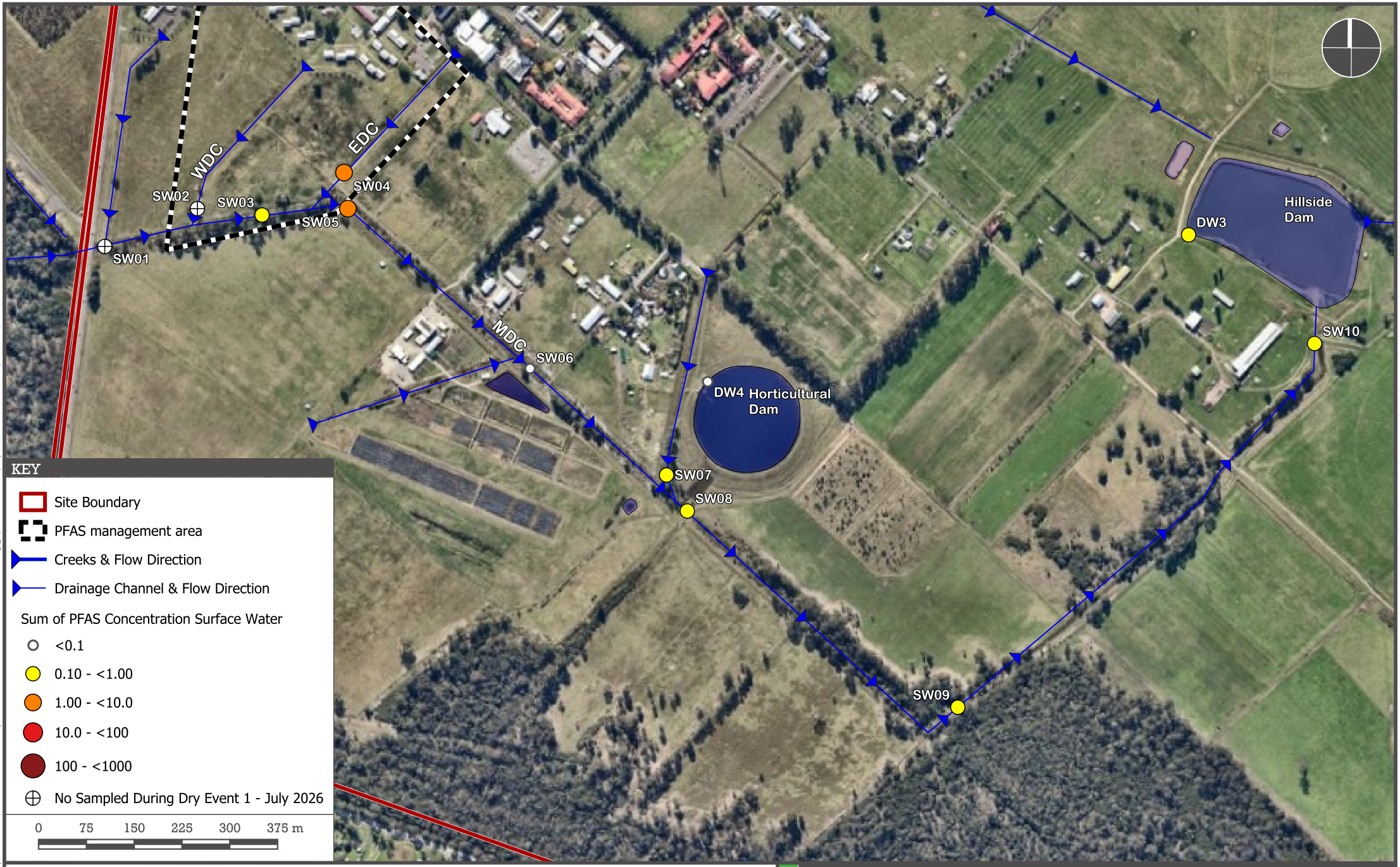
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PFOS SAC EXCEEDANCES - SURFACE WATER

PFAS Surface Water Monitoring - Dry Event 1 July 2026
2 College Street, Richmond NSW 2753

Project No. 2301034j

Figure 3



Filepath: Z:\Share\05_Projects\2023\NSW Projects\2301034 NSW WSUHC Richmond PFAS\2301034j NSW WSU EMP S/W & GW\3 Data\GIS\WSU Surface Water Monitoring.qgz | Published: 2026-07-30 | Project CRS: GDA2020 / MGA zone 56 | A4 Scale: 1:7,500

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SUM OF PFAS CONCENTRATION - SURFACE WATER

PFAS Surface Water Monitoring - Dry Event 1 July 2026
2 College Street, Richmond NSW 2753

Project No. 2301034j

Figure 4

TABLES

Table 1 : Summary of Water Analytical Data - Per- and Polyfluorinated Alkyl Substances

Surface Water Investigation – Dry Event 1 (July 2026)

Project No.: 2301034j

2 College Street
Richmond NSW 2753

	Criteria 1						
	PFAS NEMP	Sample ID	SW03	SW04	SW05	SW06	SW07
	Freshwater	Type	Water	Water	Water	Water	Water
	95%	Date	15/07/2026	15/07/2026	15/07/2026	15/07/2026	15/07/2026
PFPrS	-		< 0.01	0.02	0.03	< 0.01	< 0.01
PFBS	-		0.02	0.08	0.1	< 0.01	< 0.01
PFPeS	-		0.01	0.07	0.1	< 0.01	< 0.01
PFHxS	-		0.08	0.55	0.58	0.02	0.03
PFHpS	-		< 0.01	0.03	0.02	< 0.01	< 0.01
PFOS	0.9		0.2	**1.1**	0.79	0.04	0.18
PFNS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFDS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFBA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
PFPeA	-		0.02	0.12	0.09	< 0.01	< 0.01
PFHxA	-		0.04	0.28	0.21	< 0.01	0.01
PFHpA	-		< 0.01	0.04	0.06	< 0.01	< 0.01
PFOA	220		0.01	0.08	0.07	< 0.01	< 0.01
PFNA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFUnA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFDoA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFTrDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01

Notes:

Criteria 1 = PFAS NEMP 3.1 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DUP1 = duplicate of DW4

TRIP 1 = triplicate of DW4

BLANK = blank sample

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Water Analytical Data - Per- and Polyfluorinated Alkyl Substances

Surface Water Investigation – Dry Event 1 (July 2026)

Project No.: 2301034j

2 College Street
Richmond NSW 2753

	Criteria 1						
	PFAS NEMP	Sample ID	SW03	SW04	SW05	SW06	SW07
	Freshwater	Type	Water	Water	Water	Water	Water
	95%	Date	15/07/2026	15/07/2026	15/07/2026	15/07/2026	15/07/2026
PFTeDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
4:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
6:2 FTS	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
8:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
10:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFOA + PFOS	-		0.21	1.18	0.86	0.04	0.18
PFHxS + PFOS	-		0.28	1.65	1.37	0.06	0.21
Sum of PFAS	-		0.4	2.4	2.1	< 0.1	0.2

Notes:

Criteria 1 = PFAS NEMP 3.1 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

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Surface Water Investigation – Dry Event 1 (July 2026)

Project No.: 2301034j

2 College Street
Richmond NSW 2753

	Criteria 1						
	PFAS NEMP	Sample ID	SW08	SW09	SW10	DW3	DW4
	Freshwater	Type	Water	Water	Water	Water	Water
	95%	Date	15/07/2026	15/07/2026	15/07/2026	15/07/2026	15/07/2026
PFPrS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFBS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFPeS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFHxS	-		0.03	0.05	0.06	0.02	< 0.01
PFHpS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFOS	0.9		0.12	0.11	0.09	0.03	0.02
PFNS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFDS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFBA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
PFPeA	-		< 0.01	< 0.01	0.01	0.03	0.03
PFHxA	-		0.01	0.02	0.03	0.02	0.03
PFHpA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFOA	220		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFNA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFUnA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFDoA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFTrDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01

Notes:

Criteria 1 = PFAS NEMP 3.1 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.

Total concentrations in µg/L

– = assessment criteria not available

DUP1 = duplicate of DW4

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-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Water Analytical Data - Per- and Polyfluorinated Alkyl Substances

Surface Water Investigation – Dry Event 1 (July 2026)

Project No.: 2301034j

2 College Street
Richmond NSW 2753

	Criteria 1						
	PFAS NEMP	Sample ID	SW08	SW09	SW10	DW3	DW4
	Freshwater	Type	Water	Water	Water	Water	Water
	95%	Date	15/07/2026	15/07/2026	15/07/2026	15/07/2026	15/07/2026
PFTeDA	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSE	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
MeFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
EtFOSAA	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
4:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
6:2 FTS	-		< 0.05	< 0.05	< 0.05	< 0.05	< 0.05
8:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
10:2 FTS	-		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01
PFOA + PFOS	-		0.12	0.11	0.09	0.03	0.02
PFHxS + PFOS	-		0.15	0.16	0.15	0.05	0.02
Sum of PFAS	-		0.2	0.2	0.2	0.1	< 0.1

Notes:

Criteria 1 = PFAS NEMP 3.1 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.
Total concentrations in µg/L
– = assessment criteria not available
DUP1 = duplicate of DW4
TRIP 1 = triplicate of DW4
BLANK = blank sample
RPD = relative percent difference of duplicate/triplicate
nc = RPD not calculated, one or both samples below laboratory reporting limit
< # or ND = analyte(s) not detected in excess of laboratory reporting limit
-- = sample not analysed
Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Water Analytical Data - Per- and Polyfluorinated Alkyl Substances

Surface Water Investigation – Dry Event 1 (July 2026)

Project No.: 2301034j

Geo-Logix
2 College Street
Richmond NSW 2753

	Criteria 1						
	PFAS NEMP	Sample ID	DUP1	RPD_DUP1	TRIP 1	RPD_TRIP 1	BLANK
	Freshwater	Type	Water	Water	Water	Water	Water
	95%	Date	15/07/2026	-	15/07/2026	-	15/07/2026
PFPrS	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFBS	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFPeS	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFHxS	-		0.02	*nc*	0.02	*nc*	< 0.01
PFHpS	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFOS	0.9		0.04	*67%*	0.02	*0%*	< 0.01
PFNS	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFDS	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFBA	-		< 0.05	*nc*	< 0.05	*nc*	< 0.05
PFPeA	-		0.03	*0%*	0.02	*40%*	< 0.01
PFHxA	-		0.02	*40%*	0.02	*40%*	< 0.01
PFHpA	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFOA	220		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFNA	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFDA	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFUnA	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFDoA	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFTrDA	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01

Notes:

Criteria 1 = PFAS NEMP 3.1 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.
Total concentrations in µg/L
– = assessment criteria not available
DUP1 = duplicate of DW4
TRIP 1 = triplicate of DW4
BLANK = blank sample
RPD = relative percent difference of duplicate/triplicate
nc = RPD not calculated, one or both samples below laboratory reporting limit
< # or ND = analyte(s) not detected in excess of laboratory reporting limit
-- = sample not analysed
Bold/red indicates exceedance of assessment criteria

Table 1 : Summary of Water Analytical Data - Per- and Polyfluorinated Alkyl Substances

Surface Water Investigation – Dry Event 1 (July 2026)

Project No.: 2301034j

Geo-Logix
2 College Street
Richmond NSW 2753

	Criteria 1						
	PFAS NEMP	Sample ID	DUP1	RPD_DUP1	TRIP 1	RPD_TRIP 1	BLANK
	Freshwater	Type	Water	Water	Water	Water	Water
	95%	Date	15/07/2026	-	15/07/2026	-	15/07/2026
PFTeDA	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFOSA	-		< 0.05	*nc*	< 0.05	*nc*	< 0.05
MeFOSA	-		< 0.05	*nc*	< 0.05	*nc*	< 0.05
EtFOSA	-		< 0.05	*nc*	< 0.05	*nc*	< 0.05
MeFOSE	-		< 0.05	*nc*	< 0.05	*nc*	< 0.05
EtFOSE	-		< 0.05	*nc*	< 0.05	*nc*	< 0.05
MeFOSAA	-		< 0.05	*nc*	< 0.05	*nc*	< 0.05
EtFOSAA	-		< 0.05	*nc*	< 0.05	*nc*	< 0.05
4:2 FTS	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
6:2 FTS	-		< 0.05	*nc*	< 0.05	*nc*	< 0.05
8:2 FTS	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
10:2 FTS	-		< 0.01	*nc*	< 0.01	*nc*	< 0.01
PFOA + PFOS	-		0.04	*67%*	0.02	*0%*	ND
PFHxS + PFOS	-		0.06	*100%*	0.04	*67%*	< 0.01
Sum of PFAS	-		0.1	*nc*	< 0.1	*nc*	< 0.1

Notes:

Criteria 1 = PFAS NEMP 3.1 (HEPA 2025) Aquatic Ecosystems, Freshwater Guidance Values, 95% Species Protection.
Total concentrations in µg/L
– = assessment criteria not available
DUP1 = duplicate of DW4
TRIP 1 = triplicate of DW4
BLANK = blank sample
RPD = relative percent difference of duplicate/triplicate
nc = RPD not calculated, one or both samples below laboratory reporting limit
< # or ND = analyte(s) not detected in excess of laboratory reporting limit
-- = sample not analysed
Bold/red indicates exceedance of assessment criteria

Table 2 : Summary of Water Analytical Data - Total Suspended Solids

Surface Water Investigation – Dry Event 1 (July 2026)

Project No.: 2301034j

2 College Street
Richmond NSW 2753

	Criteria 1					
		Sample ID	SW03	SW04	SW05	SW06	SW07
		Type	Water	Water	Water	Water	Water
		Date	15/07/2026	15/07/2026	15/07/2026	15/07/2026	15/07/2026
Total Suspended Solids (TSS)	-		1,100	21	83	94	1,400

Notes:

Total concentrations in mg/L

– = assessment criteria not available

DUP1 = duplicate of DW4

TRIP 1 = triplicate of DW4

BLANK = blank sample

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 2 : Summary of Water Analytical Data - Total Suspended Solids

Surface Water Investigation – Dry Event 1 (July 2026)

Project No.: 2301034j

2 College Street
Richmond NSW 2753

	Criteria 1						
		Sample ID	SW08	SW09	SW10	DW3	DW4
		Type	Water	Water	Water	Water	Water
		Date	15/07/2026	15/07/2026	15/07/2026	15/07/2026	15/07/2026
Total Suspended Solids (TSS)	-		170	12	490	7.5	< 5

Notes:

Total concentrations in mg/L

– = assessment criteria not available

DUP1 = duplicate of DW4

TRIP 1 = triplicate of DW4

BLANK = blank sample

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

Table 2 : Summary of Water Analytical Data - Total Suspended Solids

Surface Water Investigation – Dry Event 1 (July 2026)

Project No.: 2301034j

2 College Street
Richmond NSW 2753

	Criteria 1						
		Sample ID	DUP1	RPD_DUP1	TRIP 1	RPD_TRIP 1	BLANK
		Type	Water	Water	Water	Water	Water
		Date	15/07/2026	-	15/07/2026	-	15/07/2026
Total Suspended Solids (TSS)	-		--	--	--	--	--

Notes:

Total concentrations in mg/L

– = assessment criteria not available

DUP1 = duplicate of DW4

TRIP 1 = triplicate of DW4

BLANK = blank sample

RPD = relative percent difference of duplicate/triplicate

nc = RPD not calculated, one or both samples below laboratory reporting limit

< # or ND = analyte(s) not detected in excess of laboratory reporting limit

-- = sample not analysed

Bold/red indicates exceedance of assessment criteria

ATTACHMENT A

SURFACE WATER SAMPLING

Western Sydney University – Hawksbury Campus: 2 College Street, Richmond NSW 2753



Plate 1 – SW01: Upgradient PMA – MDC.



Plate 2 – SW02: WDC discharge from PMA.



Plate 3 – SW03: MDC between WDC and EDC.



Plate 4 – SW04: EDC discharge from PMA.

MDC = Main drainage Channel
WDC = PFAS Management Area Western Drainage Channel
EDC = PFAS Management Area Eastern Drainage Channel
POC = Point of Compliance

SURFACE WATER SAMPLING

Western Sydney University – Hawksbury Campus: 2 College Street, Richmond NSW 2753



Plate 5 – SW05: MDC immediately downgradient of WDC + EDC.



Plate 6 – SW06: MDC downgradient of confluence with Native Centre drainage channel.



Plate 7 – Horticulture Dam drainage channel confluence with MDC.



Plate 8 – SW08: MDC.

MDC = Main drainage Channel
WDC = PFAS Management Area Western Drainage Channel
EDC = PFAS Management Area Eastern Drainage Channel
POC = Point of Compliance

SURFACE WATER SAMPLING

Western Sydney University – Hawksbury Campus: 2 College Street, Richmond NSW 2753



Plate 9 – SW09: MDC.



Plate 10 – SW10: MDC



Plate 11 – DW03: Hillside Dam.



Plate 12 – DW04: Horticultural Dam.

MDC = Main drainage Channel
WDC = PFAS Management Area Western Drainage Channel
EDC = PFAS Management Area Eastern Drainage Channel
POC = Point of Compliance

ATTACHMENT B

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: DW4

PROJECT INFORMATION

PROJECT NUMBER: 2301034j	**INITIALS:** ZG+PDP
DATE: 15.7.26	**WEATHER:** Sunny 16°

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	**TOTAL DEPTH (mBTOC):**	**STICK-UP (m):**
DEPTH TO PSH (mBTOC):	**THICKNESS OF PSH (m):**	**TIME:** 10:30

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
10:30			7.98	645	84.1	8.08	13.9
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) brown colour, no oily sheen, no odour, particulate matter, vegetation.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: DW4	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 10:27	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 2	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS-TSD	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: DW3

PROJECT INFORMATION

PROJECT NUMBER: 2301034j	**INITIALS:** ZG + PDP
DATE: 15.7.26	**WEATHER:** Sunny 16°

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	**TOTAL DEPTH (mBTOC):**	**STICK-UP (m):**
DEPTH TO PSH (mBTOC):	**THICKNESS OF PSH (m):**	**TIME:** 9:50.

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
9:55			7.10	539	88.5	6.27	13.7
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) clear water, organic odor, algae, no oily sheen, no plastics etc.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: DW3	**SAMPLE ID:** DUP1	**SAMPLE ID:** TRIP1.
SAMPLE TIME: 9:50 AM.	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: PFAS - GREEN	**NO. CONTAINERS** 2	**NO. CONTAINERS** 2
ANALYSIS: PFAS - TSD.	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: SW10

PROJECT INFORMATION

PROJECT NUMBER: 2301034J	**INITIALS:** ZG + PDP
DATE: 15.7.26	**WEATHER:** Sunny 16°

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	**TOTAL DEPTH (mBTOC):**	**STICK-UP (m):**
DEPTH TO PSH (mBTOC):	**THICKNESS OF PSH (m):**	**TIME:** 10:10

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
10:10			6.59	219.7	88.6	6.80	10.1
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) shallow water, slight brown colour, particulate matter, no odour & sheen,

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: SW10	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 10:05	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 2.	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS-TSS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: SW09

PROJECT INFORMATION

PROJECT NUMBER: 2301034	**INITIALS:** ZG + PDP
DATE: 15.7.26	**WEATHER:**

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	**TOTAL DEPTH (mBTOC):**	**STICK-UP (m):**
DEPTH TO PSH (mBTOC):	**THICKNESS OF PSH (m):**	**TIME:** 11:05

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
11:05			6.72	2.4	99.7	11.95	10.1
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) clear, no odour, no oily sheen,

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: SW09	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 11:02	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 2	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS - TSS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: SW08

PROJECT INFORMATION

PROJECT NUMBER: 2301030j	**INITIALS:** ZG + PDP
DATE: 15.7.26	**WEATHER:**

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	**TOTAL DEPTH (mBTOC):**	**STICK-UP (m):**
DEPTH TO PSH (mBTOC):	**THICKNESS OF PSH (m):**	**TIME:** 10:50

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
10:50			6.97	223.4	97.7	4.54	10.2
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR)

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: SW08	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 10:45	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 2	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS TSS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: SW07

PROJECT INFORMATION

PROJECT NUMBER: 2301034j	**INITIALS:** ZG + PDP
DATE: 15.7.26	**WEATHER:**

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	**TOTAL DEPTH (mBTOC):**	**STICK-UP (m):**
DEPTH TO PSH (mBTOC):	**THICKNESS OF PSH (m):**	**TIME:** 11:40

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
11:40			6.10	223.6	109.6	3.45	10.3
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) clear / slight brown colour, particulate matter, oily sheen, no odour

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: 11:BD SW07	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: & 11:37	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 2	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS-TSS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: SW06

PROJECT INFORMATION

PROJECT NUMBER: 2301034	**INITIALS:** ZG + PDP
DATE: 15.7.26	**WEATHER:** 18° sunny

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	**TOTAL DEPTH (mBTOC):**	**STICK-UP (m):**
DEPTH TO PSH (mBTOC):	**THICKNESS OF PSH (m):**	**TIME:** 11:50

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
11:50			6.60	223.0	92.7	5.70	11.8
TOTAL PURGE VOLUME (L):		APPROX. SAMPLE PURGE RATE (LPM):					

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) slight brown hue, lots of veg on top of water, no odour, particulate matter, no oily sheen.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: SW06	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 11:46	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 2	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS TSS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: SW05

PROJECT INFORMATION

PROJECT NUMBER: 2301034J	**INITIALS:** ZG + PDP
DATE: 15.7.26	**WEATHER:**

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	**TOTAL DEPTH (mBTOC):**	**STICK-UP (m):**
DEPTH TO PSH (mBTOC):	**THICKNESS OF PSH (m):**	**TIME:** 12:05

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
12:05			6.86	695	21.6	3.30	10.6
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) oily sheen, heavy organic odour, particulate matter, clear water, lots of decomposing vegetation.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: SW05	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 12:01	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 2	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS - TSS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: SW04

PROJECT INFORMATION

PROJECT NUMBER: 2301034j	**INITIALS:** ZG + PDP
DATE: 15.7.26	**WEATHER:**

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	**TOTAL DEPTH (mBTOC):**	**STICK-UP (m):**
DEPTH TO PSH (mBTOC):	**THICKNESS OF PSH (m):**	**TIME:** 12:16

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
12:16			7.13	906	87.3	7.89	12.8
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) clear, no odour, no oily sheen.

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: SW04	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 12:13	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 2	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS - TSS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

Geo-Logix

GROUNDWATER SAMPLE LOG

WELL ID: SW03

PROJECT INFORMATION

PROJECT NUMBER: 2301034j	**INITIALS:** ZG + PDP
DATE: 15 7 26	**WEATHER:**

SAMPLING METHOD: LOW FLOW: ☐ HYDRASLEEVE: ☐ BAILER: ☐

WELL GAUGING DETAILS

STANDING WATER LEVEL (mBTOC):	**TOTAL DEPTH (mBTOC):**	**STICK-UP (m):**
DEPTH TO PSH (mBTOC):	**THICKNESS OF PSH (m):**	**TIME:** 12:35

FIELD PARAMETERS

TIME	DTW (mBTOC)	TOTAL DISCHARGE (L)	pH +/- 0.1	CONDUCTIVITY (µS/CM) +/- 3%	REDOX (MV) +/- 10MV	DO (MG/L) +/-10%	TEMP (°C)
12:35			6.61	265.6	89.6	1.91	12.9
TOTAL PURGE VOLUME (L):			APPROX. SAMPLE PURGE RATE (LPM):				

OBSERVATIONS DURING SAMPLING

NOTES: (WELL CONDITION, COLOUR, CLARITY, ODOUR) dark brown in colour, very strong organic odour, no oily sheen, vegetation, particulate matter

RECHARGE BEHAVIOUR: FAST RECHARGING ☐ SLOW RECHARGING (<80% RECHARGE AFTER 2 HRS) ☐

WELL SAMPLING

DTW (mbTOC): (AT SAMPLING)

ORIGINAL	DUPLICATE	TRIPLICATE
SAMPLE ID: SW03	**SAMPLE ID:**	**SAMPLE ID:**
SAMPLE TIME: 12:31	**SAMPLE TIME:**	**SAMPLE TIME:**
NO. CONTAINERS: 2	**NO. CONTAINERS**	**NO. CONTAINERS**
ANALYSIS: PFAS, TSS	**ANALYSIS:**	**ANALYSIS:**

ORIGINAL FIELD RECORD

ATTACHMENT C

Certificate of Service and Calibration

Water Quality Meter

YSI Professional Plus

Company Name	WAM Scientific
Office Address	26 Bungarra Crescent, Chipping Norton NSW 2170
Phone Number	+61 405 241 484
Contact Name	William Pak
Instrument	YSI Pro Quatro Water Quality Meter w/ 1m Quatro Cable
Serial Number	20B122031
Client Name	Zahra Ginwala, Paula Del Prete (Geo-Logix)
Project Number	PO7245

Instrument Check

Item	Test	Test Passed	Comments
2 x Alkaline C-size Batteries	Klein Tools MM300 Multimeter	✓	Both batteries reading above 2.9V
Battery Saver Function	Operation	✓	Automatically turns off after 60 minutes if idle
Unit Display	Operation	✓	Screen visible, no damage
Keypad	Operation	✓	Responsive, no damage
Connection Port and Cable	Condition/Check	✓	Clean, no damage
Monitor Housing	Condition/Check	✓	No damage
Firmware	Version	✓	4.0.0
pH Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
pH millivolts for pH 7.00	Calibration	✓	pH 7.00 calibration range between 0 mV ± 50 mV
pH millivolts for pH 4.00	Calibration	✓	pH 4 mV range +165 to +180 from 7 buffer mV value
pH slope	Calibration	✓	Range between 55 to 60 mV/pH (ideal value 59 mV)
Response time < 90 seconds	Calibration	✓	Responds to correct value within 90 seconds
ORP Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
ORP Reading	Calibration	✓	Within ± 80 mV of reference Zobell Reading
Response time < 90 seconds	Calibration	✓	Responds to correct value within 90 seconds
Conductivity/Temp Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
Conductivity Cell	Calibration	✓	Conductivity cell constant 5.0 ± 1.0 in GLP file
Clean Sensor Readings	Calibration	✓	Clean sensor reads less than 3 uS/cm in dry air
Dissolved Oxygen Probe	Condition/Calibration	✓	Calibrated and conforms to manufacturer's specs
DO Cap	Condition/Calibration	✓	1.25 mil PE membrane (yellow membrane)
DO Sensor in Use	Condition	✓	Polarographic DO sensor
DO Sensor Value	Calibration	✓	(min 4.31 uA - max 8.00 uA) Avg 6.15 uA

Instrument Readings

Parameter	Standard Used	Reference No.	Calibration Value	Observed	Actual	Units
Temperature	Centre 370 Thermometer	Room Temp.	15.9	15.1	15.9	°C
pH	pH 4.00	417183	4.01	3.98	4.01	pH
pH	pH 7.00	419528	7.00	6.94	7.00	pH
ORP	Zobell A & B	420448/418958	249.0	252.7	249.0	mV
Conductivity	2760 µs/cm at 25°C	399819	2760	2962	2760	µs/cm
Zero Dissolved O_2	$NaSO_3$ in Distilled H_2O	426184	0.0	0.0	0.0	%
100% Dissolved O_2	100% Air Saturated H_2O	Fresh Air	100.0	99.5	100.0	%

WAM Scientific certifies that the above instrument was successfully tested according to manufacturer's standards and all necessary checks were conducted to ensure the instrument was fully operational prior to dispatch. The calibration data supplied was obtained in accordance with manufacturer's specifications using solutions of known values.

Calibrated By	William Pak
Calibration Date	13/07/2026
Calibration Due	13/01/2027

WAM SCIENTIFIC
+61 418 389 790
rentals@wamscientific.com.au
26 Bungarra Crescent, CHIPPING NORTON NSW 2170
www.wamscientific.com.au

ATTACHMENT D

Geo-Logix P/L
Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

NATA Accredited
Accreditation Number 1261
Site Number 18217

Accredited for compliance with ISO/IEC 17025 – Testing
NATA is a signatory to the ILAC Mutual Recognition Arrangement for the mutual recognition of the equivalence of testing, medical testing, calibration, inspection, proficiency testing scheme providers and reference materials producers reports and certificates.

Attention: **Zahra Ginwala**

Report **1384514-W**
Project name WSU Surface Water Sampling
Project ID 2301034J
Received Date Jul 16, 2026

Client Sample ID			**SW03**	**SW04**	**SW05**	**SW06**
Sample Matrix			**Water**	**Water**	**Water**	**Water**
Eurofins Sample No.			**S26-JI0045112**	**S26-JI0045113**	**S26-JI0045114**	**S26-JI0045115**
Date Sampled			**Jul 15, 2026**	**Jul 15, 2026**	**Jul 15, 2026**	**Jul 15, 2026**
Test/Reference	LOR	Unit				
Total Suspended Solids Dried at 103°C to 105°C	5	mg/L	1100	21	83	94
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	0.02	0.12	0.09	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.04	0.28	0.21	< 0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	[N09]0.04	[N09]0.06	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	[N09]0.01	[N09]0.08	[N09]0.07	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	84	76	78	64
13C5-PFPeA (surr.)	1	%	102	79	95	74
13C5-PFHxA (surr.)	1	%	113	87	110	92
13C4-PFHpA (surr.)	1	%	121	93	98	79
13C8-PFOA (surr.)	1	%	102	77	90	70
13C5-PFNA (surr.)	1	%	99	71	81	74
13C6-PFDA (surr.)	1	%	92	81	82	65
13C2-PFUnDA (surr.)	1	%	80	76	74	54
13C2-PFDoDA (surr.)	1	%	79	60	61	55
13C2-PFTeDA (surr.)	1	%	91	102	74	90
Perfluoroalkyl sulfonamido substances						
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	95	77	84	66

Client Sample ID			SW03	SW04	SW05	SW06
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S26-JI0045112	S26-JI0045113	S26-JI0045114	S26-JI0045115
Date Sampled			Jul 15, 2026	Jul 15, 2026	Jul 15, 2026	Jul 15, 2026
Test/Reference	LOR	Unit				
Perfluoroalkyl sulfonamido substances						
D3-N-MeFOSA (surr.)	1	%	93	105	88	74
D5-N-EtFOSA (surr.)	1	%	98	97	87	74
D7-N-MeFOSE (surr.)	1	%	69	63	69	53
D9-N-EtFOSE (surr.)	1	%	71	67	68	60
D5-N-EtFOSAA (surr.)	1	%	73	59	62	57
D3-N-MeFOSAA (surr.)	1	%	81	72	79	63
Perfluoroalkyl sulfonic acids (PFSAs)						
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	0.02	0.08	0.10	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	< 0.01	0.02	0.03	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	[N09]0.01	[N09]0.07	[N09]0.10	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]0.08	[N09]0.55	[N09]0.58	[N09]0.02
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01	[N09]0.03	[N09]0.02	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	[N09]0.20	[N09]1.1	[N09]0.79	[N09]0.04
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	118	95	107	95
18O2-PFHxS (surr.)	1	%	115	93	103	83
13C8-PFOS (surr.)	1	%	96	86	88	71
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)						
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	[J+]152	107	115	88
13C2-6:2 FTSA (surr.)	1	%	131	85	81	72
13C2-8:2 FTSA (surr.)	1	%	123	92	97	65
13C2-10:2 FTSA (surr.)	1	%	65	71	52	59
PFASs Summations						
Sum (PFHxS + PFOS)*	0.01	ug/L	0.28	1.65	1.37	0.06
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	0.37	2.25	1.9	0.06
Sum of PFASs (n=30)*	0.1	ug/L	0.4	2.4	2.1	< 0.1

Client Sample ID			SW07	SW08	SW09	SW10
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S26-JI0045116	S26-JI0045117	S26-JI0045118	S26-JI0045119
Date Sampled			Jul 15, 2026	Jul 15, 2026	Jul 15, 2026	Jul 15, 2026
Test/Reference	LOR	Unit				
Total Suspended Solids Dried at 103°C to 105°C	5	mg/L	1400	170	12	490
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.01	0.01	0.02	0.03
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01

Client Sample ID			SW07	SW08	SW09	SW10
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S26-JI0045116	S26-JI0045117	S26-JI0045118	S26-JI0045119
Date Sampled			Jul 15, 2026	Jul 15, 2026	Jul 15, 2026	Jul 15, 2026
Test/Reference	LOR	Unit				
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	76	77	76	100
13C5-PFPeA (surr.)	1	%	77	87	83	96
13C5-PFHxA (surr.)	1	%	88	89	93	113
13C4-PFHpA (surr.)	1	%	88	91	82	98
13C8-PFOA (surr.)	1	%	74	78	88	97
13C5-PFNA (surr.)	1	%	72	76	82	104
13C6-PFDA (surr.)	1	%	68	85	81	107
13C2-PFUnDA (surr.)	1	%	67	67	81	82
13C2-PFDoDA (surr.)	1	%	59	59	67	80
13C2-PFTeDA (surr.)	1	%	116	102	143	J+152
Perfluoroalkyl sulfonamido substances						
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	69	77	83	102
D3-N-MeFOSA (surr.)	1	%	89	88	97	111
D5-N-EtFOSA (surr.)	1	%	88	89	102	107
D7-N-MeFOSE (surr.)	1	%	61	59	77	81
D9-N-EtFOSE (surr.)	1	%	69	63	82	86
D5-N-EtFOSAA (surr.)	1	%	60	55	63	85
D3-N-MeFOSAA (surr.)	1	%	68	67	75	77
Perfluoroalkyl sulfonic acids (PFSAs)						
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]0.03	[N09]0.03	[N09]0.05	[N09]0.06
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	[N09]0.18	[N09]0.12	[N09]0.11	[N09]0.09
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	92	104	86	121
18O2-PFHxS (surr.)	1	%	82	84	99	108
13C8-PFOS (surr.)	1	%	75	79	86	107

Client Sample ID			SW07	SW08	SW09	SW10
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S26-JI0045116	S26-JI0045117	S26-JI0045118	S26-JI0045119
Date Sampled			Jul 15, 2026	Jul 15, 2026	Jul 15, 2026	Jul 15, 2026
Test/Reference	LOR	Unit				
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)						
1H,1H,2H,2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H,1H,2H,2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
1H,1H,2H,2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H,1H,2H,2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	91	107	132	J+159
13C2-6:2 FTSA (surr.)	1	%	88	79	101	114
13C2-8:2 FTSA (surr.)	1	%	72	96	82	109
13C2-10:2 FTSA (surr.)	1	%	57	61	73	80
PFASs Summations						
Sum (PFHxS + PFOS)*	0.01	ug/L	0.21	0.15	0.16	0.15
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	0.22	0.16	0.18	0.19
Sum of PFASs (n=30)*	0.1	ug/L	0.2	0.2	0.2	0.2

Client Sample ID			DW3	DW4	DUP1	BLANK
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S26-JI0045120	S26-JI0045121	S26-JI0045122	S26-JI0045123
Date Sampled			Jul 15, 2026	Jul 15, 2026	Jul 15, 2026	Jul 15, 2026
Test/Reference	LOR	Unit				
Total Suspended Solids Dried at 103°C to 105°C	5	mg/L	7.5	< 5	-	-
Perfluoroalkyl carboxylic acids (PFCAs)						
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	0.03	0.03	0.03	< 0.01
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.02	0.03	0.02	< 0.01
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C4-PFBA (surr.)	1	%	110	108	87	65
13C5-PFPeA (surr.)	1	%	98	117	80	69
13C5-PFHxA (surr.)	1	%	138	125	82	67
13C4-PFHpA (surr.)	1	%	116	141	87	64
13C8-PFOA (surr.)	1	%	109	134	89	65
13C5-PFNA (surr.)	1	%	118	103	83	62
13C6-PFDA (surr.)	1	%	103	113	79	68
13C2-PFUnDA (surr.)	1	%	92	98	71	57
13C2-PFDoDA (surr.)	1	%	86	95	62	57
13C2-PFTeDA (surr.)	1	%	J+168	J+200	140	118

Client Sample ID			DW3	DW4	DUP1	BLANK
Sample Matrix			Water	Water	Water	Water
Eurofins Sample No.			S26-JI0045120	S26-JI0045121	S26-JI0045122	S26-JI0045123
Date Sampled			Jul 15, 2026	Jul 15, 2026	Jul 15, 2026	Jul 15, 2026
Test/Reference	LOR	Unit				
Perfluoroalkyl sulfonamido substances						
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
13C8-FOSA (surr.)	1	%	107	121	92	69
D3-N-MeFOSA (surr.)	1	%	122	[J+]156	105	87
D5-N-EtFOSA (surr.)	1	%	129	[J+]153	105	95
D7-N-MeFOSE (surr.)	1	%	88	101	71	61
D9-N-EtFOSE (surr.)	1	%	89	107	75	63
D5-N-EtFOSAA (surr.)	1	%	83	80	59	55
D3-N-MeFOSAA (surr.)	1	%	92	101	78	57
Perfluoroalkyl sulfonic acids (PFSAs)						
Perfluorobutanesulfonic acid (PFBS)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorononanesulfonic acid (PFNS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropropanesulfonic acid (PFPrS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluoropentanesulfonic acid (PFPeS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorohexanesulfonic acid (PFHxS)[N11]	0.01	ug/L	[N09]0.02	< 0.01	[N09]0.02	< 0.01
Perfluoroheptanesulfonic acid (PFHpS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
Perfluorooctanesulfonic acid (PFOS)[N11]	0.01	ug/L	[N09]0.03	[N09]0.02	[N09]0.04	< 0.01
Perfluorodecanesulfonic acid (PFDS)[N15]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C3-PFBS (surr.)	1	%	141	122	93	64
18O2-PFHxS (surr.)	1	%	125	134	89	75
13C8-PFOS (surr.)	1	%	106	122	82	72
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)						
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)[N11]	0.05	ug/L	< 0.05	< 0.05	< 0.05	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)[N11]	0.01	ug/L	< 0.01	< 0.01	< 0.01	< 0.01
13C2-4:2 FTSA (surr.)	1	%	[J+]186	[J+]160	110	90
13C2-6:2 FTSA (surr.)	1	%	97	136	108	72
13C2-8:2 FTSA (surr.)	1	%	102	116	92	65
13C2-10:2 FTSA (surr.)	1	%	85	102	51	58
PFASs Summations						
Sum (PFHxS + PFOS)*	0.01	ug/L	0.05	0.02	0.06	< 0.01
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	0.1	0.08	0.11	< 0.05
Sum of PFASs (n=30)*	0.1	ug/L	0.1	< 0.1	0.1	< 0.1

Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description	Testing Site	Extracted	Holding Time
Total Suspended Solids Dried at 103°C to 105°C	Sydney	Jul 16, 2026	7 Days
- Method: LTM-INO-4070 Analysis of Suspended Solids in Water by Gravimetry			
Per- and Polyfluoroalkyl Substances (PFASs)			
Perfluoroalkyl carboxylic acids (PFCAs)	Sydney	Jul 17, 2026	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonamido substances	Sydney	Jul 17, 2026	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonic acids (PFSAs)	Sydney	Jul 17, 2026	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)	Sydney	Jul 17, 2026	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			

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Eurofins Environment Testing Australia Pty Ltd
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Dandenong South	Grovedale	Girraween	Mayfield West	Mitchell	Murarrie	North Hobart
VIC 3175	VIC 3216	NSW 2145	NSW 2304	ACT 2911	QLD 4172	TAS 7000
+61 3 8564 5000	+61 3 8564 5000	+61 2 9900 8400	+61 2 4968 8448	+61 2 6113 8091	+61 7 3902 4600	+61 3 8564 5000
NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261
Site# 1254	Site# 25403	Site# 18217	Site# 25079	Site# 25466	Site# 20794 & 2780	Site# 26223

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Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: WSU Surface Water Sampling
Project ID: 2301034J

Order No.:
Report #: 1384514
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Jul 16, 2026 1:42 PM
Due: Jul 23, 2026
Priority: 5 Day
Contact Name: Zahra Ginwala

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail						CANCELLED*	Total Suspended Solids Dried at 103°C to 105°C	Per- and Polyfluoroalkyl Substances (PFASs)
Sydney Laboratory - NATA # 1261 Site # 18217						X	X	X
External Laboratory								
No	**Sample ID**	**Sample Date**	**Sampling Time**	**Matrix**	**LAB ID**			
1	SW01	Jul 15, 2026		Water	S26-Jl0045110	X		
2	SW02	Jul 15, 2026		Water	S26-Jl0045111	X		
3	SW03	Jul 15, 2026		Water	S26-Jl0045112		X	X
4	SW04	Jul 15, 2026		Water	S26-Jl0045113		X	X
5	SW05	Jul 15, 2026		Water	S26-Jl0045114		X	X
6	SW06	Jul 15, 2026		Water	S26-Jl0045115		X	X
7	SW07	Jul 15, 2026		Water	S26-Jl0045116		X	X
8	SW08	Jul 15, 2026		Water	S26-Jl0045117		X	X
9	SW09	Jul 15, 2026		Water	S26-Jl0045118		X	X
10	SW10	Jul 15, 2026		Water	S26-Jl0045119		X	X
11	DW3	Jul 15, 2026		Water	S26-Jl0045120		X	X
12	DW4	Jul 15, 2026		Water	S26-Jl0045121		X	X
13	DUP1	Jul 15, 2026		Water	S26-Jl0045122			X
14	BLANK	Jul 15, 2026		Water	S26-Jl0045123			X

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NATA# 2377
Site# 2370 & 2554

Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: WSU Surface Water Sampling
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Order No.:
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Phone: 02 9979 1722
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Received: Jul 16, 2026 1:42 PM
Due: Jul 23, 2026
Priority: 5 Day
Contact Name: Zahra Ginwala

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail	CANCELLED*	Total Suspended Solids Dried at 103°C to 105°C	Per- and Polyfluoroalkyl Substances (PFASs)
Sydney Laboratory - NATA # 1261 Site # 18217	X	X	X
Test Counts	2	10	12

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Internal Quality Control Review and Glossary

General

1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follow guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013. They are included in this QC report where applicable. Additional QC data may be available on request.
2. Unless otherwise stated, all soil/sediment/solid results are reported on a dry weight basis.
3. Unless otherwise stated, all biota/food results are reported on a wet weight basis on the edible portion.
4. For CEC results where the sample's origin is unknown or environmentally contaminated, the results should be used advisedly.
5. Actual LORs are matrix dependent. Quoted LORs may be raised where sample extracts are diluted due to interferences.
6. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds where annotated.
7. SVOC analysis on waters is performed on homogenised, unfiltered samples unless noted otherwise.
8. Samples were analysed on an 'as received' basis.
9. Information identified in this report with **blue** colour indicates data provided by customers that may have an impact on the results.
10. This report replaces any interim results previously issued.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours before sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the sampling date; therefore, compliance with these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether, the holding time is seven days; however, for all other VOCs, such as BTEX or C6-10 TRH, the holding time is 14 days.

Units

mg/kg: milligrams per kilogram	**mg/L:** milligrams per litre	**ppm:** parts per million
µg/L: micrograms per litre	**ppb:** parts per billion	**%:** Percentage
org/100 mL: Organisms per 100 millilitres	**NTU:** Nephelometric Turbidity Units	**MPN/100 mL:** Most Probable Number of organisms per 100 millilitres
CFU: Colony Forming Unit	**Colour:** Pt-Co Units (CU)	

Terms

APHA	American Public Health Association
CEC	Cation Exchange Capacity
COC	Chain of Custody
CP	Client Parent - QC was performed on samples pertaining to this report
CRM	Certified Reference Material (ISO17034) - reported as percent recovery.
Dry	Where moisture has been determined on a solid sample, the result is expressed on a dry weight basis.
Duplicate	A second piece of analysis from the same sample and reported in the same units as the result to show comparison.
LOR	Limit of Reporting.
LCS	Laboratory Control Sample - reported as percent recovery.
Method Blank	In the case of solid samples, these are performed on laboratory-certified clean sands and in the case of water samples, these are performed on de-ionised water.
NCP	Non-Client Parent - QC performed on samples not pertaining to this report, QC represents the sequence or batch that client samples were analysed within.
RPD	Relative Percent Difference between two Duplicate pieces of analysis.
SPIKE	Addition of the analyte to the sample and reported as percentage recovery.
SRA	Sample Receipt Advice
Surr - Surrogate	The addition of a similar compound to the analyte target is reported as percentage recovery. See below for acceptance criteria.
TBTO	Tributyltin oxide (*bis*-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment; however, free tributyltin was measured, and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits.
TCLP	Toxicity Characteristic Leaching Procedure
TEQ	Toxic Equivalency Quotient or Total Equivalence
QSM	US Department of Defense Quality Systems Manual Version 6.0
US EPA	United States Environmental Protection Agency
WA DWER	Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should only be used as a guide and may be different when site-specific Sampling Analysis and Quality Plan (SAQP) have been implemented.

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is ≤30%; however, the following acceptance guidelines are equally applicable:

Results <10 times the LOR:	No Limit
Results between 10-20 times the LOR:	RPD must lie between 0-50%
Results >20 times the LOR:	RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range, not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 – 150%, VOC recoveries 50 – 150%

PFAS field samples containing surrogate recoveries above the QC limit designated in QSM 6.0, where no positive PFAS results have been reported or reviewed, and no data was affected.

QC Data General Comments

1. Where a result is reported as less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown are not data from your samples.
3. pH and Free Chlorine analysed in the laboratory - Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time.Analysis will begin as soon as possible after sample receipt.
4. Recovery Data (Spikes & Surrogates) - where chromatographic interference does not allow the determination of recovery, the term "INT" appears against that analyte.
5. For Matrix Spikes and LCS results, a dash "-" in the report means that the specific analyte was not added to the QC sample.
6. Duplicate RPDs are calculated from raw analytical data; thus, it is possible to have two sets of data.

Quality Control Results

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Method Blank							
Total Suspended Solids Dried at 103°C to 105°C	mg/L	< 5			5	Pass	
Method Blank							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	ug/L	< 0.05			0.05	Pass	
Perfluoropentanoic acid (PFPeA)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanoic acid (PFHxA)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanoic acid (PFHpA)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanoic acid (PFOA)	ug/L	< 0.01			0.01	Pass	
Perfluorononanoic acid (PFNA)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanoic acid (PFDA)	ug/L	< 0.01			0.01	Pass	
Perfluoroundecanoic acid (PFUnDA)	ug/L	< 0.01			0.01	Pass	
Perfluorododecanoic acid (PFDoDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotridecanoic acid (PFTrDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotetradecanoic acid (PFTeDA)	ug/L	< 0.01			0.01	Pass	
Method Blank							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	ug/L	< 0.05			0.05	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	ug/L	< 0.05			0.05	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	ug/L	< 0.05			0.05	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	ug/L	< 0.05			0.05	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	ug/L	< 0.05			0.05	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	ug/L	< 0.05			0.05	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	ug/L	< 0.05			0.05	Pass	
Method Blank							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	ug/L	< 0.01			0.01	Pass	
Perfluorononanesulfonic acid (PFNS)	ug/L	< 0.01			0.01	Pass	
Perfluoropropanesulfonic acid (PFPrS)	ug/L	< 0.01			0.01	Pass	
Perfluoropentanesulfonic acid (PFPeS)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanesulfonic acid (PFHxS)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanesulfonic acid (PFOS)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanesulfonic acid (PFDS)	ug/L	< 0.01			0.01	Pass	
Method Blank							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	ug/L	< 0.05			0.05	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	ug/L	< 0.01			0.01	Pass	
LCS - % Recovery							
Total Suspended Solids Dried at 103°C to 105°C	%	96			70-130	Pass	
LCS - % Recovery							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	%	84			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	%	110			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	%	110			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	%	94			50-150	Pass	
Perfluorooctanoic acid (PFOA)	%	107			50-150	Pass	
Perfluorononanoic acid (PFNA)	%	124			50-150	Pass	
Perfluorodecanoic acid (PFDA)	%	132			50-150	Pass	

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Perfluoroundecanoic acid (PFUnDA)	%	116			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	%	105			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	%	118			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	%	128			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	%	119			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	%	102			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	%	117			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	%	105			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	%	118			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	%	111			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	%	108			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	%	121			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	%	98			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	%	103			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	%	103			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	%	98			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	%	108			50-150	Pass	
Perfluorooctanesulfonic acid (PFOS)	%	116			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	%	88			50-150	Pass	
LCS - % Recovery							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	%	118			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	%	115			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	%	113			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	%	98			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Spike - % Recovery									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1					
Perfluorobutanoic acid (PFBA)	S26-Jl0045116	CP	%	80			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	S26-Jl0045116	CP	%	111			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	S26-Jl0045116	CP	%	126			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	S26-Jl0045116	CP	%	95			50-150	Pass	
Perfluorooctanoic acid (PFOA)	S26-Jl0045116	CP	%	101			50-150	Pass	
Perfluorononanoic acid (PFNA)	S26-Jl0045116	CP	%	104			50-150	Pass	
Perfluorodecanoic acid (PFDA)	S26-Jl0045116	CP	%	113			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	S26-Jl0045116	CP	%	119			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	S26-Jl0045116	CP	%	114			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	S26-Jl0045116	CP	%	115			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S26-Jl0045116	CP	%	114			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonamido substances				Result 1					
Perfluorooctane sulfonamide (FOSA)	S26-Jl0045116	CP	%	101			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S26-Jl0045116	CP	%	97			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S26-Jl0045116	CP	%	104			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S26-Jl0045116	CP	%	104			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S26-Jl0045116	CP	%	103			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S26-Jl0045116	CP	%	91			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S26-Jl0045116	CP	%	114			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1					
Perfluorobutanesulfonic acid (PFBS)	S26-Jl0045116	CP	%	145			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	S26-Jl0045116	CP	%	92			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S26-Jl0045116	CP	%	91			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S26-Jl0045116	CP	%	105			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S26-Jl0045116	CP	%	121			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S26-Jl0045116	CP	%	88			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	S26-Jl0045116	CP	%	94			50-150	Pass	
Spike - % Recovery									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1					
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S26-Jl0045116	CP	%	90			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S26-Jl0045116	CP	%	115			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S26-Jl0045116	CP	%	97			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S26-Jl0045116	CP	%	103			50-150	Pass	
Test	**Lab Sample ID**	**QA Source**	**Units**	**Result 1**			**Acceptance Limits**	**Pass Limits**	**Qualifying Code**
Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluorobutanoic acid (PFBA)	S26-Jl0045115	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Perfluoropentanoic acid (PFPeA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanoic acid (PFHxA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroheptanoic acid (PFHpA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanoic acid (PFOA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanoic acid (PFNA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanoic acid (PFDA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroundecanoic acid (PFUnDA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorododecanoic acid (PFDoDA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotridecanoic acid (PFTrDA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
Perfluorooctane sulfonamide (FOSA)	S26-Jl0045115	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S26-Jl0045115	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S26-Jl0045115	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S26-Jl0045115	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S26-Jl0045115	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S26-Jl0045115	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S26-Jl0045115	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1	Result 2	RPD			
Perfluorobutanesulfonic acid (PFBS)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanesulfonic acid (PFNS)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S26-Jl0045115	CP	ug/L	0.02	0.02	12	30%	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanesulfonic acid (PFOS)	S26-Jl0045115	CP	ug/L	0.04	0.04	1.0	30%	Pass	
Perfluorodecanesulfonic acid (PFDS)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1	Result 2	RPD			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S26-Jl0045115	CP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S26-Jl0045115	CP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
				Result 1	Result 2	RPD			
Total Suspended Solids Dried at 103°C to 105°C	S26-Jl0043819	NCP	mg/L	27	16	49	30%	Fail	Q15
Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluorobutanoic acid (PFBA)	S26-Jl0045748	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Perfluoropentanoic acid (PFPeA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanoic acid (PFHxA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroheptanoic acid (PFHpA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanoic acid (PFOA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanoic acid (PFNA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanoic acid (PFDA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluoroundecanoic acid (PFUnDA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorododecanoic acid (PFDoDA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotridecanoic acid (PFTrDA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotetradecanoic acid (PFTeDA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
Perfluorooctane sulfonamide (FOSA)	S26-Jl0045748	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S26-Jl0045748	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S26-Jl0045748	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S26-Jl0045748	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S26-Jl0045748	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S26-Jl0045748	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S26-Jl0045748	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1	Result 2	RPD			
Perfluorobutanesulfonic acid (PFBS)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorononanesulfonic acid (PFNS)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorooctanesulfonic acid (PFOS)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorodecanesulfonic acid (PFDS)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1	Result 2	RPD			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S26-Jl0045748	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S26-Jl0045748	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Comments

Sample Integrity

Custody Seals Intact (if used)	N/A
Attempt to Chill was evident	Yes
Sample correctly preserved	Yes
Appropriate sample containers have been used	Yes
Sample containers for volatile analysis received with minimal headspace	Yes
Samples received within HoldingTime	Yes
Some samples have been subcontracted	No

Qualifier Codes/Comments

Code	Description
J+	If the percent recoveries were greater than the upper control limit, qualify detects for the analyte in associated samples as estimated with a positive bias J+.
N09	Quantification of linear and branched isomers has been conducted as a single total response using the relative response factor for the corresponding linear/branched standard.
N11	Isotope dilution is used for calibration of each native compound for which an exact labelled analogue is available (Isotope Dilution Quantitation). The isotopically labelled analogues allow identification and recovery correction of the concentration of the associated native PFAS compounds.
N15	Where the native PFAS compound does not have labelled analogue then the quantification is made using the Extracted Internal Standard Analyte with the closest retention time to the analyte and no recovery correction has been made (Internal Standard Quantitation).
Q15	The RPD reported passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of this report.

Authorised by:

Nileshni Goundar	Analytical Services Manager
Caitlin Breeze	Senior Analyst-Inorganic



Glenn Jackson
Managing Director

Final Report – this report replaces any previously issued Report

- Indicates Not Requested

* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here.

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email: EnviroSales@eurofinsanz.com

Eurofins Environment Testing Australia Pty Ltd
ABN: 50 005 085 521

Melbourne	Geelong	Sydney	Newcastle	Canberra	Brisbane	Hobart
6 Monterey Road	19/8 Lewalan Street	179 Magowar Road	1/2 Frost Drive	Unit 1,2 Dacre Street	7 Smallwood Place	282A Argyle Street
Dandenong South	Grovedale	Girraween	Mayfield West	Mitchell	Murarrie	North Hobart
VIC 3175	VIC 3216	NSW 2145	NSW 2304	ACT 2911	QLD 4172	TAS 7000
+61 3 8564 5000	+61 3 8564 5000	+61 2 9900 8400	+61 2 4968 8448	+61 2 6113 8091	+61 7 3902 4600	+61 3 8564 5000
NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261
Site# 1254	Site# 25403	Site# 18217	Site# 25079	Site# 25466	Site# 20794 & 2780	Site# 26223

Eurofins ARL Pty Ltd
ABN: 91 05 0159 898

Perth
46-48 Banksia Road
Welshpool
WA 6106
+61 8 6253 4444
NATA# 2377
Site# 2370 & 2554

Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: WSU Surface Water Sampling
Project ID: 2301034J

Order No.:
Report #: 1384514
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Jul 16, 2026 1:42 PM
Due: Jul 23, 2026
Priority: 5 Day
Contact Name: Zahra Ginwala

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail						CANCELLED*	Total Suspended Solids Dried at 103°C to 105°C	Per- and Polyfluoroalkyl Substances (PFASs)
Sydney Laboratory - NATA # 1261 Site # 18217						X	X	X
External Laboratory								
No	**Sample ID**	**Sample Date**	**Sampling Time**	**Matrix**	**LAB ID**			
1	SW01	Jul 15, 2026		Water	S26-Jl0045110	X		
2	SW02	Jul 15, 2026		Water	S26-Jl0045111	X		
3	SW03	Jul 15, 2026		Water	S26-Jl0045112		X	X
4	SW04	Jul 15, 2026		Water	S26-Jl0045113		X	X
5	SW05	Jul 15, 2026		Water	S26-Jl0045114		X	X
6	SW06	Jul 15, 2026		Water	S26-Jl0045115		X	X
7	SW07	Jul 15, 2026		Water	S26-Jl0045116		X	X
8	SW08	Jul 15, 2026		Water	S26-Jl0045117		X	X
9	SW09	Jul 15, 2026		Water	S26-Jl0045118		X	X
10	SW10	Jul 15, 2026		Water	S26-Jl0045119		X	X
11	DW3	Jul 15, 2026		Water	S26-Jl0045120		X	X
12	DW4	Jul 15, 2026		Water	S26-Jl0045121		X	X
13	DUP1	Jul 15, 2026		Water	S26-Jl0045122			X
14	BLANK	Jul 15, 2026		Water	S26-Jl0045123			X

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Eurofins Environment Testing Australia Pty Ltd
ABN: 50 005 085 521

Melbourne	Geelong	Sydney	Newcastle	Canberra	Brisbane	Hobart
6 Monterey Road	19/8 Lewalan Street	179 Magowar Road	1/2 Frost Drive	Unit 1,2 Dacre Street	7 Smallwood Place	282A Argyle Street
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VIC 3175	VIC 3216	NSW 2145	NSW 2304	ACT 2911	QLD 4172	TAS 7000
+61 3 8564 5000	+61 3 8564 5000	+61 2 9900 8400	+61 2 4968 8448	+61 2 6113 8091	+61 7 3902 4600	+61 3 8564 5000
NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261	NATA# 1261
Site# 1254	Site# 25403	Site# 18217	Site# 25079	Site# 25466	Site# 20794 & 2780	Site# 26223

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ABN: 91 05 0159 898

Perth
46-48 Banksia Road
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WA 6106
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NATA# 2377
Site# 2370 & 2554

Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: WSU Surface Water Sampling
Project ID: 2301034J

Order No.:
Report #: 1384514
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Jul 16, 2026 1:42 PM
Due: Jul 23, 2026
Priority: 5 Day
Contact Name: Zahra Ginwala

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail	CANCELLED*	Total Suspended Solids Dried at 103°C to 105°C	Per- and Polyfluoroalkyl Substances (PFASs)
Sydney Laboratory - NATA # 1261 Site # 18217	X	X	X
Test Counts	2	10	12

Geo-Logix Pty Ltd
Building Q2, Level 3
2309/4 Daydream St
Warriewood, NSW 2102
ABN: 86 116 892 936
P: (02) 9979 1722
F: (02) 9979 1222

CHAIN OF CUSTODY

Project Manager: Zahra Ginwala
Contact email: zginwala@geo-logix.com.au & pdelprete@ge0-logix.com.au
Project Number: 2301034j
Project Name: WSU Surface Water Sampling
Date Submitted: 16/07/206

Purchase Order No: PO7250
Quote Reference:
Send Invoice to: accounts@geo-logix.com.au

TAT required: STD

ANALYSIS REQUIRED

Sample ID	Date	Matrix: soil	water	air	paint, filters	other	Con..	COMPOSITE	TCLP	TRH - C6 - C10	TRH - C10 - C40	BTEXN	VOCs	SVOCs	PAHs	OCPs	PCBs	Phenols	PFAS (80 comp)	Metals - M8	Metals - Specify **	Asbestos (WA DOH)	Asbestos (ID only)	Foreign Materials	S_{CR} ASS Suite	CEC	pH (KCl extract)	Clay Content (%)	Grain Size (by sieve)	Soil Agressivity (L2)	TSS						Hold	SUITE(S)
SW01	15.7.26		X				PFAS + Green												X												X						
SW02	"		X				"												X												X						
SW03	"		X				"												X												X						
SW04	"		X				"												X												X						
SW05	"		X				"												X												X						
SW06	"		X				"												X												X						
SW07	"		X				"												X												X						
SW08	"		X				"												X												X						
SW09	"		X				"												X												X						
SW10	"		X				"												X												X						
DW3	"		X				"												X												X						
DW4	"		X				"												X												X						
Dup 1	"		X				"												X												X						
Blank	"		X				PFAS bottle												X																		
Dup 1	"		X				Green bottle																													X	

Metals(circle):** As, Cd, Cr, Cu, Ni, Pb, Zn, Hg, Cr^{6+}, Cr^{3+}, Fe^{2+}, Fe^{3+}, Be, B, Al, V, Mn, Fe, Co, Se, Sr, Sn, Mo, Ag, Ba, Tl, Bi, Sb

Eurofins MGT Suite Codes

B1	TRH/BTEXN
B1A	TRH/MAH
B2	TRH/BTEXN/Pb
B2A	TRH/MAH/Pb
B3	PAH/Phenols
B4	TRH/BTEXN/PAH
B4A	TRH/BTEXN/PAH/Phenols
B5	TRH/BTEXN/M7
B6	TRH/BTEXN/M8
B7	TRH/BTEXN/PAH/M8
B7A	TRH/BTEXN/PAH/Phenols/M8
B8	TRH/VOC/PAH/M8
B9	TRH/BTEXN/PAH/OCP/M8
B10	TRH/BTEXN/PAH/OCP/OPP/M8
B11	Na/K/Ca/Mg/Cl/SO_4/CO_3/HCO_3/NH_3/NO_3
B11A	B11/Alkalinity
B11B	B11/EC/TDS
B12	TRH/BTEXN/Oxygenates/Ethanol
B12A	TRH/BTEXN/Oxygenates
B13	OCP/PCB
B14	OCP/OPP
B15	OCP/OPP/PCB
B16	TDS/SO_4/CH_4/Alk/BOD/COD/HPC/CUB
B17	SO_4/NO_3/Fe++/HPC/CUB
B18	Cl-/SO_4/pH
B19	N/P/K
B20	CEC/%ESP/Ca/Ma/Na/K

Chain of Custody

Relinquished by: [signature] Date/Time: 12:36 Signature: [signature] Received by: Jessica Date/Time: 16/7/26 Signature: [signature] 1:42pm 9.8 CH

#1384514

Version: V1
Issued: June 2015
Review: January 2022

Outlook

RE: Eurofins Sample Receipt Advice - Report 1384514 : Site WSU Surface Water Sampling (2301034J)

From Zahra Ginwala <ZGinwala@geo-logix.com.au>
Date Thu 16/07/2026 3:34 PM
To SH_AU25_Enviro_Sample_NSW <EnviroSampleNSW@eurofinsanz.com>

Unverified Sender: The sender of this email has not been verified. Review the content of the message carefully and verify the identity of the sender before acting on this email: replying, opening attachments or clicking links.

Hi Gabrielle,

That was a mistake – thank you for confirming.

Kind regards,

Zahra Ginwala | Environmental Geologist
NSW & QLD
Unit 2309/4 Daydream St, Warriewood NSW 2102
T: 02 9979 1722 | www.geo-logix.com.au

Geo-Logix
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From: Gabrielle Miler <EET-ELVIS@eurofinsanz.com>
Sent: Thursday, 16 July 2026 3:33 PM
To: Zahra Ginwala <ZGinwala@geo-logix.com.au>
Cc: Paula Del Prete <pdelprete@geo-logix.com.au>
Subject: Eurofins Sample Receipt Advice - Report 1384514 : Site WSU Surface Water Sampling (2301034J)

Dear Valued Client,

Please note:
Samples SW01 & SW02 not received, all analysis cancelled.

Please find attached a Sample Receipt Advice (SRA), a Summary Sheet and a scanned copy of your Chain-of-Custody (COC). It is important that you check this documentation to ensure that the details are correct such as the Client Job Number, Turn Around Time, any comments in the Notes section and sample numbers as well as the requested analysis. If there are any irregularities then please contact your Eurofins Analytical Services Manager as soon as possible to make certain that they get changed.

Kind regards
Gabrielle Miler
Sample Receipt Officer

Eurofins | Environment Testing
179 Magowar Road,

Girraween, NSW, 2145
Australia
Phone: +612 9900 8421
Email: EnviroSampleNSW@eurofins.com

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NATA Accredited
Accreditation Number 1261
Site Number 1254

Accredited for compliance with ISO/IEC 17025 – Testing
NATA is a signatory to the ILAC Mutual Recognition Arrangement for the mutual recognition of the equivalence of testing, medical testing, calibration, inspection, proficiency testing scheme providers and reference materials producers reports and certificates.

Attention: **Zahra Ginwala**

Report **1384515-W**
Project name WSU SURFACE WATER SAMPLING
Project ID 2301034j
Received Date Jul 16, 2026

Client Sample ID			**TRIP 1**
Sample Matrix			**Water**
Eurofins Sample No.			**S26-JI0045125**
Date Sampled			**Jul 15, 2026**
Test/Reference	LOR	Unit	
Perfluoroalkyl carboxylic acids (PFCAs)			
Perfluorobutanoic acid (PFBA)[N11]	0.05	ug/L	< 0.05
Perfluoropentanoic acid (PFPeA)[N11]	0.01	ug/L	0.02
Perfluorohexanoic acid (PFHxA)[N11]	0.01	ug/L	0.02
Perfluoroheptanoic acid (PFHpA)[N11]	0.01	ug/L	< 0.01
Perfluorooctanoic acid (PFOA)[N11]	0.01	ug/L	< 0.01
Perfluorononanoic acid (PFNA)[N11]	0.01	ug/L	< 0.01
Perfluorodecanoic acid (PFDA)[N11]	0.01	ug/L	< 0.01
Perfluoroundecanoic acid (PFUnDA)[N11]	0.01	ug/L	< 0.01
Perfluorododecanoic acid (PFDoDA)[N11]	0.01	ug/L	< 0.01
Perfluorotridecanoic acid (PFTrDA)[N15]	0.01	ug/L	< 0.01
Perfluorotetradecanoic acid (PFTeDA)[N11]	0.01	ug/L	< 0.01
13C4-PFBA (surr.)	1	%	86
13C5-PFPeA (surr.)	1	%	80
13C5-PFHxA (surr.)	1	%	89
13C4-PFHpA (surr.)	1	%	102
13C8-PFOA (surr.)	1	%	112
13C5-PFNA (surr.)	1	%	109
13C6-PFDA (surr.)	1	%	91
13C2-PFUnDA (surr.)	1	%	69
13C2-PFDoDA (surr.)	1	%	[J-]44
13C2-PFTeDA (surr.)	1	%	[J-]12
Perfluoroalkyl sulfonamido substances			
Perfluorooctane sulfonamide (FOSA)[N11]	0.05	ug/L	< 0.05
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)[N11]	0.05	ug/L	< 0.05
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)[N11]	0.05	ug/L	< 0.05
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)[N11]	0.05	ug/L	< 0.05
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)[N11]	0.05	ug/L	< 0.05
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)[N11]	0.05	ug/L	< 0.05
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)[N11]	0.05	ug/L	< 0.05
13C8-FOSA (surr.)	1	%	66
D3-N-MeFOSA (surr.)	1	%	[J-]11
D5-N-EtFOSA (surr.)	1	%	[J-]11

Client Sample ID			TRIP 1
Sample Matrix			Water
Eurofins Sample No.			S26-JI0045125
Date Sampled			Jul 15, 2026
Test/Reference	LOR	Unit	
Perfluoroalkyl sulfonamido substances			
D7-N-MeFOSE (surr.)	1	%	J-25
D9-N-EtFOSE (surr.)	1	%	J-21
D5-N-EtFOSAA (surr.)	1	%	55
D3-N-MeFOSAA (surr.)	1	%	64
Perfluoroalkyl sulfonic acids (PFSAs)			
Perfluorobutanesulfonic acid (PFBS)N11	0.01	ug/L	< 0.01
Perfluorononanesulfonic acid (PFNS)N15	0.01	ug/L	< 0.01
Perfluoropropanesulfonic acid (PFPrS)N15	0.01	ug/L	< 0.01
Perfluoropentanesulfonic acid (PFPeS)N15	0.01	ug/L	< 0.01
Perfluorohexanesulfonic acid (PFHxS)N11	0.01	ug/L	N090.02
Perfluoroheptanesulfonic acid (PFHpS)N15	0.01	ug/L	< 0.01
Perfluorooctanesulfonic acid (PFOS)N11	0.01	ug/L	N090.02
Perfluorodecanesulfonic acid (PFDS)N15	0.01	ug/L	< 0.01
13C3-PFBS (surr.)	1	%	92
18O2-PFHxS (surr.)	1	%	119
13C8-PFOS (surr.)	1	%	102
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)N11	0.01	ug/L	< 0.01
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)N11	0.05	ug/L	< 0.05
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)N11	0.01	ug/L	< 0.01
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)N11	0.01	ug/L	< 0.01
13C2-4:2 FTSA (surr.)	1	%	128
13C2-6:2 FTSA (surr.)	1	%	118
13C2-8:2 FTSA (surr.)	1	%	95
13C2-10:2 FTSA (surr.)	1	%	J-39
PFASs Summations			
Sum (PFHxS + PFOS)*	0.01	ug/L	0.04
Sum of WA DWER PFAS (n=10)*	0.05	ug/L	0.08
Sum of PFASs (n=30)*	0.1	ug/L	< 0.1

Sample History

Where samples are submitted/analysed over several days, the last date of extraction is reported.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description	Testing Site	Extracted	Holding Time
Per- and Polyfluoroalkyl Substances (PFASs)			
Perfluoroalkyl carboxylic acids (PFCAs)	Melbourne	Jul 17, 2026	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonamido substances	Melbourne	Jul 17, 2026	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
Perfluoroalkyl sulfonic acids (PFSAs)	Melbourne	Jul 17, 2026	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)	Melbourne	Jul 17, 2026	28 Days
- Method: LTM-ORG-2100 Per- and Polyfluoroalkyl Substances (PFAS)			

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Melbourne	Geelong	Sydney	Newcastle	Canberra	Brisbane	Hobart
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Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: WSU SURFACE WATER SAMPLING
Project ID: 2301034j

Order No.: PO7251
Report #: 1384515
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Jul 16, 2026 1:42 PM
Due: Jul 23, 2026
Priority: 5 Day
Contact Name: Zahra Ginwala

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail						HOLD*	Per- and Polyfluoroalkyl Substances (PFASs)
Melbourne Laboratory - NATA # 1261 Site # 1254						X	X
External Laboratory							
No	**Sample ID**	**Sample Date**	**Sampling Time**	**Matrix**	**LAB ID**		
1	TRIP 1	Jul 15, 2026		Water	S26-Jl0045125		X
2	TRIP 1	Jul 15, 2026		Water	S26-Jl0045126	X	
Test Counts						1	1

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Internal Quality Control Review and Glossary

General

1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follow guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013. They are included in this QC report where applicable. Additional QC data may be available on request.
2. Unless otherwise stated, all soil/sediment/solid results are reported on a dry weight basis.
3. Unless otherwise stated, all biota/food results are reported on a wet weight basis on the edible portion.
4. For CEC results where the sample's origin is unknown or environmentally contaminated, the results should be used advisedly.
5. Actual LORs are matrix dependent. Quoted LORs may be raised where sample extracts are diluted due to interferences.
6. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds where annotated.
7. SVOC analysis on waters is performed on homogenised, unfiltered samples unless noted otherwise.
8. Samples were analysed on an 'as received' basis.
9. Information identified in this report with **blue** colour indicates data provided by customers that may have an impact on the results.
10. This report replaces any interim results previously issued.

Holding Times

Please refer to the 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours before sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and despite any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the sampling date; therefore, compliance with these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether, the holding time is seven days; however, for all other VOCs, such as BTEX or C6-10 TRH, the holding time is 14 days.

Units

mg/kg: milligrams per kilogram	**mg/L:** milligrams per litre	**ppm:** parts per million
µg/L: micrograms per litre	**ppb:** parts per billion	**%:** Percentage
org/100 mL: Organisms per 100 millilitres	**NTU:** Nephelometric Turbidity Units	**MPN/100 mL:** Most Probable Number of organisms per 100 millilitres
CFU: Colony Forming Unit	**Colour:** Pt-Co Units (CU)	

Terms

APHA	American Public Health Association
CEC	Cation Exchange Capacity
COC	Chain of Custody
CP	Client Parent - QC was performed on samples pertaining to this report
CRM	Certified Reference Material (ISO17034) - reported as percent recovery.
Dry	Where moisture has been determined on a solid sample, the result is expressed on a dry weight basis.
Duplicate	A second piece of analysis from the same sample and reported in the same units as the result to show comparison.
LOR	Limit of Reporting.
LCS	Laboratory Control Sample - reported as percent recovery.
Method Blank	In the case of solid samples, these are performed on laboratory-certified clean sands and in the case of water samples, these are performed on de-ionised water.
NCP	Non-Client Parent - QC performed on samples not pertaining to this report, QC represents the sequence or batch that client samples were analysed within.
RPD	Relative Percent Difference between two Duplicate pieces of analysis.
SPIKE	Addition of the analyte to the sample and reported as percentage recovery.
SRA	Sample Receipt Advice
Surr - Surrogate	The addition of a similar compound to the analyte target is reported as percentage recovery. See below for acceptance criteria.
TBTO	Tributyltin oxide (*bis*-tributyltin oxide) - individual tributyltin compounds cannot be identified separately in the environment; however, free tributyltin was measured, and its values were converted stoichiometrically into tributyltin oxide for comparison with regulatory limits.
TCLP	Toxicity Characteristic Leaching Procedure
TEQ	Toxic Equivalency Quotient or Total Equivalence
QSM	US Department of Defense Quality Systems Manual Version 6.0
US EPA	United States Environmental Protection Agency
WA DWER	Sum of PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

QC - Acceptance Criteria

The acceptance criteria should only be used as a guide and may be different when site-specific Sampling Analysis and Quality Plan (SAQP) have been implemented.

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is ≤30%; however, the following acceptance guidelines are equally applicable:

Results <10 times the LOR:	No Limit
Results between 10-20 times the LOR:	RPD must lie between 0-50%
Results >20 times the LOR:	RPD must lie between 0-30%

NOTE: pH duplicates are reported as a range, not as RPD

Surrogate Recoveries: Recoveries must lie between 20-130% for Speciated Phenols & 50-150% for PFAS. SVOCs recoveries 20 – 150%, VOC recoveries 50 – 150%

PFAS field samples containing surrogate recoveries above the QC limit designated in QSM 6.0, where no positive PFAS results have been reported or reviewed, and no data was affected.

QC Data General Comments

1. Where a result is reported as less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown are not data from your samples.
3. pH and Free Chlorine analysed in the laboratory - Analysis on this test must begin within 30 minutes of sampling. Therefore, laboratory analysis is unlikely to be completed within holding time.Analysis will begin as soon as possible after sample receipt.
4. Recovery Data (Spikes & Surrogates) - where chromatographic interference does not allow the determination of recovery, the term "INT" appears against that analyte.
5. For Matrix Spikes and LCS results, a dash "-" in the report means that the specific analyte was not added to the QC sample.
6. Duplicate RPDs are calculated from raw analytical data; thus, it is possible to have two sets of data.

Quality Control Results

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Method Blank							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	ug/L	< 0.05			0.05	Pass	
Perfluoropentanoic acid (PFPeA)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanoic acid (PFHxA)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanoic acid (PFHpA)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanoic acid (PFOA)	ug/L	< 0.01			0.01	Pass	
Perfluorononanoic acid (PFNA)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanoic acid (PFDA)	ug/L	< 0.01			0.01	Pass	
Perfluoroundecanoic acid (PFUnDA)	ug/L	< 0.01			0.01	Pass	
Perfluorododecanoic acid (PFDoDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotridecanoic acid (PFTrDA)	ug/L	< 0.01			0.01	Pass	
Perfluorotetradecanoic acid (PFTeDA)	ug/L	< 0.01			0.01	Pass	
Method Blank							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	ug/L	< 0.05			0.05	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	ug/L	< 0.05			0.05	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	ug/L	< 0.05			0.05	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	ug/L	< 0.05			0.05	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	ug/L	< 0.05			0.05	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	ug/L	< 0.05			0.05	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	ug/L	< 0.05			0.05	Pass	
Method Blank							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	ug/L	< 0.01			0.01	Pass	
Perfluorononanesulfonic acid (PFNS)	ug/L	< 0.01			0.01	Pass	
Perfluoropropanesulfonic acid (PFPrS)	ug/L	< 0.01			0.01	Pass	
Perfluoropentanesulfonic acid (PFPeS)	ug/L	< 0.01			0.01	Pass	
Perfluorohexanesulfonic acid (PFHxS)	ug/L	< 0.01			0.01	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	ug/L	< 0.01			0.01	Pass	
Perfluorooctanesulfonic acid (PFOS)	ug/L	< 0.01			0.01	Pass	
Perfluorodecanesulfonic acid (PFDS)	ug/L	< 0.01			0.01	Pass	
Method Blank							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	ug/L	< 0.05			0.05	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	ug/L	< 0.01			0.01	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	ug/L	< 0.01			0.01	Pass	
LCS - % Recovery							
Perfluoroalkyl carboxylic acids (PFCAs)							
Perfluorobutanoic acid (PFBA)	%	100			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	%	91			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	%	105			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	%	87			50-150	Pass	
Perfluorooctanoic acid (PFOA)	%	101			50-150	Pass	
Perfluorononanoic acid (PFNA)	%	92			50-150	Pass	
Perfluorodecanoic acid (PFDA)	%	98			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	%	101			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	%	110			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	%	81			50-150	Pass	
Perfluorotetradecanoic acid (PFTeDA)	%	73			50-150	Pass	

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
LCS - % Recovery							
Perfluoroalkyl sulfonamido substances							
Perfluorooctane sulfonamide (FOSA)	%	100			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	%	103			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	%	100			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	%	98			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	%	97			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	%	95			50-150	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	%	100			50-150	Pass	
LCS - % Recovery							
Perfluoroalkyl sulfonic acids (PFSAs)							
Perfluorobutanesulfonic acid (PFBS)	%	92			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	%	81			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	%	90			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	%	94			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	%	87			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	%	96			50-150	Pass	
Perfluorooctanesulfonic acid (PFOS)	%	85			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	%	64			50-150	Pass	
LCS - % Recovery							
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)							
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	%	77			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	%	87			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	%	88			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	%	94			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Spike - % Recovery									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1					
Perfluorobutanoic acid (PFBA)	S26-Jl0045125	CP	%	109			50-150	Pass	
Perfluoropentanoic acid (PFPeA)	S26-Jl0045125	CP	%	103			50-150	Pass	
Perfluorohexanoic acid (PFHxA)	S26-Jl0045125	CP	%	112			50-150	Pass	
Perfluoroheptanoic acid (PFHpA)	S26-Jl0045125	CP	%	108			50-150	Pass	
Perfluorooctanoic acid (PFOA)	S26-Jl0045125	CP	%	113			50-150	Pass	
Perfluorononanoic acid (PFNA)	S26-Jl0045125	CP	%	110			50-150	Pass	
Perfluorodecanoic acid (PFDA)	S26-Jl0045125	CP	%	109			50-150	Pass	
Perfluoroundecanoic acid (PFUnDA)	S26-Jl0045125	CP	%	125			50-150	Pass	
Perfluorododecanoic acid (PFDoDA)	S26-Jl0045125	CP	%	127			50-150	Pass	
Perfluorotridecanoic acid (PFTrDA)	S26-Jl0045125	CP	%	77			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonamido substances				Result 1					
Perfluorooctane sulfonamide (FOSA)	S26-Jl0045125	CP	%	107			50-150	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	S26-Jl0045125	CP	%	113			50-150	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	S26-Jl0045125	CP	%	108			50-150	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	S26-Jl0045125	CP	%	109			50-150	Pass	
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	S26-Jl0045125	CP	%	118			50-150	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	S26-Jl0045125	CP	%	123			50-150	Pass	

Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	S26-Jl0045125	CP	%	108			50-150	Pass	
Spike - % Recovery									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1					
Perfluorobutanesulfonic acid (PFBS)	S26-Jl0045125	CP	%	100			50-150	Pass	
Perfluorononanesulfonic acid (PFNS)	S26-Jl0045125	CP	%	87			50-150	Pass	
Perfluoropropanesulfonic acid (PFPrS)	S26-Jl0045125	CP	%	95			50-150	Pass	
Perfluoropentanesulfonic acid (PFPeS)	S26-Jl0045125	CP	%	102			50-150	Pass	
Perfluorohexanesulfonic acid (PFHxS)	S26-Jl0045125	CP	%	91			50-150	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	S26-Jl0045125	CP	%	107			50-150	Pass	
Perfluorodecanesulfonic acid (PFDS)	S26-Jl0045125	CP	%	64			50-150	Pass	
Spike - % Recovery									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1					
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	S26-Jl0045125	CP	%	97			50-150	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	S26-Jl0045125	CP	%	106			50-150	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	S26-Jl0045125	CP	%	107			50-150	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	S26-Jl0045125	CP	%	138			50-150	Pass	
Test	**Lab Sample ID**	**QA Source**	**Units**	**Result 1**			**Acceptance Limits**	**Pass Limits**	**Qualifying Code**
Duplicate									
Perfluoroalkyl carboxylic acids (PFCAs)				Result 1	Result 2	RPD			
Perfluorobutanoic acid (PFBA)	M26-Jl0047482	NCP	ug/L	0.57	0.56	2.0	30%	Pass	
Perfluoropentanoic acid (PFPeA)	M26-Jl0047482	NCP	ug/L	1.8	1.8	3.0	30%	Pass	
Perfluorohexanoic acid (PFHxA)	M26-Jl0047482	NCP	ug/L	1.5	1.4	3.0	30%	Pass	
Perfluoroheptanoic acid (PFHpA)	M26-Jl0047482	NCP	ug/L	0.36	0.39	7.0	30%	Pass	
Perfluorooctanoic acid (PFOA)	M26-Jl0047482	NCP	ug/L	1.3	1.3	2.0	30%	Pass	
Perfluorononanoic acid (PFNA)	M26-Jl0047482	NCP	ug/L	0.05	0.05	4.0	30%	Pass	
Perfluorodecanoic acid (PFDA)	M26-Jl0047482	NCP	ug/L	0.05	0.06	8.0	30%	Pass	
Perfluoroundecanoic acid (PFUnDA)	M26-Jl0047482	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorododecanoic acid (PFDoDA)	M26-Jl0047482	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotridecanoic acid (PFTrDA)	M26-Jl0047482	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluorotetradecanoic acid (PFTeDA)	M26-Jl0047482	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
Perfluorooctane sulfonamide (FOSA)	M26-Jl0047482	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-methylperfluoro-1-octane sulfonamide (N-MeFOSA)	M26-Jl0047482	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethylperfluoro-1-octane sulfonamide (N-EtFOSA)	M26-Jl0047482	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
2-(N-methylperfluoro-1-octane sulfonamido)-ethanol(N-MeFOSE)	M26-Jl0047482	NCP	ug/L	0.06	0.07	6.0	30%	Pass	

Duplicate									
Perfluoroalkyl sulfonamido substances				Result 1	Result 2	RPD			
2-(N-ethylperfluoro-1-octane sulfonamido)-ethanol(N-EtFOSE)	M26-JI0047482	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
N-ethyl-perfluorooctanesulfonamidoacetic acid (N-EtFOSAA)	M26-JI0047482	NCP	ug/L	0.16	0.17	9.0	30%	Pass	
N-methyl-perfluorooctanesulfonamidoacetic acid (N-MeFOSAA)	M26-JI0047482	NCP	ug/L	0.11	0.11	3.0	30%	Pass	
Duplicate									
Perfluoroalkyl sulfonic acids (PFSAs)				Result 1	Result 2	RPD			
Perfluorobutanesulfonic acid (PFBS)	M26-JI0047482	NCP	ug/L	1.0	0.95	9.0	30%	Pass	
Perfluorononanesulfonic acid (PFNS)	M26-JI0047482	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
Perfluoropropanesulfonic acid (PFPrS)	M26-JI0047482	NCP	ug/L	0.03	0.03	3.0	30%	Pass	
Perfluoropentanesulfonic acid (PFPeS)	M26-JI0047482	NCP	ug/L	0.06	0.07	20	30%	Pass	
Perfluorohexanesulfonic acid (PFHxS)	M26-JI0047482	NCP	ug/L	0.29	0.28	4.0	30%	Pass	
Perfluoroheptanesulfonic acid (PFHpS)	M26-JI0047482	NCP	ug/L	0.03	0.03	1.0	30%	Pass	
Perfluorooctanesulfonic acid (PFOS)	M26-JI0047482	NCP	ug/L	0.51	0.54	6.0	30%	Pass	
Perfluorodecanesulfonic acid (PFDS)	M26-JI0047482	NCP	ug/L	0.02	0.02	23	30%	Pass	
Duplicate									
n:2 Fluorotelomer sulfonic acids (n:2 FTSAs)				Result 1	Result 2	RPD			
1H.1H.2H.2H-perfluorohexanesulfonic acid (4:2 FTSA)	M26-JI0047482	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorooctanesulfonic acid(6:2 FTSA)	M26-JI0047482	NCP	ug/L	< 0.05	< 0.05	<1	30%	Pass	
1H.1H.2H.2H-perfluorodecanesulfonic acid (8:2 FTSA)	M26-JI0047482	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	
1H.1H.2H.2H-perfluorododecanesulfonic acid (10:2 FTSA)	M26-JI0047482	NCP	ug/L	< 0.01	< 0.01	<1	30%	Pass	

Comments

Sample Integrity

Custody Seals Intact (if used)	N/A
Attempt to Chill was evident	Yes
Sample correctly preserved	Yes
Appropriate sample containers have been used	Yes
Sample containers for volatile analysis received with minimal headspace	Yes
Samples received within HoldingTime	Yes
Some samples have been subcontracted	No

Qualifier Codes/Comments

Code	Description
J-	If the percent recoveries were less than the lower control limit, qualify detects for the analyte in associated samples as estimated with a negative bias J-.
N09	Quantification of linear and branched isomers has been conducted as a single total response using the relative response factor for the corresponding linear/branched standard.
N11	Isotope dilution is used for calibration of each native compound for which an exact labelled analogue is available (Isotope Dilution Quantitation). The isotopically labelled analogues allow identification and recovery correction of the concentration of the associated native PFAS compounds.
N15	Where the native PFAS compound does not have labelled analogue then the quantification is made using the Extracted Internal Standard Analyte with the closest retention time to the analyte and no recovery correction has been made (Internal Standard Quantitation).

Authorised by:

Nileshni Goundar	Analytical Services Manager
Joseph Edouard	Senior Analyst-PFAS



Glenn Jackson
Managing Director

Final Report – this report replaces any previously issued Report

- Indicates Not Requested

* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please click here.

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Company Name: Geo-Logix P/L
Address: Bld Q2 Level 3, 2309/4 Daydream St
Warriewood
NSW 2102

Project Name: WSU SURFACE WATER SAMPLING
Project ID: 2301034j

Order No.: PO7251
Report #: 1384515
Phone: 02 9979 1722
Fax: 02 9979 1222

Received: Jul 16, 2026 1:42 PM
Due: Jul 23, 2026
Priority: 5 Day
Contact Name: Zahra Ginwala

Eurofins Analytical Services Manager : Bonnie Pu

Sample Detail						HOLD*	Per- and Polyfluoroalkyl Substances (PFASs)
Melbourne Laboratory - NATA # 1261 Site # 1254						X	X
External Laboratory							
No	**Sample ID**	**Sample Date**	**Sampling Time**	**Matrix**	**LAB ID**		
1	TRIP 1	Jul 15, 2026		Water	S26-Jl0045125		X
2	TRIP 1	Jul 15, 2026		Water	S26-Jl0045126	X	
Test Counts						1	1

Geo-Logix Pty Ltd
Building C2, Level 3
2309/4 Daydream St
Warriewood, NSW 2102
ABN: 86 116 892 936
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CHAIN OF CUSTODY

Project Manager: Zahra Ginwala
Contact email: zginwala@geo-logix.com.au & pdelprete@ge0-logix.com.au
Project Number: 2301034j
Project Name: WSU Surface Water Sampling
Date Submitted: 16/07/206

Purchase Order No: PO7251
Quote Reference:
Send Invoice to: accounts@geo-logix.com.au

TAT required: STD

ANALYSIS REQUIRED

Sample ID	Date	Matrix: soil	water	air	paint, filters	other	Comments	COMPOSITE	TCLP	TRH - C6 - C10	TRH - C10 - C40	BTEXN	VOCs	SVOCs	PAHs	OCPs	PCBs	Phenols	PFAS (30 comp)	Metals - M8	Metals - Specify **	Asbestos (WA DOH)	Asbestos (ID only)	Foreign Materials	S_{CR} ASS Suite	CEC	pH (KCl extract)	Clay Content (%)	Grain Size (by sieve)	Soil Agressivity (L2)	TSS					Hold	SUITE(S)
Trip 1	15.7.26		X				PFAS bottle												X																	
Trip 1	"		X				Green bottle																													X	

Eurofins MGT Suite Codes

B1	TRH/BTEXN
B1A	TRH/MAH
B2	TRH/BTEXN/Pb
B2A	TRH/MAH/Pb
B3	PAH/Phenols
B4	TRH/BTEXN/PAH
B4A	TRH/BTEXN/PAH/Phenols
B5	TRH/BTEXN/M7
B6	TRH/BTEXN/M8
B7	TRH/BTEXN/PAH/M8
B7A	TRH/BTEXN/PAH/Phenols/M8
B8	TRH/VOC/PAH/M8
B9	TRH/BTEXN/PAH/OCP/M8
B10	TRH/BTEXN/PAH/OCP/OPP/M8
B11	Na/K/Ca/Mg/Cl/SO_4/CO_3/HCO_3/NH_3/NO_3
B11A	B11/Alkalinity
B11B	B11/EC/TDS
B12	TRH/BTEXN/Oxygenates/Ethanol
B12A	TRH/BTEXN/Oxygenates
B13	OCP/PCB
B14	OCP/OPP
B15	OCP/OPP/PCB
B16	TDS/SO_4/CH_4/Alk/BOD/COD/HPC/CUB
B17	SO_4/NO_3/Fe++/HPC/CUB
B18	Cl-/SO_4/pH
B19	N/P/K
B20	CEC/%ESP/Ca/Ma/Na/K

Metals(circle):** As, Cd, Cr, Cu, Ni, Pb, Zn, Hg, Cr^{6+}, Cr^{3+}, Fe^{2+}, Fe^{3+}, Be, B, Al, V, Mn, Fe, Co, Se, Sr, Sn, Mo, Ag, Ba, Tl, Bi, Sb

Chain of Custody

Relinquished by: _____ Date/Time: _____ Signature: _____ Received by: Jessica Date/Time: 16/7/26 Signature: _____ 1:42pm

1384515 CH